



Isolation of biosurfactant producing bacteria from petroleum contaminated sites and their characterization

Rida Batool, Samra Ayub and Iqra Akbar

Department of Microbiology and Molecular Genetics, University of the Punjab, Quaid-e-Azam Campus, Lahore

Abstract

Biosurfactants are microbial amphiphilic compounds which can reduce surface tension between aqueous and hydrocarbon mixtures. Bacterial strains isolated from petroleum contaminated soil of various motor workshops were characterized morphologically and biochemically. Biosurfactant producing ability of the strains was determined and their emulsification activity was screened against different oils. All the selected bacterial strains showed enhanced biosurfactants production with yeast extract as nitrogen source and glucose as carbon source at optimized conditions. These strains also exhibited multiple metal and antibiotics resistance. Isolated biosurfactants of three most promising strains SF-1, SF-4 and SM-1 were extracted by solvent extraction and subjected to TLC technique. The technique indicates the glycolipid nature of the compounds and presence of rhamnose sugar, which was further confirmed by FT-IR analysis. 16srRNA analysis revealed that SF-1 and SM-1 had close resemblance with *Pseudomonas* sp. while SF-4 showed homology with *Enterobacter* sp. Isolation and screening of biosurfactant producing strains from petroleum polluted places proved to be a quick and effective means to find bacterial strains with possible industrial uses.

Keywords: Biosurfactant, TLC, FT-IR, 16s rRNA sequencing, heavy metals.

Introduction

In today's world, petroleum usage has increased to the exponential level and contaminating the environment very badly. Since 1970, more than 5 million tons of oil spills are reported by International Tanker Owners Pollution Federation affecting serious health problems. Petroleum oil is a non-volatile and low molecular compound (Budsabun, 2015). Biosurfactants are surface active compounds produced by different bacteria. These compounds are involved in reducing the surface tension of organic and non-organic mixtures. They make interfaces among various solution depending upon their polarities such as oil and water resulting in lowering the interfacial tension (El-Sheshtawy, *et al.*, 2015). Bio-surfactants are compounds mostly used because they are environment friendly, easily degraded, low toxicity and produced from cheaper substrates (Meybodi, *et al.*, 2013). These compounds have displayed promising surface activity (Hassanshahian, 2014). They are classified into various groups such as glycolipids, lipopeptides, polysaccharide-protein complexes, phospholipids, fatty acids and neutral lipids etc. (Hassanshahian, 2014).

Glycolipids and lipopeptides are low molecular weight surfactants (Ron and Rosenberg, 2001). Glycolipids are further characterized into three subgroups i.e. rhamnolipids, trehalolipids and

sophorolipids. Different bacteria are reported to produce biosurfactants including surfactin by *Bacillus subtilis* and the rhamnolipids from *Pseudomonas aeruginosa* (Batista *et al.*, 2006). Different species of *Bacillus* are reported to produce surfactin and lichenysin such as *B. coagulans*, *B. pumilus* and *B. licheniformis* (Ongena *et al.*, 2008). They are involved in the biodegradation of hydrocarbons, augmentation of oil recovery and elimination of metals from a contaminated soil. A group of biosurfactants known as polymeric biosurfactants such as emulsion, alaskan, biodispersan, liposan and mannoproteins etc. They are usually produced by *Acinetobacter* and some species of yeast. Biosurfactants have exhibited antimicrobial activity (Maier, 2003). Some biosurfactants acts as antimicrobial and therapeutic agents (Rodrigues *et al.*, 2006). Some studies showed that rhamnolipid produced by strains possess certain antibacterial and antifungal activities (Cameotra and Makkar, 2004). Biosurfactants play a significant part in the remediation of certain heavy metals as they form complexes with metals via strong bonds in soil as compared to binding of metal to the soil. This process can be helpful in the management of waste water contaminated with heavy metals (Das *et al.*, 2009). Promising applications of biosurfactants is to improve oil recovery and to develop methods for the extractions of lingering oil which situated in the sections of the

*Email: rida.mm@gmail.com

reservoir which are distant and the oil is imprisoned in the holes by duct pressure (Sen, 2008). Researchers suggest that rhamnolipid is capable of reducing metal toxicity to microbial associations in co-contaminated soils by the combination of metal complexation and in the amendment of cell surface properties through the release of LPS resulting in boosted bioremediation (Sandrin *et al.*, 2000). The current research was designed to isolate biosurfactants producing bacteria from petroleum polluted soil. Because of their diversity, biosurfactants are considered as an attractive group of compounds for their widespread use in various industrial and biotechnological processes.

