

IN VITRO ACTIVITY OF FLUOROQUINOLONES AMONG BACTERIAL ISOLATES OF CANCER PATIENTS

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ABSTRACT

Bacteraemic infections have become the major cause of mortality and morbidity in cancer patients. Fluoroquinolones have broad spectrum of antimicrobial activity and have increasingly been used for prophylaxis against infection in cancer patients. The activity of these fluoroquinolones was evaluated in cancer patients undergoing treatment against 60 g +ve and -ve bacterial strains isolated from blood cultures. The susceptibility was determined by broth dilution method according to National Committee Clinical Laboratory Standards (NCCLS), USA guidelines. The overall respective MICs at which 50% and 90% of isolates were inhibited (MIC₅₀s and MIC₉₀s) were as follows ciprofloxacin, 4 and 8 µg/mL; ofloxacin, 16 and 64 µg/mL; pefloxacin, 16 and 128 µg/mL. The percent resistance of gram negative bacterial isolates against ciprofloxacin, ofloxacin and norfloxacin was 80%, 95% and 100% respectively. In gram positive bacteria high resistance was observed against ciprofloxacin where only 15% strains were susceptible in methicillin susceptible *Staphylococcus aureus* and 12% in methicillin resistant *Staphylococcus aureus*. Where as 100% resistance were observed in case of pefloxacin, ofloxacin and norfloxacin against both methicillin resistant and methicillin sensitive *Staphylococcus aureus*. High resistance observed in this study against clinical isolates of cancer patients against potent antimicrobial agent warrants the need of monitoring susceptibility of bacterial isolates from cancer patients for better management and treatment of patients.

Key words: Fluoroquinolones, Resistance in *Pseudomonas aeruginosa*, Resistance in *Staphylococcus aureus*.

INTRODUCTION

Infections are a major cause of morbidity and mortality among immunocompromised cancer patients undergoing chemotherapy. For this reason these patients are at risk for a wide variety of bacterial, viral, and fungal infections throughout the phases of immune recovery. The careful evaluation of bacteremic episodes that develop during the course of disease is essential for the successful anti-infective treatment (Cruciani, 2000; Madani, 2000). In the face of the increasing worldwide problem of antimicrobial resistance, many classes of antimicrobial agents have become less useful for therapy (Kunin, 1997). Recently addition of the fluoride to the quinolone antibiotic compounds yielded a new class of drugs, the fluoroquinolones, which have a broader antimicrobial spectrum and improved pharmacokinetic properties (Paton and Deeve, 1988; Siporin *et al.*, 1989; Wolfson and Hooper, 1989; Hooper and Wolfson, 1991; Gootz *et al.*, 1994). Most of these agents, including norfloxacin, ciprofloxacin, ofloxacin, pefloxacin, have broader antimicrobial spectra and improved bioavailabilities in comparison with those of naladixic acid. Fluoroquinolones are bactericidal antibiotics that act by specifically targeting DNA gyrase (Just, 1993). In contrast to amino glycosides and beta-lactams, some fluoroquinolones are active against dormant and replicating bacteria (Fitton, 1992).

Fluoroquinolones is being used for the treatment of various infections, caused by both Gram positive and Gram-negative bacteria (Gootz *et al.*, 1994; Oh *et al.*, 1996; Cormicon and Jones, 1997; Hannan and Woodnutt, 2000). Some of these have been used for the infection prevention and for the treatment of established infection in neutropenic cancer patient (Karp *et al.*, 1987; Chan *et al.*, 1989 ;). However, the widespread prophylactic and therapeutic usage of quinolones in cancer patients has raised concern about the emergence of resistance to them and a reduction in their overall impact as clinically useful antibiotics. In this study we examined the in vitro activity of ciprofloxacin, pefloxacin, ofloxacin, norfloxacin against bacterial isolates obtained from patients with cancer that are being treated at our institution.

MATERIALS AND METHODS

The study was carried out at the clinical pathology labs., Institute of Nuclear Medicine and Oncology over a period between January 2004 to June 2004.

Patients:

All hospitalized cancer patients undergoing anticancer therapy with suspected blood stream infections were studied. Patient selection was made on the basis of age or gender. Patients already on antimicrobial therapy and

those having fever due to non-infectious causes, such as blood transfusion, drug infusion etc. were excluded from the study.

