

BIOLOGICAL SCREENING OF SYNTHESIZED OXALAMIDE DERIVATIVES FOR ANTIOXIDANT AND ANTICANCER PROPERTIES

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ABSTRACT

In the present study oxalamide derivatives were synthesized by treating oxalyl chloride with different derivatives of aniline (namely aniline, 5-bromo, 2,5-dimethyl, 3-bromo, 4-bromo, 3-chloro, 4-chloro, and 4-(Trifluoromethyl) aniline). The synthesized compounds were characterized using ¹H-NMR, EI-MS, and FTIR spectrometry. Synthesized oxalamides were also evaluated for their antioxidant and anticancer activities. The antioxidant activity was studied using DPPH radical scavenging assay whereas the anticancer activity was studied against blood cancer cell lines, namely RAJI and DOHH₂. The oxalamide derivatives containing bis (2, 5-dimethylphenyl), bis (3-bromophenyl), and bis (4-bromophenyl) substitutions showed significant antioxidant properties which were comparable with butylated hydroxyanisole; standard drug. However, none of the synthesized oxalamide derivatives showed any significant anticancer activity.

Keywords: oxalamide, antioxidant, anticancer, characterization, synthesis, biological-screening, amides

INTRODUCTION

Oxygenated species possessing unpaired electron, commonly known as reactive oxygen species ROS, are continuously released in the body during different enzymatic and non-enzymatic biological reactions (Dickinson and Chang, 2011; Mahmoud *et al.*, 2011). Reactive Oxygen species are involved in signal transmissions to brain and body defense mechanism against pathogens (Dickinson and Chang, 2011). However, overproduction of ROS can cause oxidative stress and can damage biomolecules such as proteins, nucleic acid, lipids. Oxidative stress eventually give rise to inflammation and promote autoimmune diseases (Balmus *et al.*, 2016; Sznarkowska *et al.*, 2017; Tian *et al.*, 2017; Zuo *et al.*, 2016) such as cancer, cardiovascular disorders, atherosclerosis, malaria, rheumatoid arthritis, Alzheimer's disease, Parkinson's disease, neurodegenerative diseases (Raziq *et al.*, 2017), cataracts, chronic inflammation, and pre-mature aging (Aruoma, 1998; Fang *et al.*, 2002; Finkel and Holbrook, 2000; Waris and Ahsan, 2006).

Antioxidants are compound that reduce the free radicals and ROS and subsequently decrease oxygen stress. Although a number of ROS inhibitors (aspirin, ibuprofen, and naproxen) have been reported (Khalaf *et al.*, 2008; Lee *et al.*, 1998) most of them have their associated side effects including allergies, non-selectivity and effects related to central nervous system (Sostres *et al.*, 2010).

Cancer is rapid, uncontrolled multiplication of abnormal cells. There are different types of cancer based on part of body affected. Common types include cancer of bladder, bone, bowel, brain, breast, cervical, eye, gallbladder, head and neck, kidney, liver, lung, anal, skin, esophageal, ovarian, pancreatic, penile, prostate, intestine, soft tissue, stomach, testicular, thyroid, uterine, vaginal, etc. Some other types of cancer are Hodgkin lymphoma, Leukemia, Mesothelioma, Myeloma, Non-Hodgkin lymphoma. It is known that cancer is affecting millions of people per annum around the globe as reported by cancer research UK (2019). In recent years, many laboratories are extensively involved in the synthesis of new anticancer agents and designing cancer therapies with least side effects, less drug resistance and having high efficacies (Giordano and Petrelli, 2008). Although there are many successes the absolute effectiveness has not been achieved against any disseminated cancer (Hanikoglu *et al.*, 2019; Nepali *et al.*, 2014).

Lymphoma is a type of cancer in which white blood cells (lymphocytes) proliferate abnormally. 90% of the lymphomas are Beta cell lymphomas. Here, we selected two beta cell lymphoma cell lines, RAJI and DOHH₂. The clonal proliferations of B-cells show a high degree of variation in terms of clinical and presenting features, histopathology, immunophenotype, and genetics.