Materials and Methods

Isolation of bacteria

Greasy soil samples from various service stations located in the vicinity of Punjab University, Lahore were collected in sterilized bottles. To isolate the biosurfactant producing microorganism, one gram of soil sample was inoculated in 100 ml of mineral salt media (Patil *et al.*, 2012) containing 1 to 5% diesel, petrol and Mobil oil instead of glucose as carbon source. After 1 week of incubation on shaker, 50 µl of subculture was taken and then transferred into fresh medium in flasks and was incubated under same conditions. In order to enrich the microbial cultures and to increase the population density, this procedure was repeated thrice. After this, 50 µl of culture was taken and spreaded on MSM agar plate to isolate biosurfactant producing microorganisms. These plates were incubated at 37 °C for 4-5 days (Ahmed *et al.*, 2016). Different colonies appeared on agar plate were observed and selected for further purification.

Qualitative screening

Hemolysis as an indication of biosurfactants production was observed on Blood agar media. This test provides information about what kind of hemolytic enzyme a bacterium possesses (Bicca *et al.*, 1999). Colonies showing hemolysis were tested further by oil spread technique. Oil spread technique is very useful as it can be employed when the activity and quantity of biosurfactants is low. For this purpose, 100 µl of crude oil was added on the surface of 50 ml of distilled water followed by the addition of 10 µl of bacterial culture. A plaque on oil surface indicates biosurfactants activity of the culture. The drop collapse and CTAB blue agar plate method was carried out as described by Bodour *et al.*, 1998; Siegmund and Wagner, 1991, respectively. Drop collapse assay is a very simple assay for screening of biosurfactant producing bacteria while CTAB assay indicates the production of anionic surfactants.

Quantitative screening

Isolates that exhibited positive results in qualitative assay were subjected to quantitative assay and their emulsification index was determined (Shoeb *et al.*, 2015).

Morphological and biochemical characterization of selected strains

Selected isolates were observed for colony morphology, Gram's nature, capsule staining, spore formation and motility. Biochemical nature of selected isolates was characterized by catalase, oxidase, citrate, indole, sugar fermentation and nitrate reduction (Cappuccino and Sherman, 2002).

Optimization of growth parameters for the production of biosurfactants

Different carbon (glucose, sucrose, fructose, and glycerol) and nitrogen sources (yeast extract, beef extract, ammonium chloride, and potassium nitrate) were examined in order to know the best combination of nutrients for biosurfactant production using MSM as a basal media. To optimize conditions for maximum biosurfactant fabrication, strains were grown at a wide range of temperature (27, 37, 45 and 50°C) and pH (3-9). Each factor was changed accordingly by keeping rest of the other constant.

Biosurfactant production in different oils

Activity of biosurfactant producing strains was checked in different oils like gangrene, coconut, sunflower, kerosene and olive oil. After 24 hours, the height of stable emulsion layer is measured by calculating emulsification index (Copper and Goldenberg, 1987).

Antibiotic and metal resistance profiling

Antibiotic resistance profiling was done to determine the susceptibility of bacteria against various antibiotics (Ampicillin, Tetracycline, Streptomycin, Chloramphenicol, Kanamycin and Erythromycin) at different concentrations by plate dilution method. Similarly, to check metal tolerance profile of biosurfactant producing strains, 5% stock solution of cobalt nitrate ($\text{Co}(\text{NO}_3)_2 \cdot 6\text{H}_2\text{O}$), Cadmium chloride ($\text{CdCl}_2 \cdot \text{H}_2\text{O}$), Copper sulfate (CuSO_4), and Zinc sulfate ($\text{ZnSO}_4 \cdot 7\text{H}_2\text{O}$) were prepared and nutrient agar plates were supplemented with respective metal stock solution. MIC of each heavy metal was determined by plate dilution method. Appearance of growth on metal / antibiotic supplemented plate indicated the positive result for resistance.

Solvent extraction of crude biosurfactants

Solvent extraction was performed for the isolation of crude biosurfactants. Bacteria were cultured in MSM broth for



48 hours on a shaker at 100 rpm at 37°C, centrifuged at 10,000 rpm for 20 minutes under cooling conditions. Supernatant was taken and concentrated HCl was added to adjust the pH at 2.0 and left it overnight at 4°C. Next day, centrifuged the acidified supernatant at 10,000 rpm for 20 minutes at 4°C and grey white precipitate was collected for further extraction of biosurfactant compounds. Ten milliliters of chloroform and methanol in the ratio (2:1 v/v) were added in order to precipitate the pellet and incubated at 30°C for 15 minutes. The content was centrifuged for 20 min under cooling conditions and supernatant was evaporated by air drying. The residue was dispensed in sodium phosphate buffer (pH 7.0) and stored at 4°C.

Purification of biosurfactants

Chloroform was used to dissolve the crude extracts and then analyzed by thin layer chromatography (TLC). About 10 µl of crude extract was placed on silica gel plate at a 2 cm above from the bottom of the plate. Tween-80 was taken as a control. Plate was dried, and developed in a solvent system of chloroform: methanol in the ratio 95:5. After developing, plate was taken out and allowed to air dry. Developed plate was observed under UV at longer and shorter wavelength and stained it with iodine fumes. Yellow colored spots were observed.

