



Pharmacological Evaluation and Synthesis of New Sulfonamides Derivatives Based on 1,4-Benzodioxane

M. Irshad^{*1}, M. A. Abbasi², Aziz-ur-Rehman², M. Akram¹, Q. Ali¹, S. Z. Siddiqui², M. Shahid³, M. Ashraf⁴, M. A. Lodhi⁵ and S. B. Jamal⁶

¹Division of Science and Technology, University of Education, Township Lahore-54770, Pakistan.

²Department of Chemistry, Government College University, Lahore-54000, Pakistan.

³Department of Chemistry and Biochemistry, University of Agriculture, Faisalabad-38040, Pakistan.

⁴Department of Pharmacy, The Islamia University of Bahawalpur, Bahawalpur-63100, Pakistan.

⁵Department of Biochemistry, Abdul Wali Khan University, Mardan-23200, Pakistan.

⁶Department of Bioinformatics, Islamic International University, Islamabad, Pakistan.

*Corresponding Author Email: misbah-irshad@ue.edu.pk

Received 18 May 2018, Revised 30 November 2018, Accepted 05 December 2018

Abstract

We report here the synthesis of a series of *N*-aryl-2,3-dihydrobenzo[1,4]dioxine-6-sulfonamide and its *N*-substituted derivatives with benzyl chloride and ethyl iodide. Initially, 2,3-dihydrobenzo[1,4]dioxine-6-sulfonyl chloride (1) was subjected to react with various aryl amines (2a-e) to afford parent compounds *N*-aryl-2,3-dihydrobenzo[1,4]dioxine-6-sulfonamide (3a-e). At second step, these parent compounds were reacted with benzyl chloride (4) and ethyl iodide (5) as to synthesize *N*-benzyl-*N*-aryl-2,3-dihydrobenzo[1,4]dioxine-6-sulfonamide (6a-e) and *N*-ethyl-*N*-aryl-2,3-dihydrobenzo[1,4]dioxine-6-sulfonamide (7a-e) in the presence of lithium hydride and *N,N'*-dimethylformamide respectively. FT-IR, Nuclear Magnetic Resonance (¹H-NMR) and Mass Spectrometry (MS) techniques were used to investigate the structures of these synthesized compounds. A fingerprinted study was conducted against some enzymes like butyrylcholinesterase (BChE), acetylcholinesterase (AChE) and lipoxygenase (LOX). This study revealed that most of them demonstrated a moderate activity against butyrylcholinesterase (BChE) and acetylcholinesterase (AChE) however promisingly a good activity against lipoxygenase enzyme was observed. Finally, an antimicrobial and hemolytic activities of these sulfonamides were probed which confirmed that the parent sulfonamides 3b have the proficient antimicrobial activities, while the derivatives 6a, 7a, 7b and 7c explored a good activity against the selected panel of bacterial and fungal species. All the compounds were further computationally docked against (LOX), (BChE) and (AChE) enzymes and these interaction highlighted the importance of sulfonamides in the inhibition of the target enzymes.

Keywords: 2,3-dihydrobenzo[1,4]dioxine-6-sulfonyl chloride, Lipoxygenase enzyme, ¹H-NMR, EI-MS, Antimicrobial and hemolytic activities, Molecular docking.

Introduction

As the first effective antibacterial agents sulfonamides were intensively investigated. A broad-spectrum of synthetic bacteriostatic antibiotics is included in this family, which are used against most gram-positive and many gram-negative microorganisms. These compounds are commonly used for therapeutic and prophylactic

purposes to fight against common bacterial diseases. Human and veterinary medicine are included in this family [1]. Furthermore, these substances are also used as feed additives in animal husbandry [2]. After tetracyclines sulfonamides are the second most widely used class of veterinary antibiotics in the European countries [3].

Sulfonamides inhibit dihydropteroate synthase just like 4-aminobenzoic acid due to structural similarities. After the discovery of penicillin and other antibiotics their utility was reduced, but later they have started to attract attention for their synergic activity, e.g. in the combination with trimethoprim. The combination of sulfamethoxazole (SMX) with trimethoprim exhibit more efficient antibacterial activity due to the sequential inhibition of the bacterial synthesis of tetrahydrofolic acid and thereby disrupting nucleic bases and acids synthesis [4-6]. Because of their ease of administration and non-interaction with defense mechanism of host some derivatives of sulfonamides are extensively used for gastrointestinal and urinary tract infections [7]. In recent times, sulfonamides have been found to be powerful carbonic anhydrase [8], COX-2 [9,10] and caspase inhibitors [11] and have applications in veterinary practices [12].

The 1,4-benzodioxane framework has often been found in biologically active lignans. Silybin [13] and americanin A [14] are used as antihepatotoxic agents; while haedoxan A [15] exhibited insecticidal activity. Silybin, containing large amount of benzodioxane, has been used as a folk medicine in Jammu Kashmir and Europe [16]. This type of natural product, which has shown a variety of bioactivities, is of synthetic interest from many years. Receptor systems are usually composed of multiple subtypes such as α -adrenoreceptors. The majority of α -adrenoreceptor antagonists displays a competitive mechanism of action and belongs to a variety of different structural classes such as yohimbanes, ergot alkaloids, quinazolines, *N*-arylpiperazines, imidazolines, phenylalkylamines, benzodioxanes, indoles, 1,4-dihydropyridines, hetero-fused 3-benzazepines and dibenzoquinolizines [17-19]. Different biological activities like antihepatotoxic [20-22], α -adrenergic blocking agent [23], anti-inflammatory [24] and D₂ antagonist/5-HT_{1A} partial agonist activity are exhibited by compounds containing dioxane ring systems [25].