Butylated hydroxytoluene (BHT), butylated hydroxyanisole (BHA), tertiary butyl hydroquinone (TBHQ), propyl gallate (PG) are commonly used synthetic antioxidants. They have limited scope because of their low thermal stability, unaesthetic features and harmful health effects. Amides have great pharmacological history and possess diverse range of applications. (Rajput and Sharma, 2018). They are famous for their anti-bacterial, anti-fungicidal, anti-convulsant, lipoxygenase inhibition, anesthetic, and platelet aggregation (Fazal-ur-Rehman *et al.*, 2019; Malki *et al.*, 2017). Amides can be synthesized from oxalic acid are diamides which are classified as oxalamides. Being diamides, they can act as bidentate ligand donating two pairs of lone pairs to a single metal and can also act as bis bidentate ligand donating their lone pairs to two different metal ions. Oxamides, as bis-bidentate ligands, can be used to synthesize supra-molecules. In pharmaceutical field, study of antioxidants, antibacterial and enzyme inhibitions remain a vital area of research as these can depreciate human skin, body, can be the reason of different disease or even can cause death.

Present work comprised of the synthesis of selected amides, their characterization, and evaluation of their antioxidant and anticancer activities. The amides synthesized were diphenyloxalamide, bis (2,5-dimethylphenyl)oxalamide, bis (3-bromophenyl)oxalamide, bis (4-bromophenyl)oxalamide, bis (3-chlorophenyl)oxalamide, bis (4-chlorophenyl)oxalamide, and bis (4-(trifluoromethyl)phenyl)oxalamide. The antioxidant activity of synthesized amides was evaluated and compared to standard antioxidants butylated hydroxyanisole (BHA). The anticancer activity of amides against RAJI and DAHH₂ cancer cells was compared to standard anticancer drugs, Imatinib and Dasatinib.

MATERIAL AND METHODS

General Experimental:

All chemicals used in this study were Reagent grade and purchased from sigma, BDH or Merck. Oxalyl chloride (Merck cat # 8070660025), triethylamine (Merck cat # 8083520100), tetrahydrofuran (Merck cat # 1097312500), n-hexane (Merck cat # 1007952500), ethyl acetate (Merck cat # 1007892500), aniline (Sigma Aldrich cat # 8222561000), 2,5-dimethylaniline (Sigma Aldrich cat # 102253), 3-bromoaniline (Sigma Aldrich cat # 180025), 4-bromoaniline (Sigma Aldrich cat # 8016000100), 3-chloroaniline (Sigma Aldrich cat # 8026120250), 4-chloroaniline (Sigma Aldrich cat # 8026130100), 4-trifluoromethylaniline (Sigma Aldrich cat # 224936), butylated hydroxyanisole (Sigma Aldrich cat # W218308), DMSO, 2, 2'-diphenyl-1-picryl hydrazyl (Sigma Aldrich cat # D9132). Aluminum plates pre coated with silica gel were used for TLC (Supelco cat # 99569). Shimadzu 8900 IR spectrophotometer, Avane Bruker 400MHz ¹H NMR spectrometer and JEOL MS Route 600H EI Mass spectrometer were used for characterization.

General procedure for the Synthesis of ligands:

Ten mMole of selected amine and 0.5mL of triethyl amine were dissolved in 80ml of tetrahydrofuran (THF), the resulting solution was cooled in ice bath ($\approx 5^{\circ}\text{C}$). To this solution 5.0mmole oxalyl chloride were added and the mixture was stirred at room temperature. The progress of the reaction was monitored using TLC. After 24 hours of stirring, reaction mixture was dried on rotavapor. The product formed was washed off using sodium bicarbonate solution and water was used to wash resulting solid. After washing solid was dried under vacuum. For further purification of oxalamides column chromatography was performed on silica gel column using n-hexane: ethyl acetate (7:3) as eluent (Fazal-ur-Rehman *et al.*, 2019).

BIOLOGICAL SCREENING

Antioxidant Activity

2, 2'-diphenyl-1-picryl hydrazyl (DPPH) Radical Scavenging Activity:

The antioxidant activity of synthesized oxamides was evaluated for DPPH free radical scavenging. The method is based on the formation of colorless hydrazine by the protonation of DPPH free radicals. A stock solution of 0.3 mM DPPH was prepared using ethanol as solvent. A series of solution for each of the synthesized oxamide was prepared using DMSO as solvent. The concentrations used were 500, 250, 150, 125, 62.5, 31.2, 15.6, 7.8 μM oxamide. A pit of a 96-well plate was used as a reaction vessel. To each of the eight wells 10 μl of respective oxamide solution was transferred followed by addition of 90 μl of DPPH stock solution in each well. A separate well was used for control. The incubation temperature was 37°C and the stirring time was 30 minutes. Absorbance of solutions at the end of reaction time was recorded at 517nm using microtiter plate reader (Spectra max plus 384 Molecular devices USA). The method was standardized using Butylated hydroxyanisole (BHA). Percent radical

scavenging activity was calculated using DMSO treated control. IC_{50} values were determined using EZ fit software (Perrella Software, USA) (Ali *et al.*, 2009; Siddiq *et al.*, 2012).