FT-IR analysis

FTIR (Fourier Transform Infra-red Spectroscopy) is a sensitive and primarily technique for identifying the functional groups present in the sample. KBr pellet of crude extracts were made and the frequency range is measured as wave numbers 4000 – 500 cm⁻¹.

16srRNA sequencing of most promising strains

For 16S rRNA sequencing, bacterial strains were sent to MacroGen Inc, Seoul, Korea for 16S rRNA gene sequencing. 16S rRNA sequence was determined using primers 518F (CCAGCAGCCGCGGTAATACG) and 800F (TACCAGGGTATCTAATCC). Reverse complementary sequence was made by reverse primer with Chromas Lite 2.01. CLC DNA Workbench software was used for editing, aligning and assembling of forward, reverse and internal sequences. BlastN was used to check the harmony sequences against GenBank. Extreme homology of the query sequences to the database sequences was determined.

Results

Isolation of bacteria

Total 30 bacterial strains were isolated from samples of soil contaminated with petroleum and its products. Ten out of thirty strains showed the biosurfactant activity.

Screening for biosurfactant production

Ten selected biosurfactant producing strains were screened for biosurfactant production by numerous tests. All the strains were positive for blood agar hemolysis, oil spreading technique, drop collapse assay and CTAB blue agar (figure 1, table 1).

Table 1: Screening of bacterial strains for biosurfactants production

Bacterial Strain	Blood Hemolysis	Oil Spreading	Drop Collapse	CTAB
SF-1	+	+	+	+
SF-2	+	+	+	+
SF-3	+	+	+	+
SF-4	+	+	+	+
SM-1	+	+	+	+
SM-2	+	+	+	+
SM-3	+	+	+	+
SW-1	+	+	+	+
SW-2	+	+	+	+
SW-3	+	+	+	+

Morphological and Biochemical characterization

Selected strains exhibited different colony morphology with pigments, texture and elevations. Majority of biosurfactant producing strains were Gram negative like, *Acinetobacter* SF-3, *SM-3* *Enterobacter*- SF-4, *Pseudomonas* SF-1, SM-1, SW-1, SW-2, whereas two strains, SF-2 and SM-2 were Gram positives. Catalase was positive for all the strains except SF-2 and SM-2. Oxidase was negative for SF-2, SF-3, SF-4, SM-2 and SM-3 whereas results of sugar fermentation were exactly opposite to oxidase. Citrate was negative for SF-2 and SM-2. Nitrate reduction was negative for SF-3, SF-4, SM-2 and SM-3 whereas Indol was positive for SF-2 and SM-2.

Optimization of growth parameters for the production of biosurfactants

Glucose was best carbon source and yeast extract was best nitrogen source to enhance the growth of strains as shown in figure 2 and 3, respectively. Sucrose is second best carbon source; fructose gave transitional growth while glycerol reduced the growth. Beef extract and potassium nitrate overall reduced growth. Temperature 37°C and pH 7 was the best condition for all strains as shown in figure 4 and 5.

Emulsification index of Biosurfactants in different oils

It was found that Gangrene oil showed a high emulsification index, Kerosene oil and olive oil showed



partial while cooking oil and coconut oil show low emulsification index (Table 2).

Antibiotic and metal resistance profiling

Most of the strains showed multiple antibiotic resistances. They were also resistant to different metals at variable concentrations. SF-1, SF-4, SM-1 were resistant to Streptomycin up to 500 $\mu\text{g ml}^{-1}$ and

FT-IR

TWEEN-80 is a biosurfactant which was taken as a control. It showed strong absorption bands which are compared with our biosurfactant producing strains (Figure 6 A). The absorption bands at 3132 cm^{-1} is due to O-H stretching, Carbonyl group stretching which is the characteristic of ester compound was found at 1720

