BIOLOGICAL CONTROL OF CHARCOAL ROT OF SUNFLOWER

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ABSTRACT

In in vitro biological control of charcoal rot disease of sunflower (caused by M. nhaseolina), Trichoderma harzianum, Arachniotus sp. and Penicillium sp. provided 93, 100 and 50 per cent disease control respectively. Trichoderma harzianum significantly promoted vigour and dry stem weight where as Arachniotus sp. increased vigour, dry stem weight, plant yield and dry root weight over the non-infected healthy plants. Penicillium sp. did not affect vigour but significantly decreased plant yield and dry stem weight.

In in vivo control Trichoderma harzianum, Arachniotus sp. and Penicillium sp. provided 85, 91 and 60 per cent disease control. Trichoderma harzianum and Arachniotus sp. increased plant vigour and head size of the sunflower. The three antagonists also significantly reduced extent of linear stem infection of the diseased plants.

INTRODUCTION

In Pakistan sunflower (Helianthus annuus L.) is known to suffer from root and stem rot disorder called charcoal rot disease (Anonymous, 1980). The disease is caused by a soil borne pathogen, Macrophomina phaseolina (Ilyas et al., 1981). Symptoms of the disease (wilting, flagging, defoliation and loss of plant vigour) become prominent just prior to crop maturity and the disease causes 20-50 per cent yield losses. (Acimovic, 1962; Ilyas et al., 1981). Commercial cultivars lack resistance against M. phaseolina (Anonymous, 1980) and chemical control of the pathogen is too expansive to be practicable (Watanabe et al., 1970 Ilyas et al., 1975). The biological control involving the use of antagonistic microorganisms appears to be feasible and an economical control measure (Dhingra, 1973; Ghaffar, 1982). This paper reports on the in vitro and in vivo control of charcoal rot of sunflower by antagonistic soil fungi.

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MATERIAL AND METHODS

Preparation of inoculum of pathogen and isolates of antagonistic fungi

A highly pathogenic isolate of M. phaseolina used in this study was recovered from an infected sunflower stem, suffering from root and stem rot disease symptoms. After growth at 30°C, stock cultures on PDA were held at 4°C until used. Isolates of $Trichoderma\ harzianum$, Arachniotus sp. and Penicillium sp. antagonistic to the growth of M. phaseolina and inhibiting respectively 47, 61 and 43 per cent mycelial growth of M. phaseolina in vitro (when the inocculum of either of the antagonists and M. phaseolina were placed 60-65 mm apart on Richard agar medium), were used for the biological control studies, and were maintained on Richard agar medium at 4°C, after their growth at 25°C.

In vitro control of root and stem rot disease

The earthen pots of 45 x 30 cm size were filled with equal amount of steam sterilized soil amended with 1% wheat straw by weight. Before amending soil with wheat straw, culture of each antagonist from one Richard agar medium plate (alongwith the medium) was thoroughly hand mixed with wheat straw of each pot. Then equal amount of M. phaseolina inoculum (obtained from a 7-day old petri dish culture) was added in the form of small PDA blocks, containing mycelium and sclerotia of the pathogen, placed at 10 and 20 cm below the soil surface in the pots. The pots without antagonist but with M. phaseolina and without antagonist and without M. phaseolina served as infested and uninfested control, respectively. Ten sunflower seeds were planted in each pot and the pots were watered frequently with sterilized water. The treatments were replicated three times. Fifteen days after plant emergence, 5 vigorously growing seedlings were left in each pot, the other were uprooted. At maturity, data were recorded on per cent plant infection, plant height, yield/plant, dry stem and dry root weight. Data were analyzed statistically and differences between treatments were determined by Duncan's Multiple Range Test at 5 per cent level of significance.

In vivo control of root and stem rot disease

The effect of each of the three antagonists on the in vivo control of the disease was studied in a field where sunflower crop was not planted for the last ten years. The treatments were planted with sunflower crop in subplots of

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1.8 x 6.0 meter size. Before sowing the seed beds were throughly ploughed and soil of each treatment was mixed with wheat straw contaminated either with M. phaseolina or with antagonist or both or with non-contaminated wheat straw (uninfested control) at the rate of 100 kg straw/acre. The calculated amount of wheat straw for each subplot (treatment) was hand mixed thoroughly with inoculum of M. phaseolina and the antagonist, each taken from 7-day old culture plates, at the rate of 50 petri plates/acre of either of M. phaseolina or the antagonist. The contaminated or uncontaminated straw was broad casted on the surface of the soil of each subplot and mixed into the soil with the help of a small hand plough. The distance between treatments (subplots) was 60 cm and between replications 90 cm, whereas plant to plant distance was 30 cm. The treatments were replicated four times.

At maturity, the data were recorded on per cent plant infection, extent of linear infection in diseased stem, plant and head size. The data were analyzed statistically and differences between treatments were determined by Duncan's Multiple Range Test at 5 per cent level of significance.

