

## ORIGINAL ARTICLE

# FREQUENCY AND ANTIMICROBIAL RESISTANCE PATTERN OF VIBRIO CHOLERAE IN STOOL SAMPLES

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## ABSTRACT

**Background:** Cholera, one of the most common cause of acute watery diarrhea in developing world, requires, along with correction of fluid and electrolyte imbalance, antibiotics in to shorten the duration of disease. Emergence of resistance with recommended antimicrobials is becoming an issue to treat *Vibrio Cholerae*. The purpose of this research was to determine the frequency and antimicrobial resistance pattern of *Vibrio cholera* isolated in stool samples at a tertiary care hospital.

**Methods:** Stool samples were received from both in-patients and out-patients in sterile leak proof containers. All *Vibrio Cholerae* isolated from stool samples were included in the study. The stool samples were inoculated on Thiosulphate-citrate-Bile salts-sucrose agar. Suspected *Vibrio cholerae* colonies were stained by Gram's stain Method. Oxidase positive colonies were isolated on Mueller Hinton agar (MHA). Slide agglutination tests were done to confirm presence of *Vibrio cholera* and its serotypes.

**Results:** A total of 352 (6.37%) Isolates of *Vibrio Cholerae* achieved from 5524 stool samples. *Vibrio Cholerae* serotype Ogawa isolated 188/352 (53.4%), while 164(46.6%) were *Vibrio cholerae* serotype Inaba. Predominant the isolates were from male patients 184/352 (52.3%), while from females were 168/352 (47.7%). Male to female ratio was 1.095:1. Mean age of patients with positive *Vibrio cholerae* isolates was  $27.15 \pm 25.61$  years. The Frequency of resistance with Co-trimoxazole, Tetracycline, Ampicillin, Chloramphenicol and Ciprofloxacin was 66.2%, 40.9%, 8.8%, 2.6% and 1.4% respectively.

**Conclusion:** The multiple antimicrobials recommended to treat Cholera have been found to develop alarming resistance among *Vibrio Cholerae* isolates. This situation needs a regular surveillance of commonly used antibiotics and urgent discovery of alternative antibiotics.

**KEYWORDS:** Frequency. Antimicrobial resistance pattern. *Vibrio cholerae*. Stool.

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## INTRODUCTION

Cholera, a serious community health issue in developing nations, has characteristic features of profuse watery diarrhea, sometimes with vomiting, causing dehydration. When not treated urgently it can be severe enough to lead to death. Along with rehydration and hygiene as part of management, it is important to control cholera with antibiotic treat-

ment especially with tetracycline.<sup>1</sup> An effective antibiotic helps in reduction of loss of fluids and period, intensity, transmission, morbidity and mortality of the disease.<sup>2,3</sup>

According to World health Organization (WHO), Cholera remains a leading community health issue in the world, particularly in Africa and Asia,<sup>4</sup> and if not diagnosed and treated properly it can be transmit-

ted to non-endemic regions.<sup>5</sup> Over 3.5 million population worldwide is affected annually, and between 100,000-130,000 deaths reported in last few years.<sup>6</sup> Notified cases of Cholera have been increased tremendously and most of the under developed nations are at risk of having outbreaks of Cholera epidemics.<sup>4</sup> Rehydration and replacement of electrolyte is the main treatment of Cholera. Antibiotic treatment is administered routinely to reduce severity and to shorten duration of illness and to decrease the chances of spread of infection. However, strains of *Vibrio cholera* (*V.cholerae*) showing resistance to antibiotics in different parts of world, as evidenced by studies.<sup>7-12</sup> Injudicious use of antibiotics is documented as a cause of development of resistance in cholera epidemics.<sup>13</sup> Prescribing inappropriate or wrong antibiotics can be dangerous and may have long term consequences.

Centers for disease control and prevention (CDC) has recommended antimicrobials from the group of tetracyclines, fluoroquinolones, macrolides and co-trimoxazole for the Cholera treatment in general,<sup>14</sup> yet local microbial susceptibility should be considered to choose the specific antibiotics.

In Pakistan, concerning susceptibility of antibiotic profile of *V.cholerae*, there is scanty information available. Consequently, there is no assurance of current antibiotic range in controlling and management of *V. cholerae*. In the current study, we reviewed and classified *V. cholerae* archived strains, determined frequently used antimicrobials resistance profile, and assessed patterns of resistance during last few years. This will help the physicians for selecting the appropriate antimicrobial management and their judicious use.