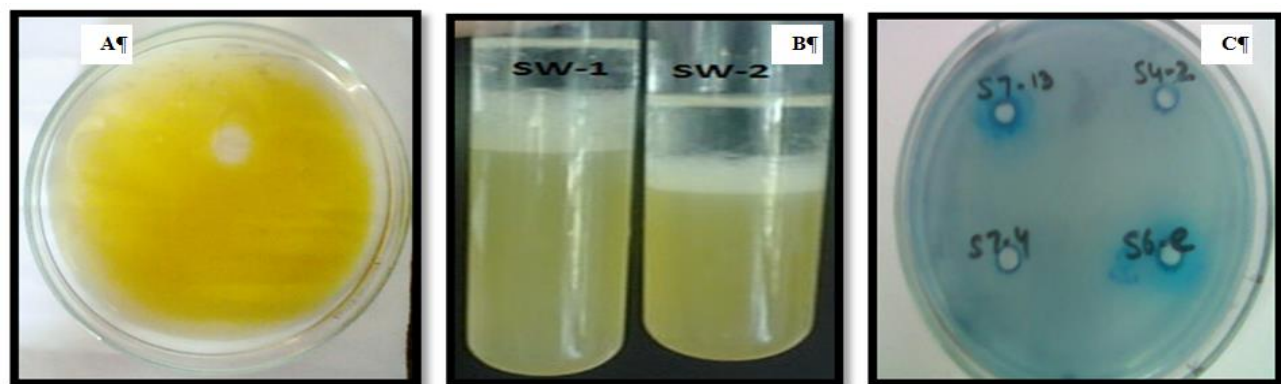


Figure 1: Biosurfactant activity of isolated bacterial strains A Oil spreading technique B Emulsification index C) CTAB Blue agar plate method

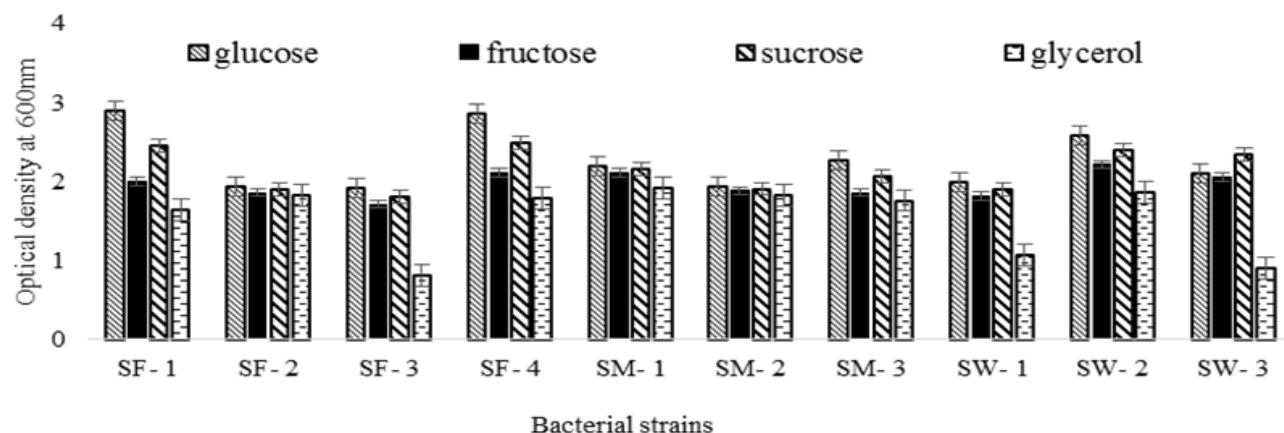


Figure 2: Production of biosurfactants by isolated bacterial strains with different carbon sources

Ampicillin up to 300 $\mu\text{g ml}^{-1}$. Their resistance to selected heavy metals was up to 800- 1000 $\mu\text{g ml}^{-1}$ as shown in table 3.

Extraction and purification of biosurfactants

In order to examine the chemical nature of honey colored residues obtained by solvent extraction, TLC was done. Yellow brown colored spots on TLC in iodine fumes indicated that biosurfactant produced by the isolates was lipid in nature. R_f value of biosurfactant produced by our strains was 0.61-0.64 which is similar to standard rhamnase sugar ($R_f = 0.63$).

cm^{-1} . Absorption bands at 1300 cm^{-1} indicates the structure identical to rhamnolipid. Strong adsorption peak at 2,920 cm^{-1} is anticipated to be the C-H stretching vibrations of the hydrocarbon chain positions. Due to the stretching vibrations of C-H of hydrocarbon chain position robust absorbance was seen at 1720 cm^{-1} . Though, absorption peaks in the range of 1,300–1,100 cm^{-1} indicated that chemical structure was identical to rhamnolipid.

The important absorption bands located at 1738 and 2929 cm^{-1} which indicated that it had similarity with rhamnolipid. The presence of ester bond due to C=O



stretching is located at 1738cm^{-1} . Absorption peaks at 2929cm^{-1} showed the stretching vibrations of hydrocarbon chains. Absorbance in the range of $1,300\text{--}1,100\text{ cm}^{-1}$ showed the occurrence of bands between carbon atoms and hydroxyl groups in the chemical structures of rhamnose rings. The absorption peak at 3139cm^{-1} indicated the presence of hydroxyl groups (Figure 6 B, 6 C and 6 D).