Additionally, molecular docking approach was used to find out the interaction mode of the synthesized compounds. The purpose of docking methodologies was to forecast the ligand and target complex and to align the molecular database

(designed inhibitors) on the basis of binding affinity to that of target. The MOE-Dock was used for docking of all the synthesized inhibitors with the binding site of target enzymes. The eventual objective of molecular docking was to get ligands with better characteristics and have good inhibition potential [26].

This research work is a productive effort to bring in pharmacologically significant compounds. In continuation of our previous work on sulfonamide synthesis [27], the designing of different *N*-substituted sulfonamides derived from 1,4-benzodioxine-6-sulfonyl chloride with an aim to inaugurate new challenges of drug having striking activity for the cure of legionnaires' diseases.

Experimental

General

Griffin and George melting point apparatus was used to record the melting points of the synthesized compounds on an open capillary tube and were not accurate. The progress of reaction and purity was confirmed by TLC; performed on silica gel plates (G-25-UV₂₅₄). The ethyl acetate and *n*-hexane solvent system in 30: 70 % was employed as mobile phase. Detection was carried out at 254 nm and ceric sulphate was used as developing reagent. The FT-IR spectra were conducted on a Jasco-320-A spectrophotometer and results were interpreted in cm⁻¹. On a Bruker spectrometers ¹H-NMR spectra were recorded with deuterated chloroform and methanol; the resolution frequency were 300 and 400 MHz. Mass spectra statistic were employed on a JMS-HX-110 spectrometer. 2,3-Dihydrobenzo[1,4]dioxine-6-sulfonyl chloride, aryl amines and the other electrophilic reagents were purchased from Merck and Alfa Aesar through local suppliers and were used without further purification. All the employed solvents were of analytical grade.

Synthetic Work

General procedure for the synthesis of N-aryl-2,3-dihydrobenzo[1,4]dioxine-6-sulfonamides in aqueous medium (3a-e)

1 mmol of various substituted aryl amines (**2a-e**) were suspended in 50 mL water in 250 mL

round bottom flask. The pH was maintained at 9.0-10.0 by adding basic aqueous solution of Na_2CO_3 at 25 °C. Then 2,3-dihydrobenzo[1,4]dioxine-6-sulfonyl chloride (1 mmol, 0.234 g; 1) was added in the reaction mass slowly over 10-15 min. The reaction was conducted by simple stirring at RT and monitored by thin layer chromatography. Conc. HCl (about 2 mL) was added slowly to adjust the pH to 2.0. The solid product were precipitate out, collected by filtration and flushed with water to afford the precursors sulfonamides (3a-e) on drying.

***N*-(3,5-Dimethylphenyl)-2,3-dihydrobenzo[1,4]dioxine-6-sulfonamide (3a)**

IR (KBr, cm^{-1}): ν_{max} : 3421 (N-H, stretching), 3031 (C-H, stretching of aromatic ring), 2918 ($-\text{CH}_2-$, stretching), 1621 (C=C, stretching of aromatic ring), 1321 ($-\text{SO}_2-$, stretching), 1115 (C-O-C, stretching of ether); $^1\text{H-NMR}$: δ (ppm) 7.30 (d, $J = 2.0$ Hz, 1H, H-5), 7.21 (dd, $J = 8.4, 2.4$ Hz, 1H, H-7), 6.85 (d, $J = 8.4$ Hz, 1H, H-8), 6.71 (s, 2H, H-2' and H-6'), 6.66 (s, 1H, H-4'), 4.23-4.25 (m, 4H, CH_2 -2 and CH_2 -3), 2.21 (s, 6H, CH_3 -1" and CH_3 -2"); EIMS: m/z 319 $[\text{M}]^+$, 255 $[\text{M}-\text{SO}_2]^+$, 214 $[\text{C}_6\text{H}_3\text{C}_2\text{H}_4\text{O}_2\text{SO}_2\text{NH}]^+$, 199 $[\text{C}_6\text{H}_3\text{C}_2\text{H}_4\text{O}_2\text{SO}_2]^+$, 135 $[\text{C}_6\text{H}_3\text{C}_2\text{H}_4\text{O}_2]^+$, 107 $[\text{C}_6\text{H}_3\text{O}_2]^+$, 105 $[\text{C}_6\text{H}_3(\text{CH}_3)_2]^+$, 90 $[\text{C}_6\text{H}_3\text{CH}_3]^+$, 75 $[\text{C}_6\text{H}_3]^+$, 65 $[\text{C}_4\text{H}_2\text{CH}_3]^+$. (Calcd. for $\text{C}_{16}\text{H}_{17}\text{NO}_4\text{S}$; 319.3854)