## METHODS

This observational study was conducted over a period of five years from January, 2011 to December, 2015 at the Department of Gastroenterology and the Department of Clinical Microbiology of Ziauddin University Hospital. Stool samples were received from both in-patients and out-patients in sterile leak proof containers. All *V. cholerae* isolated from stool samples were included in the study. All other stool bacterial pathogens were excluded from the study. Written approval from institutional

ethical committee was taken. Informed consent was taken from patient or any other patient's relative. The stool samples were inoculated on Thiosulphate-citrate-Bile salt-sucrose (TCBS) agar. Plates were incubated for 48 hours at 37°C aerobically. After 24 hour incubation plates were examined, yellow colonies suspected of *V. cholera* were stained by Gram's Method. Those colonies which stained as pink curved rods and were found to be oxidase positive were isolated on Mueller Hinton agar (MHA). The serological tests (slide agglutination test) were done to confirm the presence of *V. cholera* and its serotypes according to standard microbiological techniques.<sup>15</sup> The antisera used was from Denka Seiken co. Ltd containing specific antibodies against somatic antigen.

Antimicrobial susceptibility testing was performed on MHA medium (Oxoid Ltd., England) using modified Kirby Bauer's disk diffusion method according to Clinical and Laboratory Standards Institute (CLSI) guidelines.<sup>16</sup> *Escherichia coli* American Type Culture Collection (ATCC®) 25922 was used as control.

Data analysis was performed by using SPSS version-20. Frequency and percentages were computed for presentation of all categorical variables like micro-organisms, sex, and antimicrobial sensitivities. Mean and standard deviation was calculated for quantitative variables like age of patients.

## RESULTS

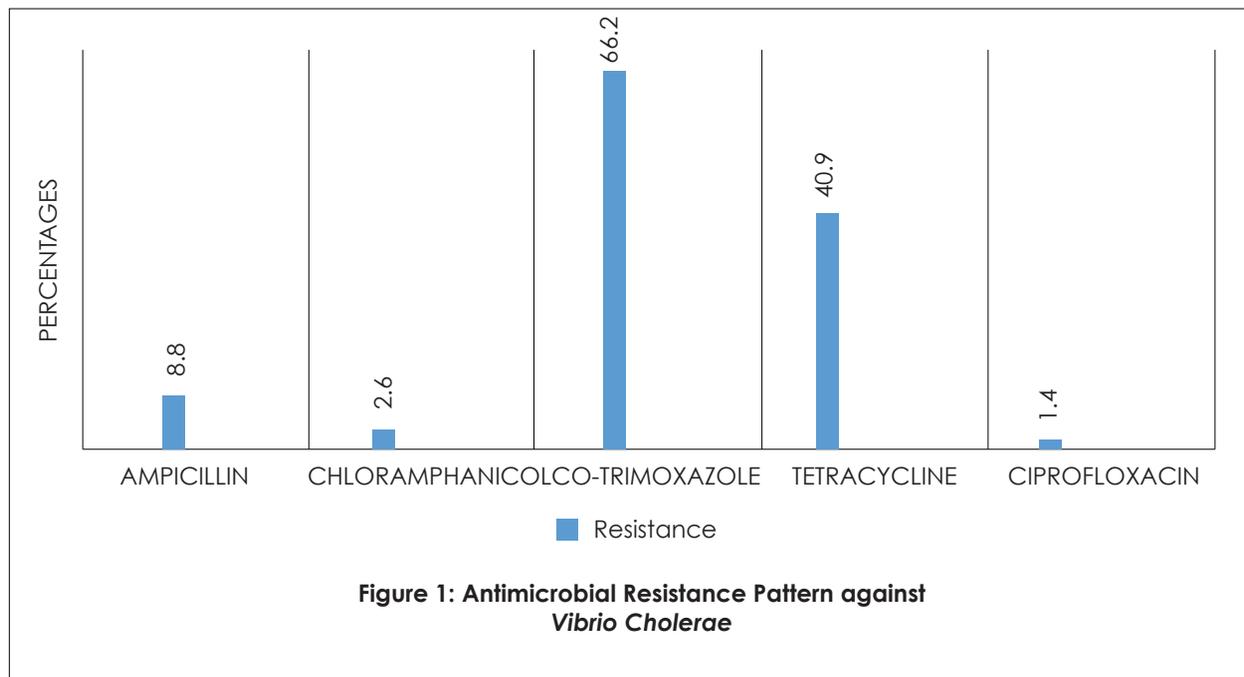
Stool samples of 5524 of in and out patients were processed for culture and antimicrobial susceptibilities during the study period. From these stool samples a total of 352 clinical isolates of *V. cholerae* were archived. The frequency of *V.cholerae* isolates were 352/5524 (6.37%). In those 352 isolates 188 (53.4%) were *V.cholerae* serotype Ogawa and 164(46.6%) were *V.cholerae* serotype Inaba (**Table 1**). Predominantly the isolates of *V.cholerae* were from male patients 184/352 (52.3%), while isolates from female patients were 168/352 (47.7%) (**Table 2**). Male to female ratio was 1.095:1. Mean age of patients with positive *V.cholerae* isolates was 27.15±25.61 years. Five common antibiotics were used to screen those strains. The Antimicrobial susceptibilities were shown in **Figure 1**.