RESULTS AND DISCUSSION

In vitro evaluation of the effect of three antagonists revealed that Trichoderma harzianum, Arachniotus sp. and Penicillium sp. significantly reduced the incidence of root and stem rot disease over the infested control (Table 1). The use of Trichoderma harzianum and Arachniotus sp. provided 93 and 100 per cent disease control, although there was no significant difference between the extent of disease control by the two antagonists, whereas with Penicillium sp. the extent of disease control was 50 per cent of the infested Arachniotus sp. applied to the seed have been reported to controd pre-and post-emergence damping-off of Urid beans caused by Rhizoctonia bataticola (Dhingra, 1973). M. Phaseolina infection significantly reduced plant height (31.5%) over the non infected healthy plants Trichoderma harzianum significantly increased plant height (16.3%) and dry stem weight (10.9%) over non-infected healthy plants, whereas Arachniotus sp. not only increased plant height (29.3%) and dry stem weight (21.8%) but also increased seed yield per plant (7.8%) and dry root weight (13%) over the noninfected healthy plants.

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Akhtar (1982) postulated that antagonists (Trichoderma harzianum and Arachniotus sp.) besides releasing antifungal metabolites, produce growth hormonic substances which increase the vigour and yield of the crop plants. Though Penicillium sp. had no effect on the in vitro increase of plant height, yet there was a significant decrease in seed yield/plant (11.7%) and dry stem weight (10%) over the healthy noninfected plants. This was, probably, on account of the fact that antifungal metabolites released by Penicillium sp. may be toxic to host plants.

In in vivo evaluation, the three antagonists not only provided significant disease control but also reduced the extent of linear infection of diseased stem over the infected control (Table 2). In doing so Trichoderma harzianum and Arachniotus sp. were statistically equally more effective than Penicillium sp. Ten per cent disease incidence with the extent of linear infection statistically equal to that of infested control, was found in noninfected control. probably due to naturally occurring soil born inoculum of the pathogen in the field soil. Although there was no significant effect of the antagonists on the height of diseased plant, each of the three antagonists significantly increased height of healthy plants. Trichoderma harzianum and Arachniotus sp. were statistically equally more effective than Penicillium sp. in stimulating an increase in height of the healthy plants (Table 2). This again may be on account of growth hormonic substances produced by the antagonists. There was no in vitro effects of Penicillium sp. on plant height (Table 1). This was probably on account of the fact that antifungal metabolites (Penicillins, etc.) released by Penicillium sp. were also toxic to the sunflower plants growing in vitro in the absence of other soil microorganisms. Under in vivo conditions. the antifungal metabolites were, perhaps, disintegrated by soil microorganisms and the hormonic substances released by the Penicillium sp. caused a significant increase in plant height. Contrary to plant height both Trichoderma harzianum and Arachniotus sp. caused a significant increase in the diameter of the flower heads of both the diseased and healthy plants. Penicillium sp. neither affected the head size of the diseased nor that of the healthy plants (Table 2). Effect of the three antagonists on yield could not be evaluated on account of birds damage to erop near maturity.

Table 1. In vitro effect of antagonistic soil saprophytes on disease incidence, plant height, seed yield/plant, dry

Treatments	Disease incidence	Plant height (cm)	Seed yield (gm)	Dry stem weight (gm)	Dry root weight (gm)
	(0/)		, o.	•	
Trichoderma harzianum +	7.0 c*	107 b	10.0 b	12 2 b	5.9 ab
M. phaseolina					
Arachinotus sp. +	0.0 c	119 a	11.0 a	13 4 a	6.1 в
M. phaseolina					
Penicillium sp. +	50.0 b	9 5 c	9.0 c	9.9 d	5,3 b
M. phaseolina					
No antagonist + M. phaseolina	100.0 a	63 d	0.0 d	10.0 d	4.2 c
(infested control)					
No antagonist +	0.0 c	. 92 с	10.2 b	11.0 с	5.4 b
No M. phaseolina					
(Noninfested control)	•				
* Figures in the same column with the same letter do not differ significantly at 5% level of significance.	th the same letter	r do not differ	significant	y at 5% level	of significance.

Table 2. In vivo effect of antagonistic saprophytic fungi on per eent mortality by Macrophomina phaseolina,

Treatments	Disease	Extent of linear	Plant height (cm)	ght (cm)	Head si	lead size (cm)
	(%)	stem above soil level (cm)	Healthy Diseased		Healthy I	lthy Diseased
Trichoderma harzianum + M. phaseolina	15 c*	25 е	199 &	141	16.0 a	988
Arachniotus sp. + M. phaseolina	9 0	24 0	20 3 a	144	15 8 a	10.2 a
Penicillium sp. + M.phaseolina	40 b	49 b	183 b	134	14.9 b	7.1 b
No antagonist + M.phaseolina (infested control)	68 a	111 &	143 с	138	14.8 խ	7.0 b
No antagonist + No M. Phaseolina	10. с	104 в	147 с	132 NS	14.4 b	7.5 b
(Non-infested control)						

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