16SrRNA sequencing of biosurfactant producing strains

The two best bacterial isolates among all biosurfactant producers were selected for 16SrRNA sequencing. These strains were SF-4 and SM-1. The 16SrRNA gene sequences with those in the NCBI sequence database (GenBank) were compared through BLAST (<http://www.nlm.nih.gov/>

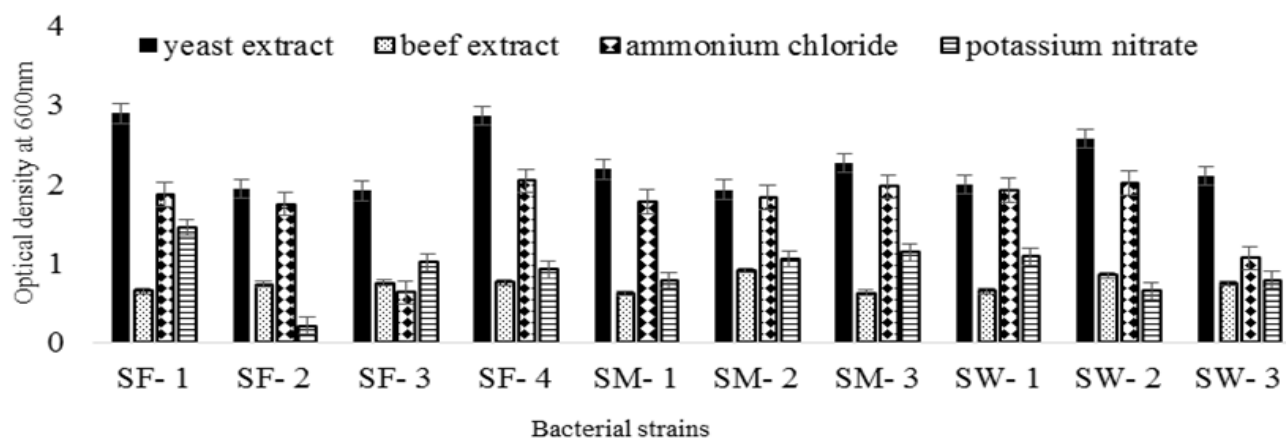


Figure 3: Production of biosurfactants by isolated bacterial strains with different nitrogen sources

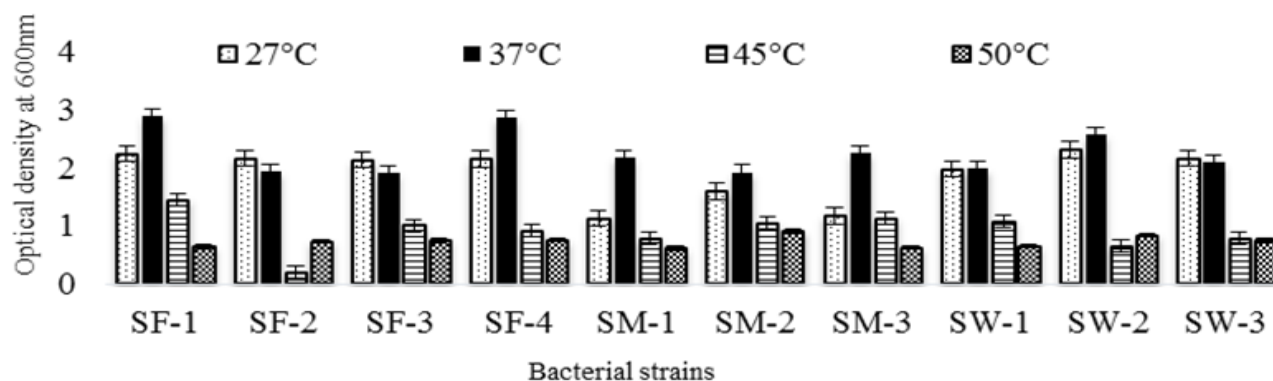


Figure 4: Production of biosurfactants by isolated bacterial strains at different growth temperature

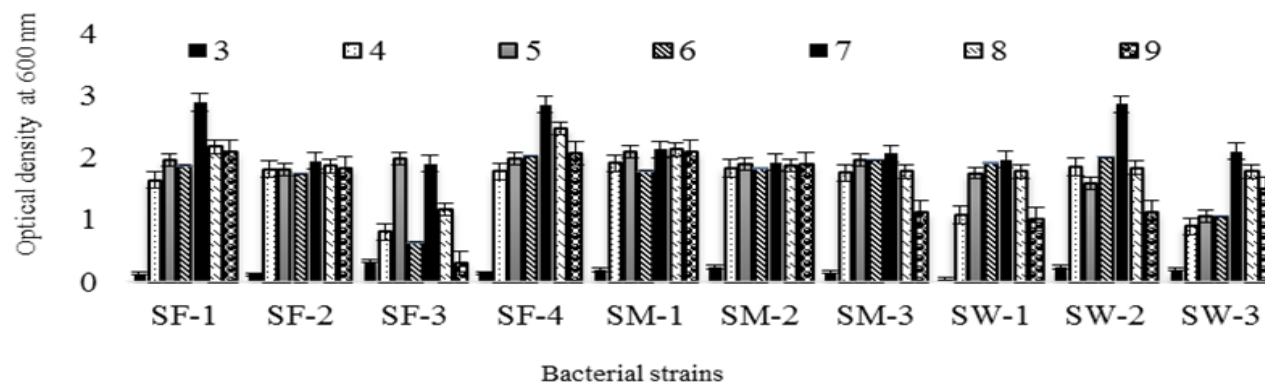


Figure 5: Production of biosurfactants by isolated bacterial strains at different pH