$$DPPH \text{ radical scavenging effect (\%)} = \left(Ac - \frac{As}{Ac} \right) \times 100$$

Where, As is the absorbance of sample and Ac is the absorbance of control.

Alamar blue assay for cell cytotoxicity analysis

Alamar blue assay was used to determine cytotoxicity of synthesized oxalamide derivatives. For each oxalamide derivatives, two cancer cell lines RAJI and DOHH₂ (The two B lymphoma cancer cell lines highly malignant Raji (Burkitt's lymphoma) and low malignant DOHH₂ (follicular lymphoma); were kindly gifted from Dr A. Kluin-Nelemans, Groningen, the Netherlands). The cell lines were seeded in 10×10^5 cells per well with 200 μ L final reaction volume. Medium used for this was RPMI-1640 medium which contained 10% heat inactivated fetal bovine serum (PAA laboratories). Different concentrations of oxalamides were then added in cells and incubated for 48 hours. After this time, the cells were incubated for 4 hours at 37°C in humidified atmosphere containing 5% CO₂ along with addition of 22 μ L Alamar blue dye 10%. After this incubation time fluorescence recording were taken at (λ_{ex} = 560 nm, λ_{em} = 590 nm) in fluorometer (O'Brien *et al.*, 2000).

RESULTS AND DISCUSSION

Chemistry

The summarized synthesis scheme is shown below (Fig. 1). Oxalamide derivatives were white solid, non-hygroscopic, amorphous, stable and soluble only in DMSO at room temperature but dissolve in acetonitrile, ethanol and ethyl acetate on heating.

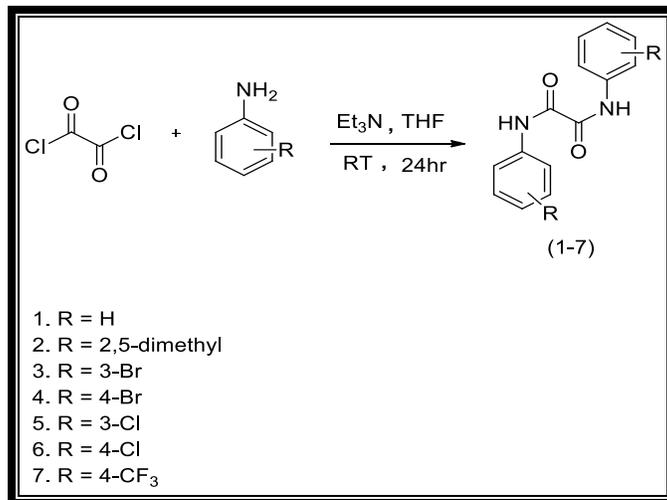
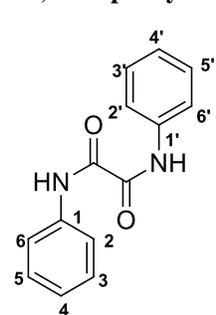


Fig. 1. Synthesis of oxalamides.

Characterization

The characterization and physicochemical data obtained from spectroscopic techniques, namely, NMR, EI-MS, and IR for the synthesized oxalamides is shown as follows.

N1,N2-diphenyloxalamide (1)



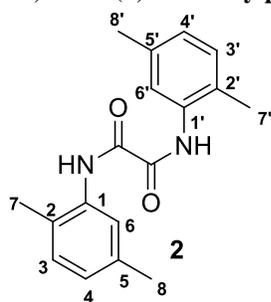
Yield: (38%), White solid.

M.p. : 252-252 °C.

¹H-NMR (DMSO, 300 MHz) δ 10.83 (s, 2H, N-H), 7.87 (d, 4H, J = 8.4 Hz, H-2, H-2', H-6', H-6'), 7.37 (t, 4H, J = 7.5 Hz, H-3, H-3', H-5, H-5'), 7.15 (t, 4H, J = 7.2 Hz, H-4, H-4').

