

## INTRODUCTION TO ANTIOXIDANT ENZYMES AND THEIR IMPLICATIONS IN PATHOPHYSIOLOGIC PROCESSES: A REVIEW ARTICLE

S. Zare<sup>3</sup>, M. Shokrzadeh<sup>2</sup> and A.G. Ebadi<sup>1</sup>

<sup>1</sup>Department of Biology, Faculty of Sciences, Urmia University, Urmia, Iran

<sup>2</sup>Department of Toxicology, Mazandaran University of Medical Sciences, Sari, Iran

<sup>3</sup>Department of Biology, Islamic Azad University of Sari Sari- 48164-194, Iran

### ABSTRACT

Aerobic organisms possess antioxidant defense systems that deal with reactive oxygen species (ROS) produced as a consequence of aerobic respiration. Reactive oxygen is related to both, the arrest of growth and the start of cell differentiation. Low concentrations of reactive oxygen intermediates may be beneficial or even indispensable in processes such as intracellular messaging and defense against microorganisms, but higher amounts of active oxygen may be harmful to cells and organisms. A wide array of non-enzymatic and enzymatic antioxidant defenses exists, including superoxide dismutase (SOD), glutathione peroxidase (GPX) and catalase (CAT). We describe their main characteristics and how these antioxidant enzymes work together against active oxygen. Small deviations from their physiological values may have a dramatic effect on the resistance of cells to oxidative damage to lipids, proteins and DNA. Consequently, toxic oxygen plays a role in aging process as well as in a number of human diseases that we list in this review.

**Key-words :** Antioxidant enzymes, aerobic organisms, ROS, GPX, CAT

### INTRODUCTION

Small amounts of reactive oxygen species (ROS), as hydroxyl radicals (HO<sup>•</sup>), superoxide anions (O<sub>2</sub><sup>•-</sup>) and hydrogen peroxide (H<sub>2</sub>O<sub>2</sub>), are constantly generated in aerobic organisms in response to both external and internal stimuli (Lee *et al.*, 1998). Low levels of ROS may be indispensable in a plethora of processes, including intracellular messaging (Schulze-Osthoff *et al.*, 1997) leading among others to proliferation or apoptosis (Vogt *et al.*, 1998), immunity (Sun *et al.*, 1998), and defense against micro-organisms (Lee *et al.*, 1998). In contrast, high doses and/or inadequate removal of ROS result in oxidative stress, which may cause severe metabolic malfunctions and damage to biological macromolecules (Lledias *et al.*, 1998).

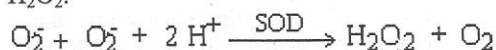
The prevention of lipid peroxidation is an essential process in all the aerobic organisms, as lipid peroxidation products can cause DNA damage. Increased lipid peroxidation and decreased antioxidant protection frequently occurs (Tampo *et al.*, 1998): epoxides may spontaneously react with nucleophilic centers in the cell and thereby covalently bind to DNA, RNA and protein (Fig. 1). Such a reaction may lead to cytotoxicity, allergy, mutagenicity and/or carcinogenicity, depending of the properties of the epoxide in question. Moreover, oxidative events may play an important role in the mechanism of action of ether lipids, and oxidizability may contribute to cellular drug sensitivity (Wagner *et al.*, 1998).

A wide array of enzymatic and non-enzymatic antioxidant defenses exist, including superoxide dismutase (SOD), glutathione peroxidase (GPX), catalase (CAT), glutathione (GSH), beta-carotene, vitamin A ascorbic acid (vitamin C) and alpha-tocopherol (vitamin E) (Mataix *et al.*, 1998). There is a interrelationship between both, the activities, and the intracellular levels of these metabolites, protecting themselves from oxygen toxicity (Grazioli *et al.*, 1998).

### I-ANTIOXIDANT ENZYMES

#### I.1. Superoxide dismutase

Superoxide dismutase (EC 1.15.1.1) destroys the free radical superoxide by converting it to peroxide that can in turn be destroyed by CAT or GPX reactions. SOD converts the highly reactive superoxide radical to the less reactive H<sub>2</sub>O<sub>2</sub>.



Another function of superoxide dismutase is to protect dehydratases (dihydroxy acid dehydratase, aconitase, 6-phosphogluconate dehydratase and fumarases A and B) against inactivation by the free radical superoxide (Benov and Fridovich, 1998).

Four classes of SOD have been identified, containing either a dinuclear Cu, Zn or mononuclear Fe, Mn or Ni cofactors (Whittaker and Whittaker, 1998). Fe-SODs and Mn-SODs show homology and possess identical metal

chelating residues at the active site, sharing substantial sequence and three dimensional structural homology, while the other superoxide dismutases are structurally unrelated. In humans, there are three forms of SOD: cytosolic Cu, Zn-SOD, mitochondrial Mn-SOD, and extracellular-SOD (EC-SOD) (Majima *et al.*, 1998). SOD catalyses the dismutation of  $O_2^-$  by successive oxidation and reduction of the transition metal ion at the active site in a Ping Pong type mechanism with remarkably high reaction rates (Hsieh *et al.*, 1998).