***N*-(4-Methylphenyl)-2,3-dihydrobenzo[1,4]dioxine-6-sulfonamide (3b)**

IR (KBr, cm^{-1}): ν_{max} : 3412 (N-H, stretching), 3022 (C-H, stretching of aromatic ring), 2914 ($-\text{CH}_2-$, stretching), 1617 (C=C, stretching of aromatic ring), 1326 ($-\text{SO}_2-$, stretching), 1145 (C-O-C, stretching of ether); $^1\text{H-NMR}$: δ (ppm) 7.27 (d, $J = 2.0$ Hz, 1H, H-5), 7.19 (dd, $J = 8.4, 2.0$ Hz, 1H, H-7), 7.03 (d, $J = 8.0$ Hz, 1H, H-8), 6.93 (d, $J = 8.0$ Hz, 2H, H-2' and H-6'), 6.84 (d, $J = 8.4$ Hz, 2H, H-3' and H-5'), 4.22-4.26 (m, 4H, CH_2 -2 and CH_2 -3), 2.26 (s, 3H, CH_3 -1"); EIMS: m/z 305 $[\text{M}]^+$, 241 $[\text{M}-\text{SO}_2]^+$, 214 $[\text{C}_6\text{H}_3\text{C}_2\text{H}_4\text{O}_2\text{SO}_2\text{NH}]^+$, 199 $[\text{C}_6\text{H}_3\text{C}_2\text{H}_4\text{O}_2\text{SO}_2]^+$, 135 $[\text{C}_6\text{H}_3\text{C}_2\text{H}_4\text{O}_2]^+$, 107 $[\text{C}_6\text{H}_3\text{O}_2]^+$, 91 $[\text{C}_6\text{H}_4\text{CH}_3]^+$, 76 $[\text{C}_6\text{H}_4]^+$, 75 $[\text{C}_6\text{H}_3]^+$, 50 $[\text{C}_4\text{H}_2]^+$. (Calcd. for $\text{C}_{15}\text{H}_{15}\text{NO}_4\text{S}$; 305.3654).

***N*-(3-Hydroxyphenyl)-2,3-dihydrobenzo[1,4]dioxine-6-sulfonamide (3c)**

IR (KBr, cm^{-1}): ν_{max} : 3455 (N-H, stretching), 3312 (O-H, stretching), 3018 (C-H, stretching of aromatic ring), 2916 ($-\text{CH}_2-$, stretching), 1612 (C=C, stretching of aromatic ring), 1324 ($-\text{SO}_2-$, stretching), 1136 (C-O-C, stretching of ether); $^1\text{H-NMR}$: δ (ppm) 8.26 (s, 1H, O-H), 7.28 (dd, $J = 8.4, 2.2$ Hz, 1H, H-7), 7.16 (d, $J = 2.4$ Hz, 1H, H-5), 7.13 (d, $J = 8.0$ Hz, 1H, H-8), 6.97 (brt, $J = 7.8$ Hz, 1H, H-5'), 6.90 (dd, $J = 1.2, 1.2$ Hz, 1H, H-2'), 6.83 (dd, $J = 8.4, 1.6$ Hz, 1H, H-4'), 6.63 (dd, $J = 8.0, 1.4$ Hz, 1H, H-6'), 4.24-4.32 (m, 4H, CH_2 -2 and CH_2 -3); EIMS: m/z 307 $[\text{M}]^+$, 243 $[\text{M}-\text{SO}_2]^+$, 214 $[\text{C}_6\text{H}_3\text{C}_2\text{H}_4\text{O}_2\text{SO}_2\text{NH}]^+$, 199 $[\text{C}_6\text{H}_3\text{C}_2\text{H}_4\text{O}_2\text{SO}_2]^+$, 135 $[\text{C}_6\text{H}_3\text{C}_2\text{H}_4\text{O}_2]^+$, 107 $[\text{C}_6\text{H}_3\text{O}_2]^+$, 93 $[\text{C}_6\text{H}_4\text{OH}]^+$, 76 $[\text{C}_6\text{H}_4]^+$, 75 $[\text{C}_6\text{H}_3]^+$, 50 $[\text{C}_4\text{H}_2]^+$. Calcd. for $\text{C}_{14}\text{H}_{13}\text{NO}_5\text{S}$; 307.3442

***N*-(Benzyl)-2,3-dihydrobenzo[1,4]dioxine-6-sulfonamide (3d)**

IR (KBr, cm^{-1}): ν_{max} : 3455 (N-H, stretching), 3022 (C-H, stretching of aromatic ring), 2913 ($-\text{CH}_2-$, stretching), 1609 (C=C, stretching of aromatic ring), 1323 ($-\text{SO}_2-$, stretching), 1127 (C-O, stretching of ether); $^1\text{H-NMR}$: δ (ppm) 7.29 (dd, $J = 2.0, 8.4$ Hz, 1H, H-7), 7.25 (d, $J = 2.0$ Hz, 1H, H-5), 7.20-7.24 (m, 5H, H-2' to H-6'), 6.92 (d, $J = 8.4$ Hz, 1H, H-8), 4.29-4.30 (m, 4H, CH_2 -2 and CH_2 -3), 4.01 (s, 2H, CH_2 -7"); EIMS: m/z 305 $[\text{M}]^+$, 241 $[\text{M}-\text{SO}_2]^+$, 228 $[\text{C}_6\text{H}_3\text{C}_2\text{H}_4\text{O}_2\text{SO}_2\text{NHCH}_2]^+$, 200 $[\text{C}_6\text{H}_3\text{O}_2\text{SO}_2\text{NHCH}_2]^+$, 199 $[\text{C}_6\text{H}_3\text{C}_2\text{H}_4\text{O}_2\text{SO}_2]^+$, 136 $[\text{C}_6\text{H}_3\text{O}_2\text{NHCH}_2]^+$, 135 $[\text{C}_6\text{H}_3\text{C}_2\text{H}_4\text{O}_2]^+$, 107 $[\text{C}_6\text{H}_3\text{O}_2]^+$, 106 $[\text{C}_6\text{H}_5\text{CH}_2\text{NH}]^+$, 77 $[\text{C}_6\text{H}_5]^+$, 75 $[\text{C}_6\text{H}_3]^+$, 51 $[\text{C}_4\text{H}_3]^+$. (Calcd. for $\text{C}_{15}\text{H}_{15}\text{NO}_4\text{S}$; 305.3254)

