

EFFECTS OF PHOTOSYNTHETIC PIGMENTS OF *VIGNA RADIATA* L. & *VIGNA UNGUICULATA* L. CULTIVATED IN LEAD CONTAMINATED SOIL AMENDED WITH CHICKEN MANURE AND MINERAL FERTILIZER

Neelofer Hamid and Faiza Jawaid

Department of Botany, University of Karachi, Karachi-75270, Pakistan

ABSTRACT

Vigna radiata and *Vigna unguiculata* plants were grown in lead (Pb) contaminated soil which is amended with different concentrations of organic fertilizer (chicken manure) and mineral fertilizer (NPK). In present investigation polluted soil was from industrial area of Karachi with elevated Pb content. The concentration of photosynthetic pigments that is chlorophyll "a", "b" and carotenoids in leaf of developing plants were monitored every one week interval up to the period of 4 weeks. Application of both fertilizer improve the concentration of chlorophyll "a", "b" and carotenoids, with time. In control of both *Vigna radiata* and *Vigna unguiculata*, chlorophyll "a", "b" and carotenoids concentration was reduces, such reduction in the concentration of leaf pigment due to the toxic effect of metals (Pb).

Key words: Carotenoides, Chlorophyll "a", Chlorophyll "b", Chicken manure, Mineral fertilizer, Metal polluted soil, *Vigna radiata*, *Vigna unguiculata*.

INTRODUCTION

Heavy metal contamination of soil, water and air has caused severe environmental hazard in the biosphere due to rapid industrialization and urbanization (Xiong, 1998). Heavy metals are a dangerous group of soil pollutants, the contamination by heavy metals causes a serious problem because they cannot be naturally degraded like organic pollutants and they accumulate in food chain (Smejkalova *et al.*, 2003). Among the existing heavy metal pollutants, lead (Pb) is the major contaminant of the soil (Gratao *et al.*, 2005; Romero *et al.*, 2006). Significant increases in the Pb content of cultivated soils have been observed near industrial areas (De Abreu *et al.*, 1998; Mishra and Choudhuri, 1999; Sharma and Dubey, 2005).

Contamination of soil by lead enhances plant uptake and causing their accumulation in different plants organ (Ernst, 1996; Zayed *et al.*, 1998; Gimmler *et al.*, 2002; Chaturvedi, 2004; Gaspar *et al.*, 2005). Lead and other heavy metal have no known biological functions (Antosiewicz, 1992; Xiong, 1998 Nicholls and Mal, 2003). In plants, the exposure to toxic concentration of lead causes oxidative stress, growth inhibition, chlorosis, browning of root tips, disturbance of photosynthesis, reduction of water and nutrient uptake and inhibition of enzymatic activities of various enzyme (Sharma and Dubey, 2005).

Soil improvement is the most important step for successful establishment of plants in heavy metal polluted soil. Addition of organic manures (Cow manure, Poultry manure and Pig manure) are well established to be involved in fertilization of plants due to its beneficial effect on physical, chemical and biological characteristics of the soil, this in turns, influences growth and increase production of plant (Scialdone *et al.*, 1980; Wong and Lau, 1985; Ye *et al.*, 1999).

The main objective of this investigation was to study the effect of heavy metal polluted soil which is amended with different concentration of chicken manure and mineral fertilizer (NPK) by moitoring the concentration of photosynthetic pigment Chlorophyll "a", "b" and carotenoides in leaves of *Vigna radiata* and *Vigna unguiculata*.

MATERIAL AND METHODS

A pot Experiment was conducted with two different edible beans *Vigna unguiculata* and *Vigna radiata*. Healthy seeds of each species were selected and sterilized with 0.1% Mercuric chloride solution for 5 minutes followed by rinsing with tap and distilled water. Seeds of *Vigna unguiculata* and *Vigna radiata* were sown in plastic begs each containing 300gm sterilized soil. After germination 12 days old seedlings were transplanted in plastic pots (12cm diameter and 15cm height) contains lead polluted soil (polluted soil collected from industrial area of Karachi), Soil was mixed with three levels of chicken manure and mineral fertilizer (50, 75 and 100 g/Kg). Soil without NPK and chicken manure served as the control. Regular watering was carried, plants watered according to requirement throughout experimental period.

Samples of soil, mineral fertilizer and chicken manure were analyzed for total lead by atomic absorption spectrophotometer following the procedure use by the Swiss national soil monitoring network (Nowack *et al.* 2001).

The leaf samples from both control and treated plants were collected in early hours of the morning and were kept in labeled sample bags. The plants samples were analyzed for following leaf pigments.