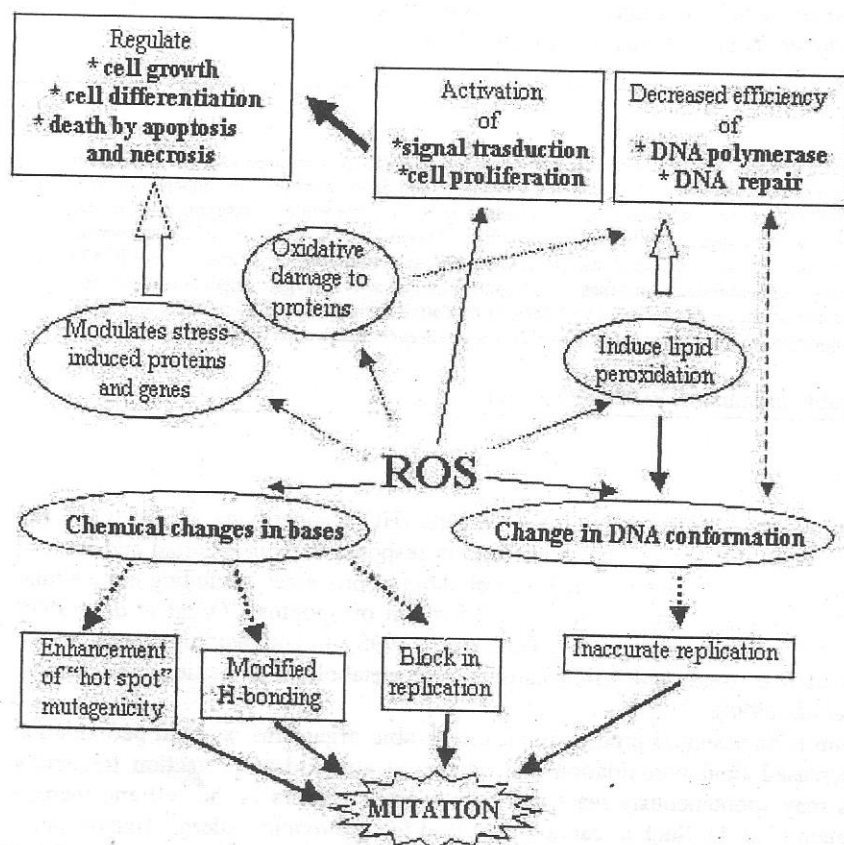


Fig.1. ROS, oxidative damage and human diseases. Interrelationship between the effect of imbalance in the reactive oxygen species (ROS) and their consequences on the cellular growth and the cellular function (top); and between ROS imbalance and the mechanisms and pathways from oxidative damage to mutation (down).

### 1.1.1 Manganese superoxide dismutase

Mn-SOD is a homotetramer (96 kDa) containing one manganese atom per subunit that cycles from Mn (III) to Mn (II) and back to Mn (III) during the two step dismutation of superoxide. The respiratory chain in mitochondria is a major source of oxygen radicals. Mn-SOD is a nuclear-encoded primary antioxidant enzyme that functions to remove these superoxide radical (Guan *et al.*, 1998). The biological importance of Mn-SOD is demonstrated among others by the following (Majima *et al.*, 1998): 1) inactivation of Mn-SOD genes in *E. coli* increases mutation frequency when grown under aerobic conditions. 2) Elimination of the gene in *Saccharomyces cerevisiae* increases its sensitivity to oxygen. 3) Lack of expression in Mn-SOD knock-out mice results in dilated cardiomyopathy and neonatal. 4) Tumor necrosis factor (TNF) selectively induces Mn-SOD, but not Cu, Zn-SOD, CAT or GPX mRNA in various mouse tissues and cultured cells. 5) Transfection of Mn-SOD cDNA into cultured cells rendered the cells resistant to paraquat, TNF and adriamycin-induced cytotoxicity, and radiation induced-neoplastic transformation. 6) Expression of human Mn-SOD genes in transgenic mice protects against oxygen-induced pulmonary injury and adriamycin-induced cardiac toxicity. Thus, the expression of Mn-SOD is essential for the survival of aerobic life and the development of cellular resistance to oxygen radical-mediated toxicity.

### 1.1.2. Copper,zinc superoxide dismutase

Cu, Zn-SOD (SOD-1) are another class of enzyme conserved throughout evolution, which usually have two identical subunits of about 32 kDa, each containing a metal cluster, the active site, constituted by a copper and a zinc atom bridged by a common ligand: His 61 (Banci *et al.*, 1998).

Whereas Mn-SOD was found in all tumors, and the ratio between the activities of Cu, Zn-SOD and Mn-SOD was not different from that of the normal tissues, tumors possess less Cu, Zn-SOD than did the more metabolically active tissues (Westman and Marklund, 1981). Cu, Zn-SOD is believed to play a major role in the first line of antioxidant defense by catalyzing the dismutation of superoxide anion radicals, to form hydrogen peroxide and molecular oxygen. Mice lacking this enzyme exhibited a pronounced susceptibility to paraquat toxicity. Most surprisingly, female homozygous knock-out mice showed a markedly reduced fertility compared with that of wild-type and heterozygous knock-out mice. They exhibited a marked increase in embryonic lethality. These data suggest a role of oxygen free radicals in causing abnormality of female reproduction in mammals (Ho *et al.*, 1998). Other recent reports involving SOD knock-outs have revealed that Mn-SOD is essential for life whereas Cu, Zn-SOD is not. Cu, Zn-SOD knock-out mice appear normal and exhibit differences only after traumatic injury, whereas Mn-SOD knockouts do not survive past 3 weeks of age (Reaume *et al.*, 1998).