***N*-(2-Phenylethyl)-2,3-dihydrobenzo[1,4]dioxine-6-sulfonamide (3e)**

IR (KBr, cm^{-1}): ν_{max} : 3398 (N-H, stretching), 3010 (C-H, stretching of aromatic ring), 2910 ($-\text{CH}_2-$, stretching), 1610 (C=C, stretching of aromatic ring), 1321 ($-\text{SO}_2-$, stretching), 1134 (C-O-C, stretching of ether); $^1\text{H-NMR}$: δ (ppm) 7.31 (d, $J = 2.0$ Hz, 1H, H-5), 7.28 (dd, $J = 8.4, 2.0$ Hz, 1H, H-7), 7.21 (d, $J = 8.4$ Hz,

1H, H-8), 7.06-7.13 (m, 5H, H-2' to H-6'), 4.27-4.28 (m, 4H, CH₂-2 and CH₂-3), 3.21 (t, *J* = 6.8 Hz, 2H, CH₂-8'), 2.77 (t, *J* = 6.8 Hz, 2H, CH₂-7'); EIMS: *m/z* 319 [M]⁺, 255 [M-SO₂]⁺, 228 [C₆H₃C₂H₄O₂SO₂NHCH₂]⁺, 200 [C₆H₃O₂SO₂NHCH₂]⁺, 199 [C₆H₃C₂H₄O₂SO₂]⁺, 136 [C₆H₃O₂NHCH₂]⁺, 135 [C₆H₃C₂H₄O₂]⁺, 120 [C₆H₅(CH₂)₂NH]⁺, 107 [C₆H₃O₂]⁺, 91 [C₇H₇]⁺, 75 [C₆H₃]⁺, 65 [C₅H₅]⁺. (Calcd. for C₁₆H₁₇NO₄S; 319.3854)

General procedure for the synthesis of compounds 6a-e and 7a-e:

Lithium hydride (0.01 g, 0.40 mmol) was added to a solution of compound (0.1 g, 3a-e) in *N,N*-dimethylformamide (DMF, 5 mL) at 25 °C. After that the reaction mixture was stirred for 30 min. The benzyl chloride (4) and ethyl iodide (5) were poured slowly and stirring continued for 1-2 hr. The complete conversion of reactants into derivatives was elucidated from TLC, the subsequent addition of cold distilled water yielded precipitates. The obtained solid was filtered, washed with distilled water and dried to yield the corresponding *N*-benzyl/ethyl derivatives of *N*-aryl-2,3-dihydrobenzo[1,4]dioxine-6-sulfonamide (6a-e and 7a-e). In some cases, compound was taken out through solvent extraction method by chloroform.

***N*-Benzyl-*N*-(3,5-dimethylphenyl)-2,3-dihydrobenzo[1,4]dioxine-6-sulfonamide (6a)**

IR (KBr, cm⁻¹): *v*_{max}: 3419 (N-H, stretching), 3031 (C-H, stretching of aromatic ring), 2927 (-CH₂-, stretching), 1613 (C=C, stretching of aromatic ring), 1324 (-SO₂-, stretching), 1122 (C-O-C, stretching of ether); ¹H-NMR: δ (ppm) 7.19-7.24 (m, 5H, H-2" to H-6"), 7.17 (d, *J* = 2.4 Hz, 1H, H-5), 7.11 (dd, *J* = 8.0, 2.4 Hz, 1H, H-7), 7.08 (d, *J* = 8.0 Hz, 1H, H-8), 6.98 (s, 2H, H-2' and H-6'), 6.95 (s, 1H, H-4'), 4.68 (s, 2H, CH₂-7"), 4.27-4.33 (m, 4H, CH₂-2 and CH₂-3), 2.14 (s, 6H, CH₃-1" and CH₃-2"); EIMS: *m/z* 409 [M]⁺, 345 [M-SO₂]⁺, 304 [C₆H₃C₂H₄O₂SO₂NCH₂C₆H₅]⁺, 199 [C₆H₃C₂H₄O₂SO₂]⁺, 135 [C₆H₃C₂H₄O₂]⁺, 107 [C₆H₃O₂]⁺, 105 [C₆H₃(CH₃)₂]⁺, 91 [C₇H₇]⁺, 90 [C₆H₃CH₃]⁺, 75 [C₆H₃]⁺, 65 [C₅H₅]⁺. (Calcd. for C₂₃H₂₃NO₄S; 409.5187)