Chlorophyll ("a" and "b") and carotenoids were extracted from the leaves and estimated by the method of Maclachlan and Zalik (1963). Data were statistically analyzed by "SPSS" program and "SIGMA PLOT" program was used for graphic presentation of the data.

RESULTS

The polluted soil contains higher lead (Pb) content (245.260µg/mg) as compare to chicken manure (53.760µg/mg) and mineral fertilizer (32.910µg/mg).

The data in table 1 illustrate that chicken manure and mineral fertilizer treatment has positive effect on the amount of chlorophyll "a" of both *Vigna radiata* and *Vigna unguiculata* as compared to control. In all chicken manure treatment of *Vigna radiata* and *Vigna unguiculata* significant ($P<0.01$) increase in the amount of chlorophyll "a" was observed through out the experiment (Fig. 1a & 1b). In *Vigna radiata* amount of chlorophyll "a" increases with the increased intensity of applied chicken manure, however, uneven increasing trend was observed in *Vigna unguiculata*. In 1st week of investigation decrease in the amount of chlorophyll "a" of *Vigna radiata* was observed after treatment with different concentration of mineral fertilizer as compare to control (Fig. 1c), afterward significant ($P<0.05$) increase was observed till the end of investigation period, on the other hand mineral fertilizer treatment of *Vigna unguiculata* showed significant ($P<0.01$) increase in the amount of chlorophyll "a" (Fig. 1d), which is continued till the end of experiment.

From Table 2 it is obvious that Significant increase in amount of chlorophyll "b" of *Vigna radiata* and *Vigna unguiculata* ($P<0.01$) was observed throughout the investigational period. The amount of chlorophyll "b" of *Vigna unguiculata* and *Vigna radiata* showed a sharp increase with the increase in the concentration of chicken manure supplied. In both species the high increase in chlorophyll "b" content was observed in chicken manure treatments as compare to control (Fig. 2a & 2b). The chlorophyll "b" content of *Vigna unguiculata* treated with mineral fertilizer elucidates increase which was higher than control (Fig. 2c) and *Vigna radiata* demonstrate higher increase in control as compare to treatment (Fig. 2d).

The result obtained for the effect of different concentration of chicken manure treatment on carotenoid amount of *Vigna radiata* and *Vigna unguiculata* were statistically significant ($P<0.01$). The carotenoid content of *Vigna radiata* treated with different concentration of chicken manure and mineral fertilizer showed a maximum increase in 50 g/Kg fertilizer treatment (Fig. 3a & 3c) and all fertilizer (chicken manure and mineral fertilizer) treatment of *Vigna unguiculata* display increase in all treatment as compare to control, the result obtained were uneven (Fig. 3b & 3d).

Table 1. Weekly changes in the amount of Chlorophyll "a" (mg/g) of *Vigna unguiculata* and *Vigna radiata* grown on lead contaminated soil under different fertilizer treatments.

species	Treatment	Fertilizer applied (mg/Kg)	Weeks			
			1 st	2 nd	3 rd	4 th
<i>Vigna radiata</i>	Control	-	1.233±0.025	1.057±0.063	1.033±0.033	0.964±0.061
	Chicken manure**	50	1.259±0.057	1.535±0.033	1.980±0.081	2.105±0.017
		75	1.345±0.061	1.629±0.038	2.016±0.023	2.332±0.067
		100	1.389±0.033	1.826±0.086	2.236±0.063	2.554±0.025
	Mineral fertilizer*	50	1.133±0.051	1.232±0.086	1.328±0.022	1.447±0.089
		75	1.065±0.076	1.156±0.089	1.196±0.061	1.249±0.033
		100	0.941±0.061	0.978±0.036	1.013±0.058	0.939±0.081
	Control	-	1.351±0.061	1.232±0.066	1.061±0.041	1.057±0.026
<i>Vigna unguiculata</i>	Chicken manure**	50	1.488±0.028	1.735±0.091	2.343±0.021	2.736±0.041
		75	1.561±0.032	1.721±0.026	2.372±0.051	2.730±0.066
		100	1.483±0.023	1.622±0.061	2.242±0.091	2.526±0.033
	Mineral fertilizer**	50	1.434±0.081	1.613±0.099	2.169±0.061	2.133±0.057
		75	1.342±0.046	1.611±0.044	2.128±0.066	2.032±0.066
		100	1.300±0.021	1.572±0.063	1.776±0.021	1.842±0.059
	Control	-	1.351±0.061	1.232±0.066	1.061±0.041	1.057±0.026