### I.1.3. Extracellular superoxide dismutase

EC-SOD is a secretory, tetrameric, copper and zinc containing glycoprotein (with a high affinity for certain glycosaminoglycans such as heparin and heparan sulfate) found in the interstitial spaces of tissues and also in extracellular fluids, accounting for the majority of the SOD activity of plasma, lymph, and synovial fluid (Adachi and Wang, 1998). EC-SOD, is not induced by its substrate or other oxidants (xanthine oxidase plus hypoxanthine, paraquat, pyrogallol, alpha-naphthoflavone, hydroquinone, catechol,  $\text{Fe}^{2+}$  ions,  $\text{Cu}^{2+}$  ions, buthionine sulfoximine, diethylmaleate, t-butyl hydroperoxide, cumene hydroperoxide, selenite, citiolone and high oxygen partial pressure) and its regulation in mammalian tissues primarily occurs in a manner coordinated by cytokines, rather than as a response of individual cells to oxidants (Buschfort *et al.*, 1997).

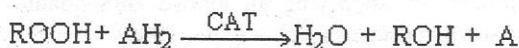
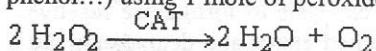
### I.1.4. Nickel superoxide dismutase

Ni-SOD has been purified from the cytosolic fraction of *Streptomyces sp.* and *Streptomyces coelicolor*. It is composed of four identical subunits of 13.4 kDa, stable at pH 4.0-8.0, and up to 70 Celsius degrees. It is inhibited by cyanide and  $\text{H}_2\text{O}_2$  but little inhibited by azide. Amino acid composition is different from iron, manganese and zinc-copper SODs. The apoenzyme, lacking in nickel, had no ability to mediate the conversion of superoxide anion to hydrogen peroxide, strongly indicating that  $\text{Ni}^{\text{III}}$  plays a main role in the activity (Youn *et al.*, 1996).

### I.2. Catalase

Catalase (EC 1.11.1.6) is a tetrameric haemin-enzyme consisting of 4 identical tetrahedrally arranged subunits of 60 kDa. Therefore, it contains 4 ferriprotoporphyrin groups per molecule, and its molecular mass is about 240 kDa. Catalase is one of the most efficient enzymes known. It is so efficient that it cannot be saturated by  $\text{H}_2\text{O}_2$  at any concentration (Lledias *et al.*, 1998).

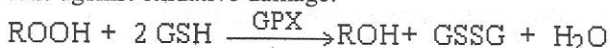
CAT reacts with  $\text{H}_2\text{O}_2$  to form water and molecular oxygen; and with H donors (methanol, ethanol, formic acid, phenol...) using 1 mole of peroxide in a kind of peroxidase activity:



$\text{H}_2\text{O}_2$  is enzymically catabolized in aerobic organism by catalase and several peroxidases. In animals,  $\text{H}_2\text{O}_2$  is detoxified by CAT and GPX. Catalase protects cells from hydrogen peroxide generated within them. Even though CAT is not essential for some cells type under normal conditions, it plays an important role in the acquisition of tolerance to oxidative stress in the adaptive response of cells (Hunt *et al.*, 1998). The increased sensitivity of transfected enriched catalase cells to adriamycin, bleomycin and paraquat is attributed to the ability of catalase in cells to prevent the drug-induced consumption of  $\text{O}_2$ . Thus, capturing  $\text{H}_2\text{O}_2$  before it can escape the cell and converting it to  $\text{O}_2$ . In this way, catalase can maintain the concentration of  $\text{O}_2$  either for repeated rounds of chemical reduction or for direct interaction with the toxin (Speranza *et al.*, 1993).

### I.3. Glutathione peroxidase

The selenium-containing peroxidases, being the more important example glutathione peroxidase (EC 1.11.1.19), catalyze the reduction of a variety of hydroperoxides (ROOH and  $\text{H}_2\text{O}_2$ ) using GSH, thereby protecting mammalian cells against oxidative damage.





There are at least five GPX isoenzymes found in mammals. Although their expression is ubiquitous, the levels of each isoform vary depending on the tissue type. *Cytosolic and mitochondrial glutathione peroxidase* (cGPX or GPX1) reduces fatty acid hydroperoxides and  $H_2O_2$  at the expense of glutathione. GPX1 and the *phospholipid hydroperoxide glutathione peroxidase* GPX4 (or PHGPX) are found in most tissues. GPX4 is located in both the cytosol and the membrane fraction. PHGPX can directly reduce the phospholipid hydroperoxides, fatty acid hydroperoxides, and cholesterol hydroperoxides that are produced in peroxidized membranes and oxidised lipoproteins (Imai *et al.*, 1998). GPX1 is predominantly present in erythrocytes, kidney, and liver, and GPX4 is highly expressed in renal epithelial cells and testes. Cytosolic GPX2 (or GPX-G1) and extracellular GPX3 (or GPX-P) are poorly detected in most tissues except for the gastrointestinal tract and kidney, respectively. Recently, a new member, GPX5, expressed specifically in mouse epididymis, is interestingly selenium-independent (De Haan *et al.*, 1998).