***N*-Benzyl-*N*-(4-methylphenyl)-2,3-dihydrobenzo[1,4]dioxine-6-sulfonamide (6b)**

IR (KBr, cm⁻¹): *v*_{max}: 3412 (N-H, stretching), 3065 (C-H, stretching of aromatic ring), 2927 (-CH₂-, stretching), 1618 (C=C, stretching of aromatic ring), 1327 (-SO₂-, stretching), 1132 (C-O-C, stretching of ether); ¹H-NMR: δ (ppm) 7.19 (d, *J* = 2.4 Hz, 1H, H-5), 7.10 (dd, *J* = 8.4, 2.4 Hz, 1H, H-7), 6.98 (d, *J* = 8.0 Hz, 1H, H-8), 6.86-6.89 (m, 5H, H-2" to H-6"), 6.84 (d, *J* = 8.0 Hz, 2H, H-2' and H-6'), 6.81 (d, *J* = 8.4 Hz, 2H, H-3' and H-5'), 4.66 (s, 2H, CH₂-7"), 4.28-4.30 (m, 4H, CH₂-2 and CH₂-3), 2.23 (s, 3H, CH₃-1"); EIMS *m/z*: 395 [M]⁺, 331 [M-SO₂]⁺, 304 [C₆H₃C₂H₄O₂SO₂NCH₂C₆H₅]⁺, 199 [C₆H₃C₂H₄O₂SO₂]⁺, 135 [C₆H₃C₂H₄O₂]⁺, 107 [C₆H₃O₂]⁺, 91 [C₇H₇]⁺, 76 [C₆H₄]⁺, 75 [C₆H₃]⁺, 65 [C₅H₅]⁺, 50 [C₄H₂]⁺. (Calcd. for C₂₂H₂₁NO₄S; 395.4925)

***N*-Benzyl-*N*-(3-hydroxyphenyl)-2,3-dihydrobenzo[1,4]dioxine-6-sulfonamide (6c)**

IR (KBr, cm⁻¹): *v*_{max}: 3413 (N-H, stretching), 3316 (O-H, stretching), 3067 (C-H, stretching of aromatic ring), 2931 (-CH₂-, stretching), 1643 (C=C, stretching of aromatic ring), 1329 (-SO₂-, stretching), 1127 (C-O-C, stretching of ether); ¹H-NMR: δ (ppm) 8.25 (s, 1H, OH), 7.34 (dd, *J* = 8.4, 2.0 Hz, 1H, H-7), 7.22 (d, *J* = 2.4 Hz, 1H, H-5), 7.12 (d, *J* = 8.0 Hz, 1H, H-8), 7.02 (brt, *J* = 7.2 Hz, 1H, H-5'), 6.92-7.01 (m, 5H, H-2" to H-6"), 6.84 (dd, *J* = 1.2, 1.2 Hz, 1H, H-2'), 6.68 (dd, *J* = 8.0, 1.2 Hz, 1H, H-4'), 6.42 (dd, *J* = 8.0, 1.2 Hz, 1H, H-6'), 4.62 (s, 2H, CH₂-7"), 4.25-4.33 (m, 4H, CH₂-2 and CH₂-3); EIMS: *m/z* 397 [M]⁺, 333 [M-SO₂]⁺, 304 [C₆H₃C₂H₄O₂SO₂NCH₂C₆H₅]⁺, 199 [C₆H₃C₂H₄O₂SO₂]⁺, 135 [C₆H₃C₂H₄O₂]⁺, 107 [C₆H₃O₂]⁺, 93 [C₆H₄OH]⁺, 91 [C₇H₇]⁺, 76 [C₆H₄]⁺, 75 [C₆H₃]⁺, 65 [C₅H₅]⁺, 50 [C₄H₂]⁺. (Calcd. for C₂₁H₁₉NO₅S; 395.4623)

***N,N*-Dibenzyl-2,3-dihydrobenzo[1,4]dioxine-6-sulfonamide (6d)**

IR (KBr, cm⁻¹): *v*_{max}: 3412 (N-H, stretching), 3034 (C-H, stretching of aromatic ring), 2945 (-CH₂-, stretching), 1613 (C=C, stretching of aromatic ring), 1327 (-SO₂-, stretching), 1135 (C-O, stretching of ether); ¹H-NMR: δ (ppm) 7.33 (dd, *J* = 2.0, 8.0 Hz, 1H, H-7), 7.28 (d, *J* = 2.4 Hz, 1H,

H-5), 7.04-7.18 (m, 10H, H-2' to H-6' and H-2'' to H-6''), 6.98 (d, $J = 8.4$ Hz, 1H, H-8), 4.29-4.32 (m, 4H, CH₂-2 and CH₂-3), 4.28 (s, 4H, CH₂-7' and CH₂-7''); EIMS: m/z 395 [M]⁺, 331 [M-SO₂]⁺, 318 [C₆H₃C₂H₄O₂SO₂NC₆H₅(CH₂)₂]⁺, 290 [C₆H₃O₂SO₂NC₆H₅(CH₂)₂]⁺, 240 [C₆H₃C₂H₄O₂NCH₂C₆H₅]⁺, 212 [C₆H₃O₂NCH₂C₆H₅]⁺, 199 [C₆H₃C₂H₄O₂SO₂]⁺, 135 [C₆H₃C₂H₄O₂]⁺, 107 [C₆H₃O₂]⁺, 91 [C₇H₇]⁺, 77 [C₆H₅]⁺, 75 [C₆H₃]⁺, 65 [C₅H₅]⁺, 51 [C₄H₃]⁺. (Calcd. for C₂₂H₂₁NO₄S; 395.4821)

N-Benzyl-N-(2-phenylethyl)-2,3-dihydrobenzo [1,4]dioxine-6-sulfonamide (6e)