Significance level = * $P<0.05$ and ** $P<0.01$

Table 2. Weekly changes in the amount of Chlorophyll “b” (mg/g) of *Vigna unguiculata* and *Vigna radiata* grown on lead contaminated soil under different fertilizer treatments.

species	Treatment	Fertilizer applied (mg/Kg)	Weeks			
			1 st	2 nd	3 rd	4 th
<i>Vigna radiata</i>	Control	-	0.428±0.061	0.449±0.041	0.666±0.022	0.693±0.036
		50	0.762±0.073	0.833±0.029	0.809±0.029	0.959±0.055
	Chicken manure**	75	0.832±0.061	0.868±0.055	0.930±0.023	0.986±0.023
		100	0.916±0.061	0.936±0.026	0.976±0.026	1.059±0.036
	Mineral fertilizer**	50	0.435±0.033	0.458±0.077	0.481±0.026	0.495±0.033
		75	0.414±0.033	0.422±0.066	0.431±0.081	0.476±0.026
<i>Vigna unguiculata</i>	Control	-	0.705±0.026	0.762±0.022	0.438±0.038	0.336±0.038
		50	0.939±0.022	0.978±0.026	1.145±0.056	1.166±0.028
	Chicken manure**	75	0.932±0.042	0.993±0.078	1.240±0.086	1.246±0.033
		100	0.922±0.066	0.976±0.047	1.045±0.054	1.133±0.086
	Mineral fertilizer**	50	0.867±0.081	0.932±0.091	0.954±0.026	0.966±0.041
		75	0.852±0.072	0.933±0.043	0.973±0.026	0.976±0.036
		100	0.872±0.088	0.941±0.033	0.977±0.033	0.989±0.089

Significance level = **P<0.01

Table 3. Weekly changes in the amount of Carotenoid (mg/g) of *Vigna unguiculata* and *Vigna radiata* grown on lead contaminated soil under different fertilizer treatments.

species	Treatment	Fertilizer applied (mg/Kg)	Weeks			
			1 st	2 nd	3 rd	4 th
<i>Vigna radiata</i>	Control	-	0.238±0.036	0.468±0.051	0.423±0.044	0.418±0.038
		50	0.386±0.063	0.798±0.036	0.924±0.051	0.971±0.061
	Chicken manure**	75	0.378±0.021	0.766±0.071	0.826±0.033	0.884±0.088
		100	0.344±0.026	0.742±0.074	0.800±0.023	0.816±0.058
	Mineral fertilizer**	50	0.366±0.033	0.671±0.033	0.773±0.074	0.835±0.066
		75	0.325±0.029	0.647±0.034	0.713±0.043	0.806±0.052
<i>Vigna unguiculata</i>	Control	-	0.403±0.022	0.423±0.042	0.451±0.022	0.413±0.063
		50	0.669±0.061	1.023±0.026	1.131±0.046	1.326±0.033
	Chicken manure**	75	0.852±0.044	1.121±0.066	1.286±0.072	1.386±0.046
		100	0.650±0.033	0.819±0.022	1.026±0.036	1.079±0.066
	Mineral fertilizer**	50	0.654±0.022	0.886±0.032	1.084±0.066	1.112±0.033
		75	0.642±0.042	0.812±0.066	0.973±0.054	1.066±0.042
		100	0.526±0.046	0.732±0.081	0.788±0.063	0.986±0.033

Significance level = **P<0.01

DISCUSSION

Lead concentration is increasing rapidly in the environment due to enlarged use of its sources by human society (Haider *et al.*, 2006), increasing lead concentration has become a global problem because of its deteriorious influence not only on plant growth (yield and quality) and environmental quality, but also on the health of human beings, therefore, much effort has been made to decontaminate the polluted soil by using chemical, physical or biological methods (Salt *et al.*, 1998).

Chlorophyll is the central part of the energy manifestation of every green plant system and therefore, any significance alteration in their levels is likely to cause marked effects on the entire metabolism of plants (Knudson *et al.*, 1977). Plant pigments are of tremendous significance in the biosphere, for the reason that the chlorophyll is earth's most important organic molecules as they are necessary for photosynthesis (Blackburn, 2007). Carotenoids are as well essential for plant and mammal survival through their photosynthetic and nutritional functions (Blackburn, 2007).