Bacterial strains and culture conditions

A total of 60 bacterial isolates of *Pseudomonas aeruginosa* and *Stypholococcus aureus* isolated from blood of patients treated between January and June 2004, were studied. Isolation was made by adding Five ml blood obtained from peripheral veins of the patients to brain heart infusion (BHI) broth (Oxoid, Hampshire, UK). The blood culture bottles were incubated at 37⁰ C and regular subcultures were done. Identification of the isolates was done by Gram staining and standard biochemical tests according to the manual of clinical microbiology (Cheesbrough, 1984). All clinical isolates were identified and stored as glycerol stocks at -70°C.

Antimicrobial agents and MIC determination

Ciprofloxacin, Pefloxacin, ofloxacin and norfloxacin was obtained from commercial suppliers. Antibiotic sensitivity against fluoroquinolones was determined in *P. areuginosa* among gram negative isolates and *S. aureus* among gram positive bacteria. Minimal Inhibitory Concentration was determined in Mueller-Hinton broth (Oxoid, UK) containing serial two-fold dilutions of each antibiotic with inoculated bacterial suspensions of 5X10⁵ CFU/ml as outlined by the National Committee for Clinical Laboratory Standards (NCCLS, 1997). The results were recorded after overnight incubation at 37°C. The MIC was defined as the lowest antibiotic concentration with no visible growth. The MIC50 and MIC90 was defined as the minimum concentration of antimicrobial that inhibited 50% or 90% of the isolates, respectively. For ciprofloxacin, NCCLS breakpoints of ≤ 1 µg/mL (susceptible), and ≥ 4 µg/mL (resistant) were applied. For ofloxacin, NCCLS breakpoints of ≤ 2 µg/mL (susceptible), and ≥ 8µg/mL (resistance) were applied. For pefloxacin, NCCLS breakpoints of ≤ 4µg/mL (susceptible), and ≥ 16 µg/mL (resistant) were applied. For norfloxacin, NCCLS breakpoints of ≤ 4µg/mL (susceptible), and ≥ 16µg/mL (resistance) were applied (NCCLS 1997, 2002).

RESULTS

Bacterial Strains:

A total 60 bacterial strains of gram negative and gram positive were isolated from patients under study. Only one isolate per patient was used for testing in order to avoid duplication.

Table 1. *In vitro* activity of fluoroquinolones agents against *Pseudomonas areuginosa*.

Antimicrobial agent	Range µg/mL	MIC50 µg/mL	MIC90 µg/mL	%age sensitivity	%age resistance
Ciprofloxacin	0.125-64	4	8	20%	80%
Ofloxacin	0.5-256	16	64	5%	95%
Pefloxacin	0.5-256	16	128	0%	100%
Norfloxacin	0.5-256	32	128	0%	100%

Table 2. *In vitro* activity of fluoroquinolones agents against methecillin susceptible *Staphylococcus aureus*.

Antimicrobial agent	Range µg/mL	MIC50 µg/mL	MIC90 µg/mL	%age sensitivity	%age resistance
Ciprofloxacin	0.125-64	8	16	15%	85%
Ofloxacin	0.5-256	32	64	0%	100%
Pefloxacin	0.5-256	32	256	0%	100%
Norfloxacin	0.5-256	64	256	0%	100%

MIC determination

The over all susceptibility results are shown in Table (1, 2 &3). Ciprofloxacin showed high activity against *P. aeuginosa* among gram negative isolates The MIC₅₀ of ciprofloxacin against gram negative bacteria was 4ug/mL and inhibited 20% of isolates of *P. areuginosa* at concentration 1µg/mL (susceptibility break point). However it inhibited all the isolates of *P. areuginosa* at concentration of 32µg/mL. Ofloxacin have moderate activity against *Pseudomonas areuginosa* with MIC ranging from 0.5-256µg/mL with more than 90% of isolates being susceptible at a concentration of 64µg/mL but 95% were resistant according to NCCLS resistance break point. The activity of pefloxacin and norfloxacin was least active against *P. areuginosa* than that of ciprofloxacin and ofloxacin for isolates tested demonstrating MIC₉₀ of 128 ug/mL (Table 2).