GPX (80 kDa) contains one selenocysteine (Sec) residue in each of the four identical subunits, which is essential for enzyme activity (Ding *et al.*, 1998). Although GPX shares the substrate,  $H_2O_2$ , with catalase, it alone can react effectively with lipid and other organic hydroperoxides. The glutathione redox cycle is a major source of protection against low levels of oxidant stress, whereas CAT becomes more significant in protecting against severe oxidant stress (Yan and Harding, 1996). In animals cells, and specially in human erythrocytes, the principal antioxidant enzyme for the detoxification of  $H_2O_2$  has for a long time been considered to be GPX, as catalase has much lower affinity for  $H_2O_2$  than GPX (Izawa *et al.*, 1996).

Cells depleted of glutathione peroxidase were more sensitive to the toxicity of paraquat and adriamycin than untransfected parental cells from which they derived but not more sensitive to bleomycin, menadione, or phenazine methosulfate. In fact that the mildly increased sensitivity to paraquat and adriamycin was the consequence of the diminished cellular content of glutathione peroxidase was confirmed by the increase in sensitivity of untransfected cells after treatment with buthionine sulfoximine, an agent which depletes cells of glutathione. These and other data strongly suggest that the enzymatic action of GPX protects cells from the toxicity of paraquat and adriamycin. The toxin that these agents engender is likely to be hydrogen peroxide or another hydroperoxide upon which glutathione peroxidase acts (Taylor *et al.*, 1993).

GPX equally protects against the oxidation of dihydrorhodamine 123 (an indicator dye) by peroxynitrite, requiring GSH as reductant. Thus, there is also a function of GPX and potentially of other selenoproteins containing selenocysteine or selenomethionine, in the GSH-dependent maintenance of a defense line against peroxynitrite-mediated oxidations, as a peroxynitrite reductase (Sies *et al.*, 1997).

## II. REACTIVE OXYGEN SPECIES IN PATHOPHYSIOLOGIC PROCESSES

ROS generated during metabolism can enter into reactions that, when uncontrolled, can become impaired and affect certain processes leading to clinical manifestations (Babovic *et al.*, 1998). Direct effects include peroxidative changes in membranes and other cellular components, including oxidative DNA damage (Wiseman and Halliwell, 1996). Normally, the body is protected by a wide range of fluids by metal-binding macromolecules. SOD, GPX, and CAT within cells remove superoxide and peroxides before they react with metal catalysis to form more reactive species. Finally, peroxidative chain reactions initiated by reactive species that escaped enzymatic degradation are terminated by chain-breaking antioxidants, including water-soluble ascorbate, lipid-soluble vitamin E and ubiquinone. To optimize performance, oxidative stress must be controlled by supplying all known antioxidant nutrients and by minimizing effects of substances that stimulate reactive oxygen species (Miller *et al.*, 1993).

An unbalanced production of reactive oxygen intermediates has been postulated to play a role in the pathogenesis of a number of clinical disorders such as ischemia/reperfusion, atherosclerosis, neurodegenerative diseases, allergy and cancer. Besides it has been established its relationship with other specific pathologies: Alzheimer's disease, Parkinson's disease, allergic encephalomyelitis, chronic granulomatous disease, Down's syndrome, hepatitis, arthritis, HIV infection, diabetic complications, cataract formation and ulcer (Table 1). *Helicobacter pylori* generated substantial amounts of superoxide radicals. *H. pylori* infection has a different effect on mitochondrial and cytoplasmic SOD in the gastric mucous, reflected by a pronounced increase in the cytokine inducible Mn-SOD and a marginal decrease in the constitutive Cu, Zn-SOD (Götz *et al.*, 1996). In a similar fashion, linkage studies have revealed that mutations in Cu, Zn-SOD are responsible for 10-15 % of cases of the fatal motor neuron disease familial amyotrophic lateral sclerosis (Leah *et al.*, 1998). These patients have mutations in the gene encoding cytosolic Cu, Zn-SOD, and multiple lines of evidence from cell culture and transgenic models indicate that these mutations cause Cu, Zn-SOD to acquire toxic properties (Price *et al.*, 1998).

ROS have been implicated in many lung diseases including those associated with exposure to asbestos, nitrogen dioxide, ozone, paraquat, hyperoxia, carbon tetrachloride, and the anticancer drugs bleomycin and adriamycin. Phagocytic cells have been implicated in the generation of ROS during inflammation (Shull *et al.*, 1991).

Table 1. Antioxidant enzymes and human diseases.