**Table:1 Vibrio Cholera Serotype**

|       |                       | Frequency | Percent | Valid Percent | Cumulative Percent |
|-------|-----------------------|-----------|---------|---------------|--------------------|
| Valid | Vibrio Cholerae Ogawa | 188       | 53.4    | 53.4          | 53.4               |
|       | Vibrio Choleraenaba   | 164       | 46.6    | 46.6          | 100.0              |
|       | Total                 | 352       | 100.0   | 100.0         |                    |

**Table:2 Gender**

|       |        | Frequency | Percent | Valid Percent | Cumulative Percent |
|-------|--------|-----------|---------|---------------|--------------------|
| Valid | Male   | 184       | 52.3    | 52.3          | 52.3               |
|       | Female | 168       | 47.7    | 47.7          | 100.0              |
|       | Total  | 352       | 100.0   | 100.0         |                    |



## DISCUSSION

Cholera is an endemic disease in Pakistan.<sup>17</sup> Contamination of water and food, unhealthy environment and poor sanitation are the causes of outbreaks of Cholera.<sup>18</sup> Incidence of Cholera in Pakistan is estimated as 1.64/1000 annually.<sup>19</sup>

*V.cholerae* is a curved gram negative bacilli that comprises both pathogenic and non-pathogenic strains. It has three serotypes (Ogawa, Inaba and rarely Hikojima). Clinical manifestations and sign

and symptoms of Ogawa serotype are similar to Inaba serotype. Irrespective of serotype patients with concurrent infections and immunodeficiency are at high risk of having morbidity and mortality. Emergence of resistance with antimicrobials has remained a problem for years in Pakistan with changing pattern of sensitivity among Ampicillin, Chloramphanicol, Co-trimoxazole and Tetracycline.<sup>20</sup>

In studies from China the pattern of antimicrobial susceptibility was found 38.5% to Co-trimoxazole,

11% to tetracycline and 45.9% to nalidixic acid.<sup>12</sup> In India, strains during epidemics have shown an increasing pattern of resistance with nalidixic acid and Co-trimoxazole.<sup>21</sup> The antibiotic of choice does not only depend on the prevailing effectiveness of the antibiotic against *V.cholerae* isolate involved, but a number of factors such as age, sex, and physiological status.

In the current study of *V.cholerae* isolates, not only the pattern of resistance was observed but also the resistance with multiple antimicrobials among isolates of *V.cholerae* was studied. In this study the frequency of *V.cholerae* isolates was 6.37%. A recent study in Baghdad showed that 3.4% of stool samples were positive for *V.cholerae*.<sup>22</sup> A similar study in India showed isolation of *V.cholerae* in 24.8% of stool samples of pediatric patients presented with acute watery diarrhea.<sup>23</sup> In our study serotype Ogawa was isolated in 46.5% of stool samples while serotype Inaba was present in 53.5% of stool samples. It was in contrast to a study from India, Karnataka, where the burden of *V.cholerae* serotype Ogawa was 87.7% and *V.cholerae* serotype, Inaba, 8.3%.<sup>24</sup> Another study from India also showed frequency of *V.cholerae* serotype Ogawa and *V.cholerae* serotype Inaba as 69.5% and 30.5% respectively.<sup>25</sup>