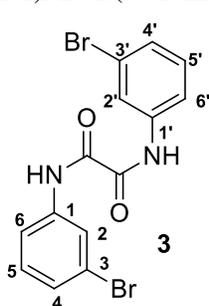
MS (EI): 240.0[M⁺], 120.93.

IR (KBr) ν_{max} cm⁻¹: 3303.8 (NH), 3055.0 (sp²-CH), 1668.3 (C=O), 1596.9, 1523.7 (C=N, C=C), 1436.9 (CH bending aromatic), 1311.5 (C-N).

N1,N2-bis(2,5-dimethylphenyl)oxalamide (2)

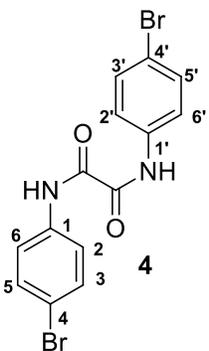
Yield: (39%), White solid.

M.p.: 228 °C.

 $^1\text{H-NMR}$ (DMSO, 300 MHz) δ 10.16 (s, 2H, N-H), 7.33 (s, 2H, H-6, H-6'), 7.16 (d, 2H, $J = 7.8\text{Hz}$, H-4, H-4'), 7.00 (d, 2H, $J = 7.8\text{Hz}$, H-3, H-3'), 2.28 (s, 6H, 2-CH₃), 2.28 (s, 6H, 2-CH₃).
MS (EI): 296.1[M⁺, 100%], 148.0, 121.0.IR (KBr) ν_{max} cm⁻¹: 3255.6 (NH), 3050.6 (sp²-CH), 2920.0 (sp³-CH), 1670.2 (C=O), 1577.1 (C=N), 1521.7 (C=C), 1471.6 (C-H bending aromatic), 1217.4 (C-N)**N1,N2-bis(3-bromophenyl)oxalamide (3)**

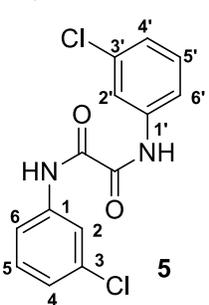
Yield: (49.8%), White solid.

M.p. : 217-219 °C.

 $^1\text{H-NMR}$ (DMSO, 300 MHz) δ 11.05 (s, 2H, N-H), 8.13 (s, 2H, H-2, H-2'), 7.86 (t, 2H, $J = 2.4\text{ Hz}$, H-5, H-5'), 7.35 (m, 4H, H-6, H-4', H-6').
MS (EI): 398.3[M⁺], 399.3[M+1], 400.3[M+2], 199.1, 171.0[100%].IR (KBr) ν_{max} cm⁻¹: 3340.5 (NH), 3055.0 (sp²-CH), 1691.5 (C=O), 1583.4 (C=N), 1525.6 (C=C), 1400.2 (C-H bending aromatic), 1294.1 (C-N).**N1,N2-bis(4-bromophenyl)oxalamide (4)**

Yield: (79.3%), White solid.

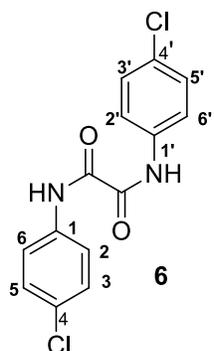
M.p. : 260 °C.

 $^1\text{H-NMR}$ (DMSO, 300 MHz) δ 10.97 (s, 2H, N-H), 7.84 (d, 4H, $J = 6.6\text{ Hz}$, H-3, H-5, H-3', H-5'), 7.57 (d, 4H, $J = 6.6\text{ Hz}$, H-2, H-6, H-2', H-6').
MS (EI): 398.1[M⁺], 399.1[M+1], 400.0[M+2], 199.1, 171.1[100%].IR (KBr) ν_{max} cm⁻¹: 3292.3 (NH), 3045.4 (sp²-CH), 1662.5 (C=O), 1589.2 (C=N), 1510.2 (C=C), 1390.6 (C-H bending aromatic), 1294.1 (C-N).**N1,N2-bis(2-chlorophenyl)oxalamide (5)**

Yield: (31%), White solid.

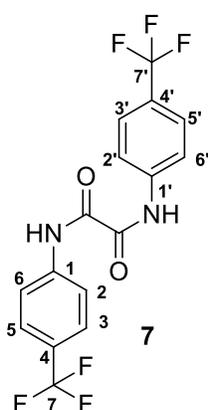
M.p. : 199-201 °C.