Disease	Specification	Main key enzyme/s
Allergy	Intolerance to aspirin	GPX (Pearson <i>et al.</i> , 1991)
Allergy	Intolerance to other drugs	SOD (Ezeamuzie and Al-Hage, 1998)
Allergy	Intolerance to some foods	GPX (Malmgrem <i>et al.</i> , 1986)
Allergy	Reaction in skin tests	SOD (Cramer <i>et al.</i> , 1996)
Cancer	Bowel	CAT, GPX, SOD (Kennedy <i>et al.</i> , 1998)
Cancer	Breast	GPX (Lee <i>et al.</i> , 1998)
Cancer	Colorectal	COX-2 <sup>a</sup> (Chinery <i>et al.</i> , 1998)
Cancer	Kidney	CAT, GPX, SOD (Okamoto <i>et al.</i> , 1994)
Cancer	Leukemia	CAT, GPX, SOD (Sentüker <i>et al.</i> , 1997)
Cancer	Liver	CAT, GPX, SOD (Kennedy <i>et al.</i> , 1998)
Cancer	Skin	GPX (Shisler <i>et al.</i> , 1998)
Cardiological and vessels injuries	Ischemia	SOD (Wang <i>et al.</i> , 1998)
Cardiological and vessels injuries	Atherosclerosis	SOD (Petyaev <i>et al.</i> , 1998)
Infectious disease	Arthritis	COX-2 <sup>a</sup> (Avramidis <i>et al.</i> , 1998)
Infectious disease	<i>Helicobacter pylori</i>	SOD (Götz <i>et al.</i> , 1996)
Infectious disease	Hepatitis	GPX (Downey <i>et al.</i> , 1998)
Infectious disease	HIV	GPX (Banki <i>et al.</i> , 1998)
Infectious disease	Influenza virus	CAT, GPX, SOD (Jacoby and Choi, 1994)
Genetic disorder	Chronic granulomatous disease	CAT (Chang <i>et al.</i> , 1998)
Genetic disorder	Down's syndrome	SOD (Sustrová and Saríková, 1997)
Metabolic malfunction	Diabetes	CAT, SOD (Yan and Harding, 1997)
Neurodegenerative disease	Allergic encephalomyelitis	NOS <sup>b</sup> (Ding <i>et al.</i> , 1997)
Neurodegenerative disease	Alzheimer's disease	SOD (Furuta <i>et al.</i> , 1995)
Neurodegenerative disease	Amyotrophic lateral sclerosis	SOD (Offen <i>et al.</i> , 1998)
Neurodegenerative disease	Huntington's disease	SOD (Price <i>et al.</i> , 1998)
Neurodegenerative disease	Parkinson's disease	GPX (Offen <i>et al.</i> , 1998)
Neurodegenerative disease	Prion disease	SOD (Price <i>et al.</i> , 1998)
Ophthalmologic problem	Cataract	CAT, SOD (Yan and Harding, 1997)

<sup>a</sup>COX-2: cyclooxygenase-2; <sup>b</sup>NOS: nitric oxide synthase

Preservation of leukocyte SOD inducibility appears to correlate with longevity in elderly individuals and may be of value in predicting resistance to malignancy or fatal cardiovascular events (Niwa *et al.*, 1990). Oxygen species are key participants in damage resultant from ischemia/reperfusion. Brain and heart tissues are protected from this oxidative injury by antioxidant enzymes as SOD and GPX. Overexpression of both enzymes confers significant protection against both infarction and brain edema in transgenic mice (Wang *et al.*, 1998). Transgenic mice with overexpression of human SOD-1 are studied along with matched nontransgenic controls. In the transgenic hearts with overexpression of SOD-1 the burst of superoxide generation was almost totally quenched. This event was accompanied by a 2-fold increase in the recovery of contractile function, a 2.2-fold decrease in infarct size, and a greatly improved recovery of high energy phosphates compared with that in nontransgenic controls. These results demonstrate that superoxide is an important mediator of postischemic injury and that increasing intracellular SOD-1 dramatically protects the heart from this injury. Thus, increasing intracellular SOD-1 expression may be a highly effective approach to decrease the cellular injury that occurs following reperfusion of ischemic tissues (Vogt *et al.*, 1998). Concerning MnSOD, results exist suggesting that its messenger RNA maybe induced by oxygen radicals or by other chemical mediators, such as cytokines (Matsuyama *et al.*, 1994).

The pathogenesis of influenza virus infections of the lungs is also mediated by oxidative stress. Influenza infections cause airway epithelial inflammation and oxidant-mediated damage. In this setting, cellular antioxidant enzymes may protect airway epithelial cells against damage resulting from toxic oxygen radicals produced by activated leukocytes (Jacoby and Choi, 1994). Such infections might therefore be expected to induce expression of stress-response genes and genes encoding antioxidant enzymes and to activate transcriptional regulatory proteins.

### III. PERSPECTIVE

As a conclusion, abnormalities in the cellular regulation and expression of antioxidant enzymes have a role in cell division cycle and in the balance of life (Supek *et al.*, 1997). This understanding illustrates the importance of the antioxidant defense system in maintaining normal cellular physiology and fighting against diseases. Besides, as described above, active oxygen intermediates scavenging has been proposed as one of the mechanisms to promote immunity (Sun *et al.*, 1998).

Thus, when antioxidant, free radical scavenging systems are overwhelmed, pathologic conditions may result. Defense systems against free radical in human are a proof of the main role of antioxidant enzymes in blood cells detoxification, showing the coordinated enzymatic mechanism, and the interrelationships between all these enzymatic activities. Undoubtedly, we bet for the therapeutic implications of antioxidant activities and their possible future clinical application. Finally, we want to emphasize the importance of the study of all the regulation mechanisms at molecular level.

### ACKNOWLEDGEMENTS

This work was supported by Islamic Azad University. We wish to thank Dr. Heidari for critically reading the manuscript.