Our study showed that 66.2% isolates were resistant to Co-trimoxazole, while pattern of resistance of *V.cholerae* with Tetracycline, Ampicillin, Chloramphenicol and Ciprofloxacin was 40.9 %, 8.8%, 2.6% and 1.4% respectively. A study from Iran also showed similar pattern of resistance of antimicrobials with *V.cholerae* strains in stool. This study exhibited the resistance with Co-trimoxazole, Tetracycline, Ampicillin and Ciprofloxacin as 43%, 29%, 27% and 0% respectively.<sup>26</sup> Chloramphenicol was not used in this study to treat *V.cholerae*. Similarly, a study from Pakistan, conducted in 2001-2, in Rawalpindi, showed a better trend of resistance of antimicrobials with *V.cholerae*. In this study *V.cholerae* showed resistance with Co-trimoxazole, Tetracycline, Ampicillin and Ciprofloxacin with a frequency of 95.8%, 0%, 4.17% and 0% respectively.<sup>27</sup> This situation is alarming for a developing country, as in the past, Tetracycline, Co-trimoxazole have been used as the drug of choice in patients with Cholera related diarrhea. This predicts that there should be a continuous surveillance of antibiotics used in the treatment of Cholera because of ever changing pattern of resistance of *V.cholerae*.

### CONCLUSION

The current study concludes the emergence of resistance with commonly used antibiotics to treat *V. cholerae*, including some of the antibiotics recommended by WHO for treatment in Pakistan. The frequency of resistance with antimicrobials is increasing with Co-Trimoxazole, Tetracycline and

ampicillin, which are the antibiotics of choice for Cholera. The situation is alarming, especially in a country where Cholera outbreaks are common especially after floods. However, Chloramphenicol and Ciprofloxacin are still highly effective for treatment of Cholera. To prevent further spread of resistance, these antibiotics should be used judiciously, and should be limited to those patients who are moderate to severely dehydrated due to Cholera<sup>28</sup>. The multi-drug resistance in *V. cholerae* demands the continuous surveillance of commonly used antimicrobials. Also, there is a need to search for alternative antimicrobials which can help in treating Cholera effectively.

### REFERENCES

1. Volk W, Benjamin d, Kadner R, & Parson T, Eds. Essentials of Medical Microbiology 4<sup>th</sup>edn. Philadelphia: J. B. Lippincott Company. 1991.
2. Rahaman MM, Majid MA, Alam AKMJ, Islam MR. Effects of doxycycline in actively purging cholera patients: a double-blind clinical trial. *Antimicrob Agents Chemother.* 1976; 10:610-2.
3. Sack DA, Sack RB, Nair GB, Siddique AK, Cholera. *Lancet* 2004; 363: 223-33.
4. WHO. Cholera, 2006. *WklyEpidemiol Rec.* 2007; 82: 273-4.
5. WHO. Prevention and control of cholera outbreaks: WHO policy and recommendation. [Internet] 2013. Available from: <http://www.who.int/cholera/technical/prevention/control/en/index4.html>.
6. WHO, Global Health Observatory (GHO): Number of reported cholera cases, August 2011.[Internet] 2011. Available from: [http://www.who.int/gho/epidemic\\_diseases/cholera/cases\\_text/en/index.html](http://www.who.int/gho/epidemic_diseases/cholera/cases_text/en/index.html).
7. Tabatabaei SM, Khorashad AS. Antimicrobial Resistance Patterns of *Vibrio cholerae* Strains Isolated From Afghan and Iranian Patients in Iran. *Int J Infect.* 2015; 2:e22822.
8. ThapaShrestha U, Adhikari N, Maharjan R, Banjara MR, Rijal KR, Basnyat SR et al. Multidrug resistant *Vibrio cholerae* O1 from clinical and environmental samples in Kathmandu city. *BMC Infect Dis.* 2015;15:104.
9. Shah BK, Sharma S, Shakya G, Upadhyay BP. Multiple drug resistance *Vibrio cholerae*, *Salmonella* and *shigella* from Nepalgunj Cholera outbreak and different hospitals of Nepal. *Nepalese journal of Biosciences.* 2012; 2:31-9.
10. Mahmood A. Emergence of nalidixic acid resistant *Vibrio cholerae* O-1 in Karachi. *J Pak Med Assoc.* 1999;49:286.
11. Garrigue GP, Ndayo M, Sicard JM, Fonkoua MC, Lemao G, Durand JP, et al. [Antibiotic resistance of strains of *Vibrio cholerae* isolated in Douala (Cameroon)]. *Bull SocPatholExotFiliales.* 1986; 79:305-12.
12. Wang R, Lou J, Liu J, Zhang L, Li J, Kan B. Antibiotic resistance of *Vibrio Cholerae* O1 El Tor strains from