In case of methicillin susceptible *S. aureus* MIC₅₀ against ciprofloxacin was 8 µg/mL and ranged from 0.125-64 µg/mL. Only 15% were susceptible at NCCLS break point of 2 µg/mL. In case of methicillin resistant *S. aureus* were 12%. In other fluoroquinolones (ofloxacin, pefloxacin, and norfloxacin) 100% resistance was observed against both methicillin susceptible and methicillin resistant *S. aureus* strains (Table 2 & 3).

Table 3. *In vitro* activity of fluoroquinolones against methicillin resistant *Staphylococcus aureus*.

Antimicrobial agent	Range µg/mL	MIC ₅₀ µg/mL	MIC ₉₀ µg/mL	%age sensitivity	%age resistance
Ciprofloxacin	0.125-64	8	16	12%	88%
Ofloxacin	0.5-256	32	64	0%	100%
Pefloxacin	0.5-256	32	256	0%	100%
Norfloxacin	0.5-256	64	256	0%	100%

DISCUSSION

Our results indicated comparative activity of different quinolones against Gram-negative and gram positive microorganisms isolate from cancer patients. Ciprofloxacin was found active against gram negative and gram positive microorganism. It inhibited all the isolates of *P. aeruginosa* at concentration of 32 µg/mL with only 20% being susceptible. The MIC of ciprofloxacin against members of the family *Pseudomonas* ranged from 0.125- 64 µg/mL, with more than 90% of isolates being susceptible at a concentration 8 µg/mL. The MIC₉₀ values for ofloxacin against isolates tested was 64 µg/mL which is eight fold higher than ciprofloxacin. In case of pefloxacin and norfloxacin 90% of isolates have MIC₁₂₈ µg/mL. None of the isolate were found susceptible at NCCLS susceptible break point. *In vitro* activity against gram negative indicated that MIC values of ciprofloxacin are generally lower than those of ofloxacin, pefloxacin and norfloxacin but higher than recommended NCCLS. Thus most of the strains are resistant.

Segatore *et al.* (1999) conducted a comparative survey on 334 clinical isolates of *P. aeruginosa* and found higher number of ciprofloxacin resistant strains with 17.1% was susceptible to ciprofloxacin. A decreased susceptibility to fluoroquinolone was observed among gram negative isolates in Taiwan after wide use of these antimicrobial agents in different study periods reported (Sheng *et al.*, 2002). In Europe antimicrobial susceptibility of isolates from 3136 bacteraemic versus 17261 non-bacteraemic patients was reported in MYSTIC surveillance programme and found ciprofloxacin generally exhibited the lowest activities against the most commonly isolated organisms (Unal *et al.*, 2004).

Antimicrobial susceptibility of the pathogens of bacteraemia in the UK and Ireland between 2001-2002 was determined and a resistance rate of *Pseudomonas aeruginosa* to ciprofloxacin was found 7% (Reynolds *et al.*, 2004). Ciprofloxacin and other fluoroquinolones against *S. aureus* shown decreased susceptibility in our study. Ciprofloxacin resistance is considerably higher among methicillin resistance *S. aureus* than among methicillin sensitive *S. aureus* as shown in table 2 and 3. Goldstein and Acer in 1995 reported wide spread emergence of decreased susceptibility to ciprofloxacin and other fluoroquinolone against infection with *S. aureus*. Raviglione *et al.* (1990) studied ciprofloxacin resistance in methicillin resistant *Staphylococcus aureus* in an acute care hospital and 83% resistance was observed. Lei *et al.* (2001) reported 79.4% resistance to ciprofloxacin in methicillin resistant *S. aureus*. The activity of five different fluoroquinolones against *S. aureus* strain (13 methicillin resistant strains and nine methicillin susceptible strains and found ciprofloxacin and norfloxacin were the least active quinolones as compare to trovafloxacin, levofloxacin and sparfloxacin was studied (Sierra *et al.*, 2002).

This study provides important information on the current resistance pattern among bacterial pathogens of our patients against fluoroquinolones. It can also be argued that according to the accepted selective pressure theory hypothesis that a casual relationship exist between antimicrobials use and the development of resistance. An extensive use of fluoroquinolones in the wards either for therapy or antimicrobial prophylaxis might have contributed to such high resistance rate.

Antimicrobial resistance continues to increase and going surveillance of microbial pathogens is essential for better patient management. This study also warrants the need of rapid laboratory detection of resistance to prevent the spread of resistance among bacterial strains.

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