### REFERENCES

- Adachi T and X. L. Wang (1998). Association of extracellular-superoxide dismutase phenotype with the endothelial constitutive nitric oxide synthase polymorphism. *FEBS Lett.*, 433: 166-168.
- Avramidis N, A. Kourounakis, L. Hadjipetrou and V. Senchuk (1998). Anti-inflammatory and immunomodulating properties of grape melanin. Inhibitory effects on paw edema and adjuvant induced disease. *Arzneimittelforschung*, 48: 764-761.
- Babovic S, M. J. Im, M. F. Angel and P. N. Manson (1998). Role of reactive oxygen species in optic nerve compression injury: a preliminary study. *Ann Plas Surg.*, 40: 156-159.
- Banci L, M. Benedetto, I. Bertini, R. Del Conte, M. Piccioli and M. S. Viezzoli (1998). Solution structure of reduced monomeric Q133M2 copper, zinc superoxide dismutase (SOD). Why is SOD a dimeric enzyme? *Biochemistry*, 37: 11780-11791.
- Banki K, E. Hutter, N. J. Gonchoroff and A. Perl (1998). Molecular ordering in HIV-induced apoptosis. Oxidative stress, activation of caspases, and cell survival are regulated by transaldolase. *J Biol Chem.*, 273: 11944-11953.
- Benov, L and I. Fridovich (1998). Growth in iron-enriched medium partially compensates *Escherichia coli* for the lack of manganese and iron superoxide dismutase. *J Biol Chem.*, 273: 10313-10316.
- Buschfort C, M. R. Müller, S. Seeber, M. F. Rajewsky and J. Thomale (1997). DNA excision repair profiles of normal and leukemic human lymphocytes: functional analysis at the single-cell level. *Cancer Res.*, 57: 651-658.
- Chang Y, B. H. Segal, S. M. Holland, G. F. Miller and K. J. Kwon-Chung (1998). Virulence of catalase-deficient *aspergillus nidulans* in p47(phox)<sup>-/-</sup> mice. Implications for fungal pathogenicity and host defense in chronic granulomatous disease. *J Clin Invest.*, 101: 1843-1850.
- Chinery R, R. D. Beauchamp, Y. Shyr, S. C. Kirkland, R. J. Coffey and J. D. Morrow (1998). Antioxidants reduce cyclooxygenase-2 expression, prostaglandin production, and proliferation in colorectal cancer cells. *Cancer Res.*, 58: 2323-2327.
- Crameri R, A. Faith, S. Hemmann, R. Jaussi, C. Ismail, G. Menz and K. Blaser (1996). Humoral and cell-mediated autoimmunity in allergy to *Aspergillus fumigatus*. *J Exp Med.*, 184: 265-270.
- De Haan J, C. Bladier, P. Griffiths, M. Kelner, R. D. O'Shea, N. S. Cheung, R. T. Bronson, M. J. Silvestro, S. Wild, S. S. Zheng, P. M. Beart, P. J. Herzog and I. Kola (1998). Mice with a homozygous null mutation for the most abundant glutathione peroxidase, Gpx1, show increased susceptibility to the oxidative stress-inducing agents paraquat and hydrogen peroxide. *J Biol Chem.*, 273: 22528-22536.
- Ding L, Z. Liu, Z. Zhu, G. Luo, D. Zhao and J. Ni (1998). Biochemical characterization of selenium-containing catalytic antibody as a cytosolic glutathione peroxidase mimic. *Biochem J.*, 332: 251-255.