In the present investigation, chlorophyll “a”, chlorophyll “b” and carotenoid content of control samples was more affected as compared to fertilizer treatment. Our work is supported by the work of Sinha and Gupta (2005)

who suggested that chlorophyll contents of *Sesbania cannabina* increased with increase in the Fly Ash amendment. Amendments other than fly ash enhance the accumulation of metals (Fe, Mn, Zn, Cu, Pb and Ni).

Lead (Pb) inhibits the chlorophyll synthesis by impairing uptake of essential elements such as Mg and Fe by plants (Burzynski, 1987). Jaleel *et al* (2009) suggested that, impaired chlorophyll development by heavy metals may be due to the interference to proteins (enzymatic proteins) responsible for chlorophyll biosynthesis. Chlorophyll "a" and Chlorophyll "b" content was decreased in two mustard cultivars under heavy metal treatment (Mobin and Khan, 2006). Heavy metal can accumulate in higher levels in the aerial part preferentially in the chloroplast and disturb the enzyme of chlorophyll biosynthesis by interacting with SH group of chloroplast synthesizing enzyme (Tawfik, 2008). The Key enzyme of chlorophyll biosynthesis, d-amino laevulinate dehydrogenase, is strongly inhibited by Pb ions (Prasad and Prasad 1987). An enhancement of chlorophyll degradation occurs in Pb treated plants due to increased chlorophyllase activity (Drazkiewicz, 1994).

Carotenoids are a non enzymatic antioxidant, playing vital role in the protection of plant from the adverse impact of reactive oxygen species (Tawfik, 2008). Likewise, Sengar *et al.* suggested that Carotenoids protect chlorophyll from photooxidative destruction and therefore, a reduction in carotenoid could have a serious consequence on chlorophyll pigments, although the total carotenoid content was also slightly reduced by Pb, there was no correlation between Pb concentration and the fall in carotenoid content (Sengar *et al.*, 2008).

CONCLUSION

Thus present study clearly indicated that the application of chicken manure and mineral fertilizer was the most effective for stimulating the growth and metabolic processes of plants in lead contaminated soils.

REFERENCES

- Antosiewicz, D.M. (1992). Adaptation of plants to an environment polluted with heavy metals. *Byul. Izobr.* 61: 281–299.
- Blackburn, G.A. (2007). Hyper spectral remote sensing of plant pigments. *J Exp. Bot.* 58(4): 855-867.
- Burzynski, M. (1987). The influence of lead and cadmium absorption and distribution of potassium, calcium, magnesium and iron in cucumber seedling. *Acta Physiol Plant* 9:229-238.
- Chaturvedi, I. (2004). Phytotoxicity of cadmium and its effect on two genotypes of *Brassica juncea* L. *Emir J Agric Sci.* 16 (2): 01-08.
- De Abreu, C.A., M.F De Abreu and J.C De Andrade (1998). Distribution of lead in the soil profile evaluated by DTPA and Mehlich-3 solutions. *Bragantia* 57:185-192.
- Drazkiewicz, M. (1994). Chlorophyll-occurrence, functions, mechanism of action, effects of internal and external factors. *Photosynthetica* 30:321-331.
- Ernst, W.H.O. (1996). Bioavailability of heavy metals and decontamination of soil by plants. *Appl Geochem* 11: 163-167.
- Gaspar, G.M., P. Mathe, L. Szabo, B. Orgovanyi, N. Uzingier and A. Anton (2005). After- effect of heavy metal pollution in a brown forest soil. *Acta Biol Szegediensis* 49(1-2):71-72.
- Gimmler, H., J. Carandang, A. Boots, E. Reisberg and M. Woiatke (2002). Heavy metal content and distribution within a woody Plant during and after seven years continuous growth on municipal solid waste bottom slag rich in heavy metals. *J Appl Bot* 76:203-217.
- Gratao, P.L., M.N.V Prasad, P.F Cardoso, P.J Lea and R.A Azevedo (2005). Phytoremediation: green technology for the clean up of toxic metals in the environment. *Braz J Plant Physiol.* 17:53-64
- Sengar, R.S., M. Gautam, S.K Garg, R Chaudary and K. Sengar (2008). Effect of lead on seed germination , seedling growth , chlorophyll content and Nitrate reductase activity of Mung beans (*Vigna radiata*). *Rec. J. Phytochemistry* 2(2): 61-68.
- Haider, S., S. Kanwal, F. uddin and R. Azmat (2006). Phytotoxicity of Pb: II. Changes in chlorophyll absorption spectrum due to toxic metal Pb stress on *Phaseolus mungo* and *Lens culinaris*. *Pak. J Biol. Sci.* 9(11): 2062-2068.
- Jaleel, A.C., K. JayaKumar, Z. Chang-Xig and M.M Azooz (2009). Antioxidant potentials protect *Vigna radiata* (L.) Wilczek plants from soil Cobalt stress and improve growth and pigment composition. *Plants Omics Journal* 2(3): 120-126.
- Knudson, L.L., T.W Tibbitts and G.E Edwards (1977). Measurement of ozone injury by determination of leaf chlorophyll concentration. *Plant physiol.*, 60: 606-608.
- MacLachlan, S. and S. Zalik (1963). Extraction and estimation of chlorophyll. *Can J Bot.*, 41: 1053.