Table 2: Emulsification index (%) of isolated biosurfactant producing bacteria with different oils

Bacterial Strains	Gangrene Oil	Kerosene Oil	Coconut Oil	Cooking Oil	Olive Oil
SF-1	50	27	27	34	40
SF-2	16	11	20	10	16
SF-3	13	16	20	10	22
SF-4	34	20	20	40	30
SM-1	53	28	30	37	27
SM-2	50	40	34	50	43
SM-3	30	34	37	43	43
SW-1	20	25	16	16	20
SW-2	32	30	30	20	25
SW-3	30	28	31	22	30

Table 3: Heavy metal resistance profiling of isolated biosurfactant producing bacteria

Bacterial Strain	Heavy Metals ($\mu\text{g ml}^{-1}$)				
	CdCl ₂	ZnSO ₄	CoCl ₂	CoNO ₃	CuSO ₄
SF-1	+1000	+800	+900	+800	+1000
SF-2	+600	+600	+600	+600	+600
SF-3	+600	+600	+600	+600	+600
SF-4	+900	+800	+800	+700	+700
SM-1	+900	+1000	+800	+800	+800
SM-2	+600	+600	+600	+700	+600
SM-3	+600	+600	+700	+700	+600
SW-1	+600	+600	+700	+700	+700
SW-2	+700	+700	+700	+700	+700
SW-3	+700	+700	+700	+700	+700

+ indicates growth up to...

BLAST). 16SrRNA gene sequence of the strain SM-1 was most closely related to *Pseudomonas* sp. while SF-4 showed homology with *Enterobacter* sp. (Figure 7 A, B). Sequences obtained were submitted to Genbank under the accession number KP794925 (*Enterobacter cloacae* strain SF-4) and KP794924 (*Pseudomonas aeruginosa* strain SM-1).

Discussion

The current study is aimed on the isolation of biosurfactant producing bacteria from hydrocarbon polluted soil. According to several published studies, investigators reported the successful isolation of biosurfactant producing organisms from soil polluted with hydrocarbons or mineral oils (Mukherjee *et al.*, 2009; Mulligan, 2005). In this study ten biosurfactant producing strains were isolated from petroleum contaminated soil. Hemolytic activity seems to be a good selection measure in the exploration for biosurfactant producing bacteria.

Biosurfactants producing strains hemolysed the red blood cells and showed α or β hemolysis. In present study,

selected strains exhibited hemolytic activity which is principal screening technique for biosurfactant producing bacteria. Other methods such as oil spreading technique, drop collapse assay, emulsification index need to be carried out for further confirmation (Shoeb *et al.*, 2015). The drop from supernatant of all the isolated strains collapsed and control remained in beaded form. Diameter of a sample and concentrations of biosurfactant in a drop have a direct relationship and if the drop having no biosurfactant remains in beaded form due to the hydrophobicity of oily surface that cause aggregation of droplets (Bodour and Miller-Maier, 1998). Selected strains also showed oil displacement activity which was confirmed by oil spreading technique (Patil *et al.*, 2012). Emulsification activity of all isolated strains was checked; all of them formed very stable emulsions. The cells developed a strong hydrophobic character during exponential phase and it may lose when cell enters into a stationary phase. Emulsification reduced oil drop size less than 1 μm (Bredholt *et al.*, 2002). CTAB blue agar plate assay is good for screening of new anionic biosurfactant production strains or mutants (Perfumo *et al.*, 2006). Three biosurfactant producing strains, including SF-



1, SF-4 and SM-1 produced blue haloes around their wells. The size of halo depends on rhamnolipids production (Siegmund and Wagner, 1991).