- the seventh pandemic in China, 1961-2010. *Int J Antimicrob Agents*. 2012; 40:361-4.
13. Ahmed S, Bardhan PK, Iqbal A, Mazumder RN, Khan AI, Islam MS, et al. The 2008 cholera epidemic in Zimbabwe: experience of the icddr,b team in the field. *J Health Popul Nutr*. 2011; 29:541-6.
  14. Centers of Disease Control and Prevention. Antibiotic Treatment: Recommendations for the use of antibiotics for the treatment of cholera. [Internet] 2012. Available from: <http://www.cdc.gov/cholera/treatment/antibiotic-treatment.html>.
  15. Koneman EW, Allen Sd, Janda Wm, Procop GW, Schreckenberger PC, Woods Gl, et al. Color atlas and textbook of diagnostic microbiology, 6th ed. Philadelphia. Lippincott Williams & Wilkins. 2006.
  16. Clinical and Laboratory Standards Institute (CLSI). Performance Standards for Antimicrobial Susceptibility Testing. Twentieth Informational Supplement, M100-S20. Vol. 30. Wayne, PA: CLSI. 2010; 1-153.
  17. Afzal M. Population growth and economic development in Pakistan. *Open Demograph J*. 2009; 2:1-71.
  18. Noor A, akhem J, Ehtesham M, Xu W. Cholera in Pakistan. *Malmö Hodskola, HaskOchsamhälle*. 2010; 10:205-6.
  19. Mohammad A. The global burden of Cholera. *Bull world Health Organ* 2012; 90:209-18.
  20. Lodhi M, Munir T, Karamat K. Dehydrating diarrhea in children due to *V. cholerae*. *Pak Armed Forces Med J*. 2006; 56:50-5.
  21. Mandal J, Dinooop KP, Parija SC. Increasing Antimicrobial Resistance of *Vibrio cholerae* O1 Biotype El Tor Strains Isolated in a Tertiary-care Centre in India. *J Health Popul Nutr*. 2012; 30:12-6.
  22. Jameel SK, Mostafa AS, Abdulmohsin AM, Mohamed NS, Najji SR, Mohammed TT. The Isolation of *V. cholerae* and Other Enteric Bacteria with Molecular Characterization of *V. cholerae* During the Outbreak of Baghdad/Iraq in 2015. *Advances in Microbiology*. 2016 (6): 699-715
  23. Kuttiaat VS, Lodha R, Das, B, Kohli U. Prevalence of Cholera in Pediatric Patients with Acute Dehydrating diarrhea. *Indian J Pediatr*. 2010; 77:67-71.
  24. Kulkarni S, Chillarge C. Antibiotic Susceptibility Pattern of *Vibrio Cholerae* Causing Diarrhea Outbreaks in Bidar, North Karnataka, India. *Int J Curr Microbiol App Sci*. 2015; 4:957-61.
  25. Roychowdhury A, Pan A, Dutta D, Mukhopadhyay AK, Ramamurthy T, Nandy RK, et al. Emergence of Tetracycline-Resistant *V. Cholerae* O1 Serotype Inaba, in Kolkata, India. *Jpn J Infect Dis*. 2008; 61:128-9.
  26. Tabatabaei SM, Khorashad AS. Antimicrobial Resistance Pattern of *V. Cholerae* Strains Isolated From Afghan And Iranian Patients in Iran. *Int J Infect*. 2015; 2:e 22822.
  27. Rafi S, Hussain A, Saeed W, Ali A, Khwaja SA. Changing epidemiology and sensitivity pattern of *Vibrio cholera* at Rawalpindi. *Pak J Med Sci*. 2004; 20:357-60.
  28. Nelson EJ, Nelson DS, Salman MA, Sack DA. Antibiotics for both moderate and severe Cholera. *N Engl J Med*. 2010; 364:5-7.