- Ding M, J. L. Wong, N. E. Rogers, L. J. Ignarro and R. R. Voskuhl (1997). Gender differences of inducible nitric oxide production in SJL/J mice with experimental autoimmune encephalomyelitis. *J Neuroimmunol.*, 77: 99-106.
- Downey J, C. D. Bingle, S. Cottrell, N. Ward, D. Churchman, M. Dobrota and C. J. Powell (1998). The LEC rat reduced hepatic selenium, contributing to the severity of spontaneous hepatitis and sensitivity to carcinogenesis. *Biochem Biophys Res Commun.*, 244: 463-467.
- Ezeamuzie C and M. Al-Hage (1998). Effects of some anti-asthma drugs on human eosinophil superoxide anions release and degranulation. *Int Arch Allergy Immunol.*, 115: 162-168.
- Furuta A, D. L. Price, C. A. Pardo, J. C. Troncoso, Z. S. Xu, N. Taniguchi and L. J. Martin (1995). Localization of superoxide dismutases in Alzheimer's disease and Down's syndrome neocortex and hippocampus. *Am J Pathol.*, 146: 357-367.
- Götz J, C. I. Va Kan, H. W. Verspaget, I. Biemond, C. B. Lamers and R. A. Veenendaal (1996). Gastric mucosal superoxide dismutases in *Helicobacter pylori* infection. *Gut.*, 38: 502-506.
- Grazioli V, R. Schiavo, E. Casari, F. Marzatico, R. Rodriguez, Y. Baena and P. Gaetani (1998). Antioxidant enzymatic activities and lipid peroxidation in cultured human chondrocytes from vertebral plate cartilage. *FEBS Lett.*, 431: 149-153.
- Guan Y, M. J. Hickey, G. E. Borgstahl, R. A. Hallewell, J. R. Lepock, D. O'Connor, Y. Hsieh, H. S. Nick, D. N. Silverman and J. A. Tainer (1998). Crystal structure of Y34F mutant human mitochondrial manganese superoxide dismutase and the functional role of tyrosine 34. *Biochemistry*, 37: 4722-4730.
- Ho Y, M. Gargano, J. Cao, R. T. Bronson, I. Heimler, and R. J. Hutz (1998). Reduced Fertility in Female Mice Lacking Copper-Zinc Superoxide Dismutase. *J Biol Chem.*, 273: 7765-7769.
- Hsieh Y, Y. Guan, C. Tu, P. J. Bratt, A. Angerhofer, J. R. Lepock, M. J. Hickey, J. A. Tainer, H. S. Nick and D. N. Silverman (1998). Probing the active site of human manganese superoxide dismutase: the role of glutamine 143. *Biochemistry*, 37: 4731-4739.
- Hunt C, J. E. Sim, S. J. Sullivan, T. Featherstone, W. Golden, C. V. Kapp-Herr, R. A. Hock, R. A. Gomez, A. J. Parsian and D. R. Spitz (1998). Genomic instability and catalase gene amplification induced by chronic exposure to oxidative stress. *Cancer Res.*, 58: 3986-3992.
- Imai H, K. Narashima, M. Arai, H. Sakamoto, N. Chiba and Y. Nakagawa (1998). Suppression of leukotriene formation in RBL-2H3 cells that overexpressed phospholipid hydroperoxide glutathione peroxidase. *J. Biol. Chem.*, 273: 1990-1997.
- Izawa S, Y. Inoue and A. Kimura (1996). Importance of catalase in the adaptive response to hydrogen peroxide: analysis of acatalasaemic *Saccharomyces cerevisiae*. *Biochem. J.*, 320: 61-67.
- Jacoby D and A. M. Choi (1994). Influenza virus induces expression of antioxidant genes in human epithelial cells. *Free Radic Biol. Med.*, 16, 821-824.
- Kennedy C, R. Cueto, S. A. Belinsky, J. F. Lechner and W. A. Pryor (1998). Overexpression of hMTH1 mRNA: a molecular marker of oxidative stress in lung cancer cells. *FEBS Lett.*, 429: 17-20.
- Leah R, D. W. Casareno and J. D. Gitlin (1998). The copper chaperone CCS directly interacts with copper/zinc superoxide dismutase. *J. Biol. Chem.*, 273: 23625-23628.
- Lee Y, S. S. Galoforo, C. M. Berns, J. C. Chen, B. H. Davis, J. E. Sim, P. M. Corry and D. R. Spitz (1998). Glucose deprivation-induced cytotoxicity and alterations in mitogen-activated protein kinase activation are mediated by oxidative stress in multidrug-resistant human breast carcinoma cells. *J. Biol. Chem.*, 273: 5294-5299.
- Lledías F, P. Rangel and W. Hansberg (1998). Oxidation of catalase by singlet oxygen. *J. Biol. Chem.*, 273: 10630-10637.
- Majima H, T. D. Oberley, K. Furukawa, M. P. Mattson, H. C. Yen, L. I. Szewda and D. K. S. Clair (1998). Prevention of Mitochondrial Injury by Manganese Superoxide Dismutase Reveals a Primary Mechanism for Alkaline-induced Cell Death. *J. Biol. Chem.*, 273: 8217-8224.
- Malmgren R, G. Unge, O. Zetterström, H. Theorell and K. De Wahl (1986). Lowered glutathione-peroxidase activity in asthmatic patients with food and aspirin intolerance. *Allergy*, 41: 43-45.
- Mataix J, J. L. Quiles, J. R. Huertas, M. Battino and M. Mañas (1998). Tissues specific interactions of exercise, dietary fatty acids, and vitamin E in lipid peroxidation. *Free Rad. Biol. Med.*, 24: 511-521.
- Matsuyama T, S. Shimizu, H. Nakamura, M. Tagaya and M. Sugita (1994). Effects of recombinant superoxide dismutase on manganese superoxide dismutase gene expression in gerbil hippocampus after ischemia. *Stroke*, 25: 1417-1424.
- Miller J, E. Brzezinska-Slebodzinska and F. C. Madsen (1993). Oxidative stress, antioxidants, and animal function. *J. Dairy Sci.*, 76: 2812-2823.