 $^1\text{H-NMR}$ (CDCl₃, 300 MHz) δ 9.29 (s, 2H, N-H), 7.78 (s, 2H, H-2, H-2'), 7.49 (d, 2H, $J = 6.0\text{ Hz}$, H-4, H-4'), 7.31 (t, 2H, $J = 6.0\text{ Hz}$, H-5, H-5'), 7.19 (d, 2H, $J = 6.0\text{ Hz}$, H-6, H-6').
MS (EI): 308.2[M⁺], 309.3[M+1], 310.3[M+2], 154.1, 127.1[100%].IR (KBr) ν_{max} cm⁻¹: 3305.8 (NH), 3076.2 (sp²-CH), 1668.3 (C=O), 1589.2 (C=N), 1521.7(C=C), 1425.3(Aromatic CH bending), 1272.9 (C-N).

N1, N2-bis(4-chlorophenyl)oxalamide (6)

Yield: (72%), White solid.

M.p. : 293-295 °C.

¹H-NMR (CDCl₃, 300 MHz) δ 10.96 (s, 2H, N-H), 7.89 (d, 4H, *J* = 6.6Hz, H-3, H-5, H-3', H-5'), 7.44 (d, 4H, *J* = 6.3 Hz, H-2, H-6, H-2', H-6').MS (EI): 308.0[M⁺], 308.9[M+1], 310.0[M+2], 154.9, 127.0[100%].IR (KBr) ν_{max} cm⁻¹: 3296.1 (NH), 3099.4 (sp²-CH), 1662.5 (C=O), 1589.2 (C=N), 1514.0 (C=C), 1392.5(C-H bending aromatic), 1291.6 (C-N).**N1,N2-bis(4-(trifluoromethyl) phenyl)oxalamide (7)**

Yield: (60%), White solid.

M.p. : 305-307 °C.

¹H-NMR (DMSO, 300 MHz) δ 11.21 (s, 2H, *N-H*), 8.09 (d, 4H, *J* = 6.3Hz, H-3, H-5, H-3', H-5'), 7.77 (d, 4H, *J* = 6.3 Hz, H-2, H-6, H-2', H-6').MS (EI): 376.0[M⁺], 377[M+1], 188.0, 159.9[100%].IR (KBr) ν_{max} cm⁻¹: 3357.8 (NH), 3294.2 (sp²-CH), 1679.9 (C=O), 1600.8 (C=N), 1423.4 (C-H bending aromatic), 1249.8 (C-N).Results of all spectroscopic analysis confirmed the synthesis of oxalamide and its purity. ¹H-NMR, EI-MS and IR spectra of compound 3 are shown in (Fig. 2).**BIOLOGICAL SCREENING****Antioxidant Studies:**

Results for antioxidant screening of oxalamide derivatives are shown in Table 1. DPPH radical scavenging assay showed that three out of the seven synthesized oxalamide derivatives exhibited significant antioxidant activity which was close to the activity of standard drug (BHA). The half maximal inhibitory concentration for compound 2, 3, and 4 was 51.2, 55.4, and 47.3 μM , whereas, for BHA the IC₅₀ was 44.2 μM . The efficiency of compound 2, 3, and 4 was 86, 80, and 93% of the standard BHA. Four oxalamide derivatives were found to be inactive in this screening. These results revealed that substitution on oxalamide effected the antioxidant property of compound. This may be due to difference in electronegativity of the substituents. Compounds 5, 6, and 7 substituted with highly electronegative group showed no activity while compounds 2, 3 and 4 showed good activities. The unsubstituted oxalamides showed no antioxidant properties.

DPPH scavenging activity of the compounds synthesized in this study was also compared with some of the reported analogues like anilines, phenylenediamines, aminophenols, polyanilines and HCl doped anilines. It was reported that polyanilines doped with hydrochloric acid showed higher efficiency as compared to polyanilines while the later were more efficient than anilines (Gizdavic-Nikolaidis *et al.*, 2004). The IC₅₀ of the compounds synthesized in this study were bis(2,5-dimethylphenyl)oxalamide (51.2 μM), bis(3-bromophenyl)oxalamide (55.4 μM), and bis(4-bromophenyl)oxalamide (47.3 μM) which is much better when compared to *o*-phenylenediamine (590 μM), *p*-phenylenedamine (600 μM), 2-aminophenol (400 μM), 3-aminophenol (330 μM), and 4-aminophenol (300 μM) (Bendary *et al.*, 2013; Wright *et al.*, 2001). The high efficiency of diamines is attributed to the resonance stability of the product formed after radical scavenging (Wright *et al.*, 2001). The compounds synthesized in this study had higher resonance stability due to their high aromaticity which resulted in low IC₅₀ values. The compounds which didn't show significant antioxidant activity had electron withdrawing substitutions and their negative inductive effect would have affected their resonance stabilization.