IR (KBr, cm⁻¹): ν_{\max} : 3423 (N-H, stretching), 3043 (C-H, stretching of aromatic ring), 2921 (-CH₂-, stretching), 1618 (C=C, stretching of aromatic ring), 1328 (-SO₂-, stretching), 1142 (C-O-C, stretching of ether); ¹H-NMR: δ (ppm) 7.32 (dd, $J = 8.4, 2.0$ Hz, 1H, H-7), 7.26 (d, $J = 2.4$ Hz, 1H, H-5), 7.17 (d, $J = 8.0$ Hz, 1H, H-8), 6.91-7.15 (m, 10H, H-2' to H-6' and H-2'' to H-6''), 4.31 (s, 4H, CH₂-2 and CH₂-3), 4.20 (s, 2H, CH₂-7''), 2.72 (t, $J = 7.6$ Hz, 2H, CH₂-8'), 2.58 (t, $J = 7.6$ Hz, 2H, CH₂-7'); EIMS: m/z 409 [M]⁺, 345 [M-SO₂]⁺, 318 [C₆H₃C₂H₄O₂SO₂NC₆H₅(CH₂)₂]⁺, 290 [C₆H₃O₂SO₂NC₆H₅(CH₂)₂]⁺, 254 [C₆H₃C₂H₄O₂NC₆H₅(CH₂)₂]⁺, 226 [C₆H₃O₂NC₆H₅(CH₂)₂]⁺, 199 [C₆H₃C₂H₄O₂SO₂]⁺, 135 [C₆H₃C₂H₄O₂]⁺, 107 [C₆H₃O₂]⁺, 91 [C₇H₇]⁺, 77 [C₆H₅]⁺, 75 [C₆H₃]⁺, 65 [C₅H₅]⁺, 51 [C₄H₃]⁺. (Calcd. for C₂₃H₂₃NO₄S; 409.5187).

N-(3,5-Dimethylphenyl)-N-ethyl-2,3-dihydrobenzo[1,4]dioxine-6-sulfonamide (7a)

IR (KBr, cm⁻¹): ν_{\max} : 3428 (N-H, stretching), 3021 (C-H, stretching of aromatic ring), 2927 (-CH₂-, stretching), 1623 (C=C, stretching of aromatic ring), 1328 (-SO₂-, stretching), 1127 (C-O-C, stretching of ether); ¹H-NMR: δ (ppm) 7.06 (dd, $J = 8.4, 2.4$ Hz, 1H, H-7), 7.01 (d, $J = 2.4$ Hz, 1H, H-5), 6.95 (d, $J = 8.4$ Hz, 1H, H-8), 6.62 (s, 2H, H-2' and H-6'), 6.59 (s, 1H, H-4'), 4.27-4.33 (m, 4H, CH₂-2 and CH₂-3), 3.54 (q, $J = 7.2$ Hz, 2H, CH₂-1''), 2.23 (s, 6H, CH₃-1''' and CH₃-2'''), 1.01 (t, $J = 7.2$ Hz, 3H, CH₃-2''); EIMS: m/z 347 [M]⁺, 283 [M-SO₂]⁺, 268 [C₆H₃C₂H₄O₂NCH₂C₆H₃(CH₃)₂]⁺, 242 [C₆H₃C₂H₄O₂SO₂NC₂H₅]⁺, 240 [C₆H₃O₂NCH₂C₆H₃(CH₃)₂]⁺,

227 [C₆H₃C₂H₄O₂SO₂NCH₂]⁺, 199 [C₆H₃C₂H₄O₂SO₂]⁺, 135 [C₆H₃C₂H₄O₂]⁺, 107 [C₆H₃O₂]⁺, 105 [C₆H₃(CH₃)₂]⁺, 90 [C₆H₃CH₃]⁺, 65 [C₄H₂CH₃]⁺, 75 [C₆H₃]⁺. (Calcd. for C₁₈H₂₁NO₄S; 347.4587).

N-Ethyl-N-(4-methylphenyl)-2,3-dihydrobenzo [1,4]dioxine-6-sulfonamide (7b)

IR (KBr, cm⁻¹): ν_{\max} : 3443 (N-H, stretching), 3028 (C-H, stretching of aromatic ring), 2929 (-CH₂-, stretching), 1636 (C=C, stretching of aromatic ring), 1324 (-SO₂-, stretching), 1145 (C-O-C, stretching of ether); ¹H-NMR: δ (ppm) 7.16 (d, $J = 2.0$ Hz, 1H, H-5), 7.06 (dd, $J = 8.4, 2.0$ Hz, 1H, H-7), 6.99 (d, $J = 8.0$ Hz, 1H, H-8), 6.92 (d, $J = 8.0$ Hz, 2H, H-2' and H-6'), 6.86 (d, $J = 8.8$ Hz, 2H, H-3' and H-5'), 4.25-4.28 (m, 4H, CH₂-2 and CH₂-3), 3.56 (q, $J = 7.2$ Hz, 2H, CH₂-1''), 2.31 (s, 3H, CH₃-1'''), 1.03 (t, $J = 7.2$ Hz, 3H, CH₃-2''); EIMS: m/z 333 [M]⁺, 269 [M-SO₂]⁺, 254 [C₆H₃C₂H₄O₂NCH₂C₆H₄CH₃]⁺, 242 [C₆H₃C₂H₄O₂NSO₂C₂H₅]⁺, 227 [C₆H₃C₂H₄O₂SO₂NCH₂]⁺, 226 [C₆H₃O₂NCH₂C₆H₄CH₃]⁺, 199 [C₆H₃C₂H₄O₂SO₂]⁺, 135 [C₆H₃C₂H₄O₂]⁺, 107 [C₆H₃O₂]⁺, 91 [C₆H₄CH₃]⁺, 76 [C₆H₄]⁺, 75 [C₆H₃]⁺, 50 [C₄H₂]⁺. (Calcd. for C₁₇H₁₉NO₄S; 333.4542).