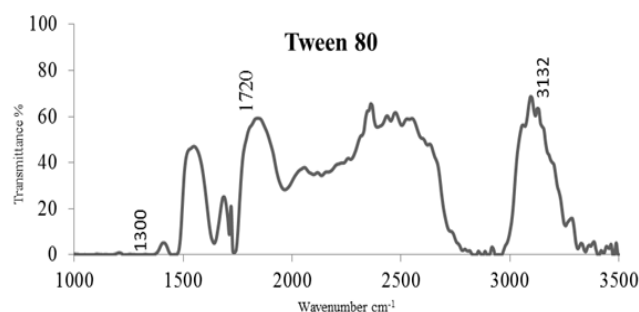


Figure 6 A: FT-IR spectrum of Tween 80

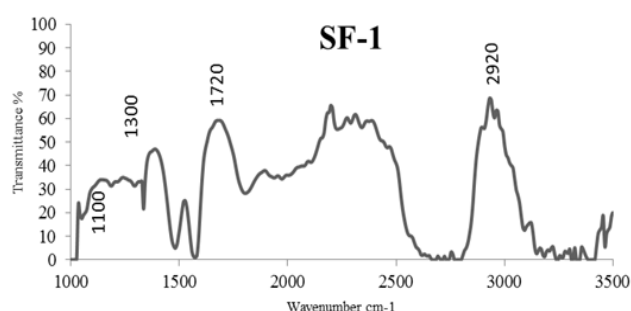


Figure 6 B: FT-IR spectrum of purified biosurfactant producing strain SF-1

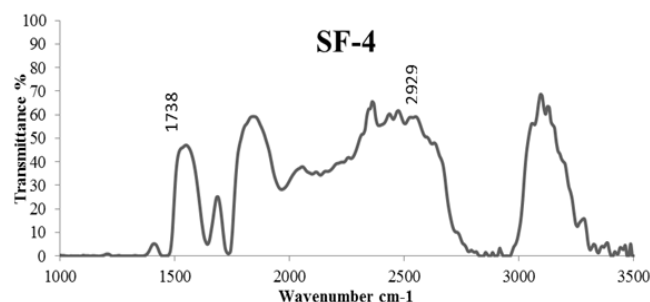


Figure 6 C: FT-IR spectrum of purified biosurfactant producing strain SF-4

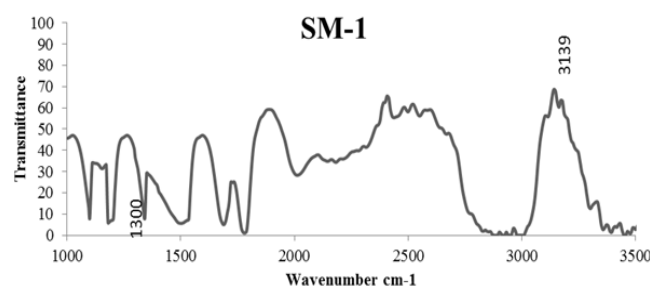


Figure 6 D: FT-IR spectrum of purified biosurfactant producing strain SM-1

Biosurfactants production is greatly influenced by the environmental factors and growth conditions such as pH, temperature, carbon sources and nitrogen sources by influencing the cellular growth or activity. Similar findings were reported by Kim *et al.* (1997). It is described that biosurfactant producing bacteria showed maximum growth by using glucose and yeast extract (Khopade *et al.*, 2012). Beef extract and potassium nitrate overall reduced microbial growth (Hamzah 2013). Maximum growth of biosurfactants producing bacteria was at temperature 37°C and at pH 7. It is in accordance with the findings of previously published reports (Guerra-Santos *et al.*, 1986; Hamzah 2013). In order to determine the potential of biosurfactants, emulsification activity estimation is one of the principle methods. Emulsifying activities (E24) regulate the yield of bioemulsifier (Shoeb *et al.*, 2015). The most potent oil biodegrading strain in current research was *Pseudomonas* showed highest emulsification index (Shoeb *et al.*, 2015).

It was found that Gangrene oil showed a high emulsification index, Kerosene oil and olive oil also exhibited good response (Abouseoud *et al.*, 2007; Chander *et al.*, 2012). Many of the strains showed multiple antibiotic resistances. Bacterial strains sequestered from contaminated sites were 40-200 times more resistant to heavy metals than related strains isolated from unpolluted sites (Bopp *et al.*, 1983). It was found that the heavy metals were found among hydrocarbons and their presence produces a difficulty in biodegradation. So as a result, the microorganisms isolated from hydrocarbon contaminated sites show less sensitivity to metals prevailing on the site, such as quicksilver, copper and chromium (Fredrickson *et al.*, 1995). Stress resistance in isolates has evidently shown the presence of pollutants at the sampling place (Shoeb *et al.*, 2015). Thin layer chromatography (TLC) is a simple technique, used to purify the products in small quantities (Symmank *et al.*, 2002). The biosurfactant produced by the isolates was lipid in nature as it produced yellow brown colored spots in iodine fumes with R_f value of (0.61) identical to standard rhamnose sugar ($R_f = 0.63$) (Stahl, 1969, Vyas and Dave, 2011). Most of researchers reported *Pseudomonas* as an efficient producer of rhamnolipid biosurfactant (Vyas and Dave, 2011). In FT-IR spectrum of partially purified biosurfactants produced by SF-1, SF-4 & SM-1 further confirmed the presence of a rhamnolipid structure. The adsorption bands found are in accordance with the findings of Guo *et al.* (2009) presenting the existence of rhamnose rings and long hydrocarbon chains (Moussa *et al.*, 2014). Most of the bacteria that are reported as hydrocarbon degraders belong to *Pseudomonas* (Widada *et al.*, 2002). In the present study, bacterial strain SM-1 showed homology to *Pseudomonas aeruginosa* whereas