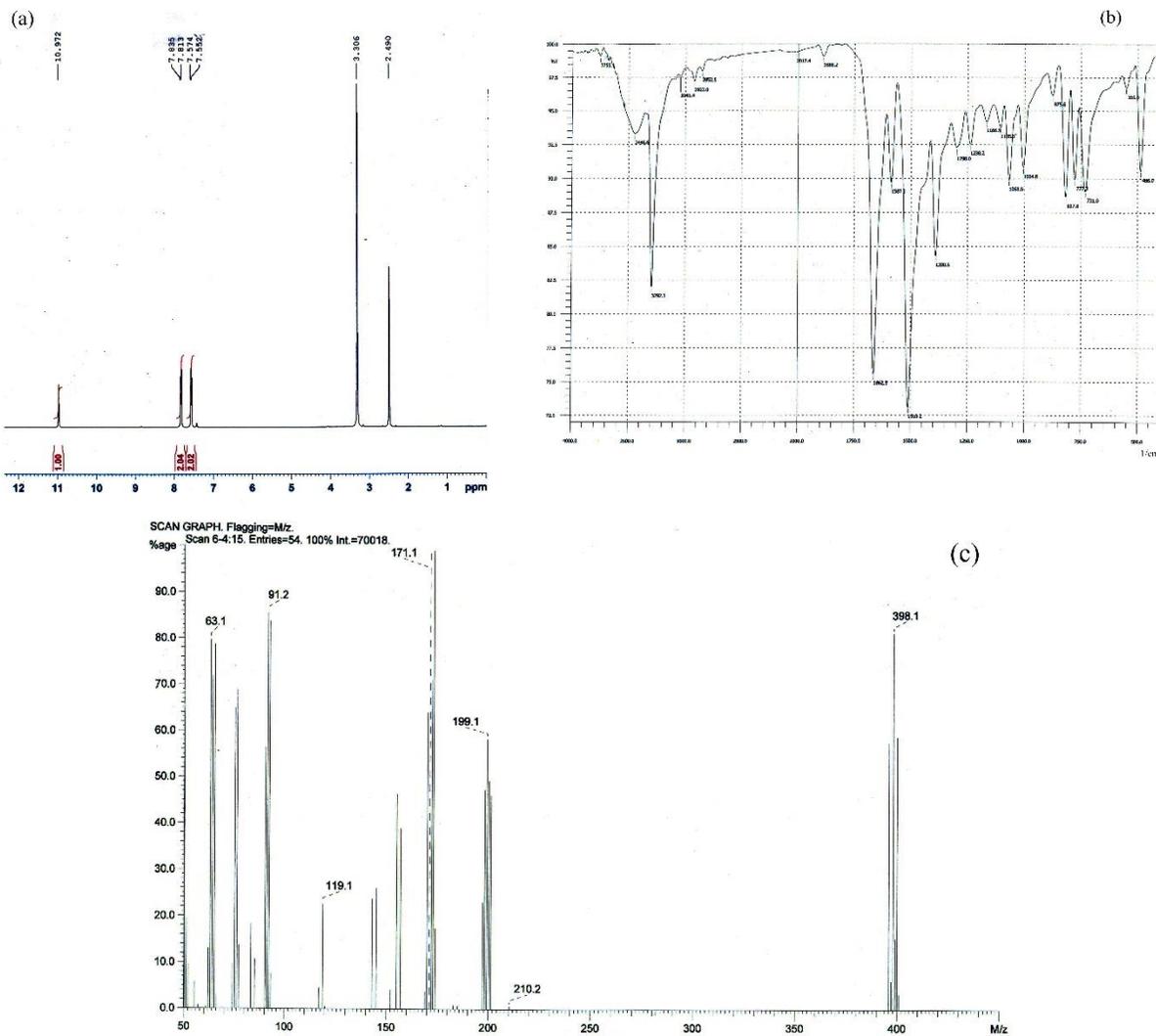


Fig. 2. Characterization of synthesized oxamides a) NMR Spectrum, b) IR Spectrum, and c) EI-MS spectrum.