- Niwa Y, K. Ishimoto and T. Kanoh (1990). Induction of superoxide dismutase in leukocytes by paraquat: correlation with age and possible predictor of longevity. *Blood*, 76: 835-841.
- Offen D, P. M. Beart, N. S. Cheung, C. J. Pascoe, A. Hochman, S. Gorodin, E. Melamed, R. Bernard and O. Bernard (1998). Transgenic mice expressing human Bcl-2 in their neurons are resistant to 6-hydroxydopamine and 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine neurotoxicity. *Proc. Natl. Acad. Sci.*, 95: 5789-5794.
- Okamoto K, S. Toyokuni, K. Uchida, O. Ogawa, J. Takenawa, Y. Kakehi, H. Kinoshita, Y. Hattori-Nakakuki, H. Hiai and O. Hoshida (1994). Formation of 8-hydroxy-2'-deoxyguanosine and 4-hydroxy-2-nonenal-modified proteins in human renal-cell carcinoma. *Int. J. Cancer*, 58: 825-829.
- Pearson D, V. J. Suarez-Mendez, J. P. Day and P. F. Miller (1991). Selenium status in relation to reduced glutathione peroxidase activity in aspirin-sensitive asthma. *Clin. Exp. Allergy*, 21: 203-208.
- Petyaev I, M. J. M. Mitchinson, J. Hunt and P. J. Coussons (1998). Superoxide dismutase activity of antibodies purified from the human arteries and atherosclerotic lesions. *Biochem. Soc. Trans.*, 26: S43.
- Price D, S. S. Sangram and D. R. Borchelt (1998). Genetic neurodegenerative diseases: the human illness and transgenic models. *Science*, 286: 1079-1083.
- Reaume A, J. L. Elliot, E. K. Hoffman, N. W. Kowall, R. J. Ferrante, D. F. Siwek, H. M. Wilcox, D. G. Flood, M. F. Beal, R. H. Brown Jr., R. W. Scott and W. D. Snider (1996). Motor neurons in Cu/Zn superoxide dismutase-deficient mice develop normally but exhibit enhanced cell death after axonal injury. *Nature Genet.*, 13: 43-47.
- Sentüker S, B. Karahalil, M. Inal, H. Yilmaz, H. Müslümanoğlu, G. Gedikoglu and M. Dizdaroglu (1997). Oxidative DNA base damage and antioxidant enzyme levels in childhood acute lymphoblastic leukemia. *FEBS Lett.*, 416: 286-290.
- Schulze-Osthoff K, M. K. Bauer, M. Vogt and S. Wesselborg (1997). Oxidative stress and signal transduction. *Int. J. Vitam. Nutr. Res.*, 67: 336-342.
- Shisler J, T. G. Senkevich, M. J. Berry and B. Moss (1998). Ultraviolet-induced cell death blocked by a selenoprotein from a human dermatotropic poxvirus. *Science*, 279: 40-41.
- Shull S, N. H. Heintz, M. Periasamy, M. Manohar, Y. M. Janssen, J. P. Marsh and B. T. Mossman (1991). Differential regulation of antioxidant enzymes in response to oxidants. *J. Biol. Chem.*, 266: 24398-24403.
- Sies H, V. S. Sharov, L. O. Klotz and K. Briviba (1997). Glutathione peroxidase protects against peroxynitrite-mediated oxidations. A new function for selenoproteins as peroxynitrite reductase. *J. Biol. Chem.*, 272: 27812-27817.
- Speranza M, A. C. Bagley and R. E. Lynch (1993). Cells enriched for catalase are sensitized to the toxicities of bleomycin, adriamycin, and paraquat. *J. Biol. Chem.*, 268: 19039-19043.
- Sun J, Y. Chen, M. Li and Z. Ge (1998). Role of antioxidant enzymes on ionizing radiation resistance. *Free Rad. Biol. Med.*, 24: 586-593.
- Supek F, L. Supekova, H. Nelson and N. Nelson (1997). Function of metal-ion homeostasis in the cell division cycle, mitochondrial protein processing, sensitivity to mycobacterial infection and brain function. *J. Exp. Biol.*, 200: 321-330.
- Sustrová M and V. Sariková (1997). Down's syndrome-effect of increased gene expression in chromosome 21 on the function of the immune and nervous system. *Bratisl. Lek. Listy.*, 98: 221-228.
- Tampo Y, M. Tsukamoto and M. Yonaha (1998). The antioxidant action of 2-methyl-6-(p-methoxyphenyl)-3,7-dihydroimidazo[1,2- $\alpha$ ]pyrazin-3-one (MCLA), a chemiluminescence probe to detect superoxide anions. *FEBS Lett.*, 430: 348-352.
- Taylor S, L. D. Davenport, M. J. Speranza, G. T. Mullenbach and R. E. Lynch (1993). Glutathione peroxidase protects cultured mammalian cells from the toxicity of adriamycin and paraquat. *Arch. Biochem. Biophys.*, 305: 600-605.
- Vogt M, M. K. Bauer, D. Ferrari and K. Schulze-Osthoff (1998). Oxidative stress and hypoxia/reoxygenation trigger CD95 (APO-1/Fas) ligand expression in microglial cells. *FEBS Lett.*, 429: 67-72.
- Wagner B, G. R. Buettner, L. W. Oberley and C. P. Burns (1998). Sensitivity of K562 and HL-60 cells to edelfosine, an ether lipid drug, correlates with production of reactive oxygen species. *Cancer Res.*, 58: 2809-2816.
- Wang P, H. Chen, H. Qin, S. Sankarapandi, M. W. Becher, P. C. Wong and J. L. Zweier (1998). Overexpression of human copper,zinc-superoxide dismutase (SOD-1) prevents postischemic injury. *Proc. Natl. Acad. Sci.*, 95: 4556-4560.
- Westman N and S. L. Marklund (1981). Copper- and zinc-containing superoxide dismutase and manganese-containing superoxide dismutase in human tissues and human malignant tumors. *Cancer Res.*, 41: 2962-2966.
- Whittaker M and J. W. Whittaker (1998). A glutamate bridge is essential for dimer stability and metal selectivity in manganese superoxide dismutase. *J. Biol. Chem.*, 273: 88-22193.



- Wiseman, H and B. Halliwell (1996). Damage to DNA by reactive oxygen and nitrogen species: role in inflammatory disease and progression to cancer. *Biochem J.*, 313: 17-29.
- Yan, H and J. J. Harding (1997). Glycation-induced inactivation and loss of antigenicity of catalase and superoxide dismutase. *Biochem J.*, 328: 599-605.
- Youn, H, E. J. Kim, J. H. Roe, Y. C. Hah and S. O. Kang (1996). A novel nickel-containing superoxide dismutase from *Streptomyces* spp. *Biochem J.*, 318: 889-896.

(Accepted for publication May 2005)