N-Ethyl-N-(3-hydroxyphenyl)-2,3-dihydrobenzo [1,4]dioxine-6-sulfonamide (7c)

IR (KBr, cm⁻¹): ν_{\max} : 3454 (N-H, stretching), 3067 (C-H, stretching of aromatic ring), 3312 (O-H, stretching), 2912 (-CH₂-, stretching), 1645 (C=C, stretching of aromatic ring), 1327 (-SO₂-, stretching), 1156 (C-O-C, stretching of ether); ¹H-NMR: δ (ppm) 8.20 (s, 1H, O-H), 7.30 (dd, $J = 8.4, 1.6$ Hz, 1H, H-7), 7.25 (d, $J = 2.4$ Hz, 1H, H-5), 7.19 (d, $J = 8.0$ Hz, 1H, H-8), 7.00 (brt, $J = 7.6$ Hz, 1H, H-5'), 6.87 (dd, $J = 1.2, 2.0$ Hz, 1H, H-2'), 6.64 (dd, $J = 1.2, 8.0$ Hz, 1H, H-4'), 6.45 (dd, $J = 8.0, 1.2$ Hz, 1H, H-6'), 4.28-4.31 (m, 4H, CH₂-2 and CH₂-3), 3.51 (q, $J = 7.0$ Hz, 2H, CH₂-1''), 0.95 (t, $J = 7.2$ Hz, 3H, CH₃-2''); EIMS: m/z 335 [M]⁺, 271 [M-SO₂]⁺, 256 [C₆H₃C₂H₄O₂NCH₂C₆H₄OH]⁺, 242 [C₆H₃C₂H₄O₂NSO₂C₂H₅]⁺, 228 [C₆H₃O₂NCH₂C₆H₄OH]⁺, 227 [C₆H₃C₂H₄O₂SO₂NCH₂]⁺, 199 [C₆H₃C₂H₄O₂SO₂]⁺, 135 [C₆H₃C₂H₄O₂]⁺, 107 [C₆H₃O₂]⁺, 93 [C₆H₄OH]⁺, 76 [C₆H₄]⁺, 75 [C₆H₃]⁺, 50 [C₄H₂]⁺. (Calcd. for C₁₆H₁₇NO₅S; 335.3931).

N-Benzyl-N-ethyl-2,3-dihydrobenzo[1,4]dioxine-6-sulfonamide (7d)

IR (KBr, cm^{-1}): ν_{max} : 3417 (N-H, stretching), 3034 (C-H, stretching of aromatic ring), 2924 ($-\text{CH}_2-$, stretching), 1615 (C=C, stretching of aromatic ring), 1326 ($-\text{SO}_2-$, stretching), 1128 (C-O, stretching of ether); $^1\text{H-NMR}$: δ (ppm) 7.42 (dd, $J = 2.1, 8.4$ Hz, 1H, H-7), 7.34 (d, $J = 2.4$ Hz, 1H, H-5), 7.28-7.32 (m, 5H, H-2' to H-6'), 7.01 (d, $J = 8.4$ Hz, 1H, H-8), 4.23-4.30 (m, 4H, CH_2 -2 and CH_2 -3), 4.19 (s, 2H, CH_2 -7'), 3.15 (q, $J = 7.2$ Hz, 2H, CH_2 -1"), 0.88 (t, $J = 7.2$ Hz, 3H, CH_3 -2"); EIMS: m/z 333 $[\text{M}]^+$, 269 $[\text{M-SO}_2]^+$, 256 $[\text{C}_6\text{H}_3\text{C}_2\text{H}_4\text{O}_2\text{SO}_2\text{NCH}_2\text{C}_2\text{H}_5]^+$, 241 $[\text{C}_6\text{H}_3\text{C}_2\text{H}_4\text{O}_2\text{SO}_2\text{N}(\text{CH}_2)_2]^+$, 199 $[\text{C}_6\text{H}_3\text{C}_2\text{H}_4\text{O}_2\text{SO}_2]^+$, 135 $[\text{C}_6\text{H}_3\text{C}_2\text{H}_4\text{O}_2]^+$, 107 $[\text{C}_6\text{H}_3\text{O}_2]^+$, 77 $[\text{C}_6\text{H}_5]^+$, 75 $[\text{C}_6\text{H}_3]^+$, 51 $[\text{C}_4\text{H}_3]^+$. (Calcd. for $\text{C}_{17}\text{H}_{19}\text{NO}_4\text{S}$; 333.4234)

N-Ethyl-N-(2-phenylethyl)-2,3-dihydrobenzo[1,4]dioxine-6-sulfonamide (7e)