Table 1. Antioxidant activity screening of synthesized oxalamides.

Sample ID	R	Antioxidant Activity IC ₅₀ (μM± SEM)
1	H	N.A.
2	2,5-dimethyl	51.2±0.34
3	3-Br	55.4 ± 0.38
4	4-Br	47.3 ± 0.24
5	3-Cl	N.A.
6	4-Cl	N.A.
7	4-CF ₃	N.A.
BHA (standard)	-	44.2 ± 0.04

N.A. = No Activity

Anticancer studies:

Anticancer activities of oxalamides are shown in Table 2. Oxalamide derivatives were found to be inactive against cancer cell lines and hence don't possess anticancer activity against beta lymphoma.

In this study a series of N,N-bis substituted oxalamide derivatives were synthesized by reacting amine with oxalyl chloride in the presence of triethylamine. Synthesized oxalamides were characterized using different spectroscopic techniques. The synthesized compounds were screened for antioxidant properties. It was observed that bis (2,5-dimethylphenyl)oxalamide, bis (3-bromophenyl)oxalamide, and bis (4-bromophenyl)oxalamide had good antioxidant properties. The compounds have potential to undergo further pharmacological studies. However, synthesized oxalamide derivatives did not show any anticancer properties.

Table 2. Anticancer activity studies for synthesized oxalamides.

Sample ID	R	Raji (Burkitt's Lymphoma)	DOHH ₂ (Follicular lymphoma)
1	H	N.A.	N.A.
2	2,5-dimethyl	N.A.	N.A.
3	4-Br	N.A.	N.A.
4	3-Br	N.A.	N.A.
5	4-Cl	N.A.	N.A.
6	3-Cl	N.A.	N.A.
7	4-CF ₃	N.A.	N.A.
(Standard) Imatinib	-	65.44 ± 3.76	53.10 ± 1.37
(Standard) Dasatinib	-	33.64 ± 1.9	47.08 ± 0.75

N.A. = No Activity

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REFERENCES

- Ali, S., S. Yasmeen, N. Afza, A. Malik, L. Iqbal, M. Lateef, N. Riaz and M. Ashraf (2009). Mutininside, new antioxidant phenolic glucoside from *Abutilon muticum*. *Journal of Asian Natural Products Research*, 11(5): 457-464.
- Aruoma, O.I. (1998). Free radicals, oxidative stress, and antioxidants in human health and disease. *Journal of the American oil chemists' society*, 75(2): 199-212.
- Balmus, I.M., A. Ciobica, A. Trifan and C. Stanciu (2016). The implications of oxidative stress and antioxidant therapies in inflammatory bowel disease: clinical aspects and animal models. *Saudi journal of gastroenterology*, 22(1): 3-17.
- Bendary, E., R.R. Francis, H.M.G. Ali, M.I. Sarwat and S. El Hady (2013). Antioxidant and structure-activity relationships (SARs) of some phenolic and anilines compounds. *Annals of Agricultural Sciences*, 58(2): 173-181.
- Dickinson, B.C. and C.J. Chang (2011). Chemistry and biology of reactive oxygen species in signaling or stress responses. *Nature chemical biology*, 7(8): 504.
- Fang, Y.-Z., S. Yang and G. Wu (2002). Free radicals, antioxidants, and nutrition. *Nutrition*, 18(10): 872-879.
- Fazal-ur-Rehman, S., A.A. Wasim, S. Iqbal, M.A. Khan, M. Lateef and L. Iqbal (2019). Synthesis, lipoygenase inhibition activity and molecular docking of oxamide derivative. *Pakistan Journal of Pharmaceutical Sciences*, 32(3(Suppl)): 1253-1259.
- Finkel, T. and N.J. Holbrook (2000). Oxidants, oxidative stress and the biology of ageing. *Nature*, 408(6809): 239.
- Giordano, S. and A. Petrelli (2008). From single-to multi-target drugs in cancer therapy: when aspecificity becomes an advantage. *Current medicinal chemistry*, 15(5): 422-432.
- Gizdavic-Nikolaidis, M., J. Travas-Sejdic, P.A. Kilmartin, G.A. Bowmaker and R.P. Cooney (2004). Evaluation of antioxidant activity of aniline and polyaniline. *Current Applied Physics*, 4(2): 343-346.
- Hanikoglu, A., H. Ozben, F. Hanikoglu and T. Ozben (2019). Hybrid compounds and oxidative stress induced apoptosis in cancer therapy. *Current medicinal chemistry*: Doi: 10.2174/0929867325666180719145819.