- Mishra, A. and M.A Choudhuri (1999). Monitoring of Phytotoxicity of Lead and Mercury from germination and early seedling growth indices in two rice cultivars. *Water Air Soil Pollu.*, 114:339-346.
- Mobin, M. and N.A Khan (2006). Photosynthetic activity, pigment composition and antioxidative response of two mustard (*Brassica Juncea*) cultivars differing in photosynthetic capacity subjected to cadmium stress. *J Plant Physiol* 164: 601-610.
- Nicholls, A.M. and T.K Mal (2003). Effects of lead and copper exposure on growth of an invasive weed, *Lythrum salicaria* L. *Ohio J Sci* 103(5):129-133.
- Nowack, B., J.M Obrecht, M. Schluep, R. Schulin, W. Hansmann and V. Koppel (2001). Elevated Lead and Zinc contents in remote alpine soils of the swiss national park. *J Environ Qual.*, 30: 919-926.
- Prasad, D.D.K and A.R.K Prasad (1987). Altered d-aminolaevulinic acid metabolism by lead and mercury in germinating seedlings of Bajra (*Pennisetum typhoideum*). *J Plant Physiol.*, 127:241-249.
- Ramadan, M.A.E. and E. Al-Ashkar (2007). The effect of different fertilizers on the heavy metals in soil and tomato plant. *Aust. J. basic & Appl. Sci.*, 1(3): 300-306.
- Ramadan, M.A.E., S.M. Adam and Z.F. Fawzy (2007). The distribution of heavy metals in soil and squash organs under different rates from poultry manures and biofertilizer. *J. Appl. Sci. Res.*, 3(7): 581-586.
- Romerio, S., A.M.M.A Lagoa, P.R Furlani, C.A de Abreu, M.F de Abreu and N.M Erismann (2006). Lead uptake and tolerance of *Ricinus communis* L. *Braz J Plant Physiol.*, 18(4):483-489.
- Salt, D.E., R.D Smith and I Raskin (1998). Phytoremediation. *Annu Rev Plant Physiol.*, 49: 643-668.
- Scialdone, D., D. Scogamiglio and A.U Ramunni (1980). The short and medium term effects of organic amendments on lead availability. *Water Air Soil Pollut.*, 13: 267-274.
- Sharma, P and R.S Dubey (2005). Lead toxicity in plants. *Braz. J. Plant Physiol.* 17(1): 35-52.
- Sinha, S. and A.K Gupta (2005). Translocation of metals from fly ash amended soil in the plant of *Sesbania cannabina* L. Ritz: Effect on antioxidants. *Chemosphere*, 61: 1204-1214.
- Smejkalova, M., O. Mikanova and L. Boruvka (2003). Effects of heavy metal concentrations on biological activity of soil micro-organisms, *J. Plant Soil Environ*, 49(7): 321-326.
- Tawfik, K.M. (2008). A monitory field study at El Seaf- Helwan Faba bean farms irrigated by industrial waste water and polluted water with sewage. *J Appl. Sci. Rec.*, 4(5): 492-499.
- Xiong, Z.T. (1998). Lead uptake and effects on seed germination and plant growth of a Pb accumulator, *Brassica perkinsensis* Rupr. *Bull Environ Contam Toxicol.*, 60:285-291.
- Wong, M.H. and W.M Lau (1985). The effects of applications of phosphate, lime, EDTA, refuse compost and pig manure on the Pb contents of crops. *Agric Wastes*, 12: 61-75.
- Ye, Z.H, J.W.C Wong, M.H Wong, C.Y Lan and A.J.M Baker (1999). Lime and pig manure as ameliorants for revegetating lead/zinc mine tailings: a greenhouse study. *Bioresour Technol.*, 69: 35-43.
- Zayed, A., C.M Lytle, J.H Qian and N. Terry (1998). Chromium accumulation, translocation and chemical speciation in vegetable crops. *Planta*, 206: 293-299.

(Accepted for publication May, 2009)