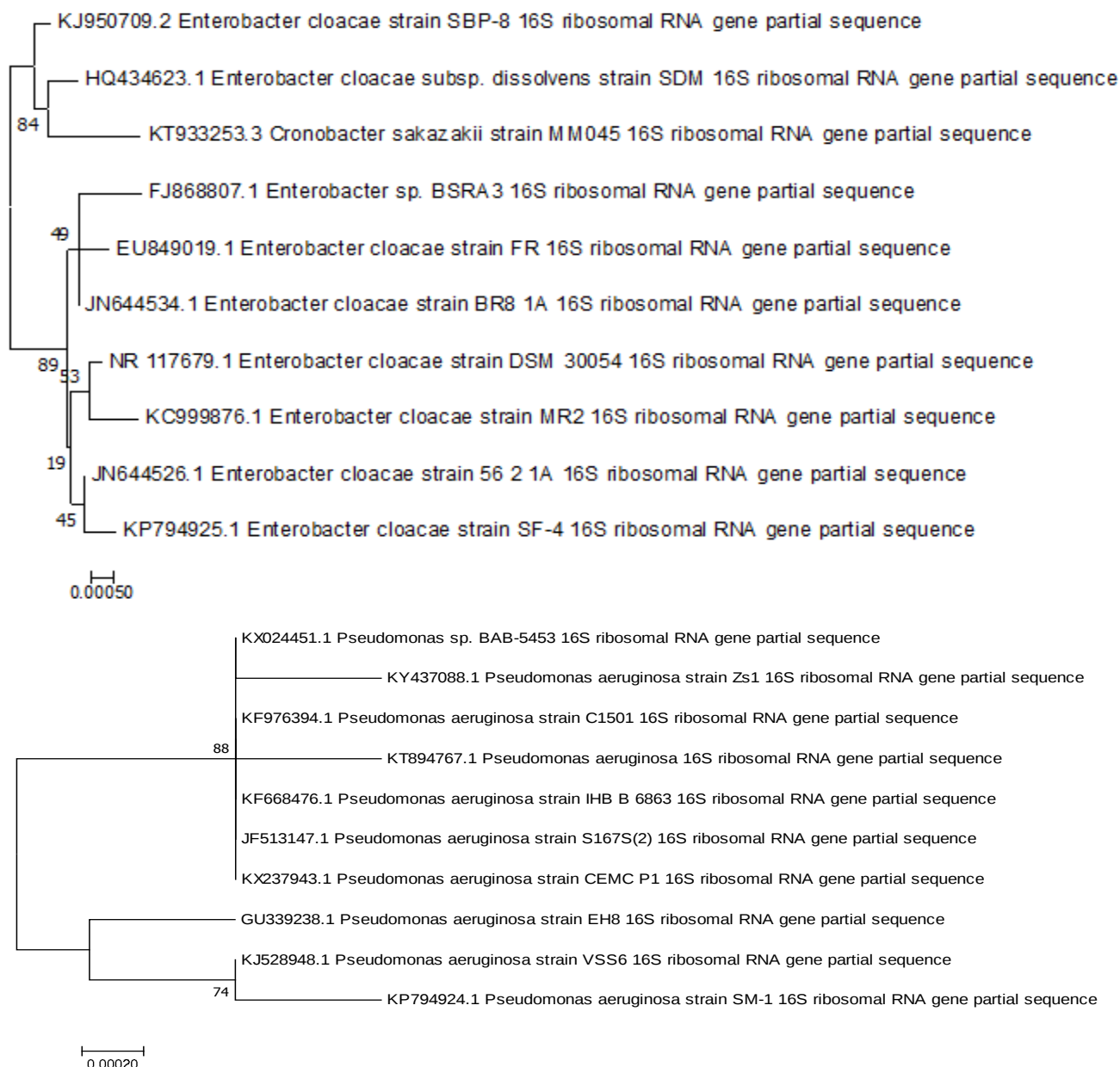


Figure 7: The evolutionary history was inferred using the Neighbor-Joining method (Saitou and Nei, 1987). Evolutionary analyses were conducted in MEGA7 (Kumar *et al.*, 2106). A: Strain SF-4; B: Strain SM-1

bacterial strain (SF-4) showed belonging to *Enterobacter cloacae*. Production of biosurfactant by *Enterobacter cloacae* LK5 isolated from oil contaminated soil has reported previously (Sarin and Sarin, 2008).

Future prospective

Future of biosurfactants will depend on their production cost and applications. Biosurfactants can be used in environmental biotechnologies due to their low toxicity and

biodegradability but their high production cost is major hindrance in the commercial success of biosurfactants. They may be helpful in the bioremediation of petroleum and lubricant contaminated sites.

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