- Khalaf, N.A., A.K. Shakya, A. Al-Othman, Z. El-Agbar and H. Farah (2008). Antioxidant activity of some common plants. *Turkish Journal of Biology*, 32(1): 51-55.
- Lee, S.K., Z. Mbwambo, H. Chung, L. Luyengi, E. Gamez, R. Mehta, A. Kinghorn and J. Pezzuto (1998). Evaluation of the antioxidant potential of natural products. *Combinatorial Chemistry and High Throughput Screening*, 1(1): 35-46.
- Mahmoud, E.A., J. Sankaranarayanan, J.M. Morachis, G. Kim and A. Almutairi (2011). Inflammation responsive logic gate nanoparticles for the delivery of proteins. *Bioconjugate Chemistry*, 22(7): 1416-1421.
- Malki, F., A. Touati and S. Moulay (2017). Comparative study of antioxidant activity of some amides. *Journal of Analytical and Pharmaceutical Research*, 5(3): 1-5.
- Nepali, K., S. Sharma, M. Sharma, P. Bedi and K. Dhar (2014). Rational approaches, design strategies, structure activity relationship and mechanistic insights for anticancer hybrids. *European journal of medicinal chemistry*, 77: 422-487.
- O'brien, J., I. Wilson, T. Orton and F. Pognan (2000). Investigation of the Alamar Blue (resazurin) fluorescent dye for the assessment of mammalian cell cytotoxicity. *European Journal of Biochemistry*, 267(17): 5421-5426.
- Rajput, P. and A. Sharma (2018). Synthesis and biological importance of amide analogues. *Journal of Pharmacology and Medicinal Chemistry*, 2(1): 22-31.
- Raziq, N., M. Saeed, M.S. Ali, S. Zafar, M. Shahid and M. Lateef (2017). A new glycosidic antioxidant from *Ranunculus muricatus* L.(Ranunculaceae) exhibited lipoygenase and xanthine oxidase inhibition properties. *Natural product research*, 31(11): 1251-1257.
- Siddiq, F., I. Fatima, A. Malik, N. Afza, L. Iqbal, M. Lateef, S. Hameed and S.W. Khan (2012). Biologically active bergenin derivatives from *Bergenia stracheyi*. *Chemistry and biodiversity*, 9(1): 91-98.
- Sostres, C., C.J. Gargallo, M.T. Arroyo and A. Lanas (2010). Adverse effects of non-steroidal anti-inflammatory drugs (NSAIDs, aspirin and coxibs) on upper gastrointestinal tract. *Best practice and research Clinical gastroenterology*, 24(2): 121-132.
- Statistics, W.C. (2019) *Worldwide cancer statistics*, 2019. Available online: <https://www.cancerresearchuk.org/health-professional/cancer-statistics/worldwide-cancer> Accessed: June 08, 2019.
- Sznarkowska, A., A. Kostecka, K. Meller and K.P. Bielawski (2017). Inhibition of cancer antioxidant defense by natural compounds. *Oncotarget*, 8(9): 15996-16016.
- Tian, T., Z. Wang and J. Zhang (2017). Pathomechanisms of oxidative stress in inflammatory bowel disease and potential antioxidant therapies. *Oxidative medicine and cellular longevity*, 2017: 1-18.
- Waris, G. and H. Ahsan (2006). Reactive oxygen species: role in the development of cancer and various chronic conditions. *Journal of carcinogenesis*, 5: 14.
- Wright, J.S., E.R. Johnson and G.A. DiLabio (2001). Predicting the Activity of Phenolic Antioxidants: Theoretical Method, Analysis of Substituent Effects, and Application to Major Families of Antioxidants. *Journal of the American Chemical Society*, 123(6): 1173-1183.
- Zuo, T., M. Zhu and W. Xu (2016). Roles of oxidative stress in polycystic ovary syndrome and cancers. *Oxidative medicine and cellular longevity*, 2016: 1-14.

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