IR (KBr, cm^{-1}): ν_{max} : 3429 (N-H, stretching), 3027 (C-H, stretching of aromatic ring), 2921 ($-\text{CH}_2-$, stretching), 1619 (C=C, stretching of aromatic ring), 1328 ($-\text{SO}_2-$, stretching), 1145 (C-O, stretching of ether); $^1\text{H-NMR}$: δ (ppm) 7.27 (dd, $J = 8.2, 2.4$ Hz, 1H, H-7), 7.19 (d, $J = 2.4$ Hz, 1H, H-5), 6.99-7.25 (m, 5H, H-2' to H-6'), 6.97 (d, $J = 8.0$ Hz, 1H, H-8), 4.27-4.30 (m, 4H, CH_2 -2 and CH_2 -3), 3.30 (t, $J = 7.2$ Hz, 2H, CH_2 -8'), 3.20 (q, $J = 7.2$ Hz, 2H, CH_2 -1"), 2.81 (t, $J = 7.2$ Hz, 2H, CH_2 -7'), 1.07 (t, $J = 7.2$ Hz, 3H, CH_3 -2"); EIMS: m/z 347 $[\text{M}]^+$, 283 $[\text{M-SO}_2]^+$, 256 $[\text{C}_6\text{H}_3\text{C}_2\text{H}_4\text{O}_2\text{SO}_2\text{NCH}_2\text{C}_2\text{H}_5]^+$, 255 $[\text{C}_6\text{H}_3\text{O}_2\text{N}(\text{CH}_2)_2\text{C}_2\text{H}_5\text{C}_6\text{H}_5]^+$, 241 $[\text{C}_6\text{H}_3\text{C}_2\text{H}_4\text{O}_2\text{SO}_2\text{N}(\text{CH}_2)_2]^+$, 199 $[\text{C}_6\text{H}_3\text{C}_2\text{H}_4\text{O}_2\text{SO}_2]^+$, 135 $[\text{C}_6\text{H}_3\text{C}_2\text{H}_4\text{O}_2]^+$, 107 $[\text{C}_6\text{H}_3\text{O}_2]^+$, 91 $[\text{C}_7\text{H}_7]^+$, 75 $[\text{C}_6\text{H}_3]^+$, 65 $[\text{C}_5\text{H}_5]^+$. (Calcd. for $\text{C}_{18}\text{H}_{21}\text{NO}_4\text{S}$; 347.4587).

Enzyme inhibition studies***Butyrylcholinesterase and Acetylcholinesterase assay***

The BChE and AChE inhibition activity were carried out by taking 100 μL of the reaction mixture containing 60 μL of 50 mM Na_2HPO_4 buffer bearing pH 7.7, 10 μL of both the test compound and AChE/BChE having strength of 0.5

mM were added in each cell. After mixing the contents were pre-read at 405 nm and incubated at 37 °C for few mins. After incubation the 10 μL of substrate was planted; butyrylthiocholine chloride for BChE and acetylthiocholine iodide for AChE along with 10 μL DTNB (0.5 mM well^{-1}). After 15 min of incubation at 37 °C. Using Eserine as control the absorbance was computed at 405 nm [28]. The % inhibition was found by

$$\text{Inhibition (\%)} = \frac{\text{Control} - \text{Test}}{\text{Control}} \times 100$$

Where Control = Total enzyme activity without inhibitor

Test = Activity in the presence of test compound

Lipoxygenase assay

Lipoxygenase (LOX) activity was assayed according to the method [29-31] with small modifications. Baicalein (0.5 mM well^{-1}) was used as a positive control. IC_{50} values were calculated using the same procedure as mentioned for butyrylcholinesterase and acetylcholinesterase enzyme.

Statistical analysis

All the measurements were completed in triplicate and statistical analysis was performed by Microsoft Excel 2010. Results are offered as mean \pm sem.

Antimicrobial activity***Microbial strains***

All the synthesized samples were individually tested against a set of microorganisms, including two gram-positive bacteria: *Staphylococcus aureus* (*S. aureus*), and *Bacillus subtilis* (*B. subtilis*); two gram-negative bacteria: *Escherichia coli* (*E. coli*) and *Pasteurella multocida* (*P. multocida*) (local isolate) and four pathogenic fungi, *Candida albicans* (*C. albicans*), *Microsporium canis* (*M. canis*), *Aspergillus flavus* (*A. flavus*) and *Fusarium solani* (*F. solani*). The pure bacterial and fungal strains were obtained from Department of Clinical Medicine and

Surgery, University of Agriculture, Faisalabad, Pakistan. Purity and identity were verified by the Department of Microbiology, University of Agriculture, Faisalabad, Pakistan. In agar nutrient the bacterial and fungal strains were cultured at 37 °C and 28 °C, respectively. The incubation lasts for a day [32].

Disc diffusion method

The antimicrobial activity of the synthesized compounds was determined by disc diffusion method [32]. Amoxicillin (30 µg/dish) (Oxoid, UK) and Flumequine (30 µg/disk) (Oxoid, UK) were used as positive reference for bacteria and fungi, respectively to compare sensitivity of strain/isolate in analyzed microbial species. Plates, after 2 hr at 4 °C, were incubated at 37 °C for 18 hr for bacteria and at 28 °C for 24 hr for fungal strains. Antimicrobial activity was evaluated by measuring the diameter of the growth inhibition zones (zone reader) in millimeters for the organisms and comparing to the controls.

Hemolytic activity

Hemolytic activity of the compound was studied by taking 3 mL of freshly obtained heparinized human blood was collected from volunteers after consent, counseling and bovine from the Department of Clinical Medicine and Surgery, University of Agriculture, Faisalabad, Pakistan. Blood was centrifuged for 5 min and cells were washed three times with 5 mL of chilled (4 °C) sterile isotonic phosphate-buffered saline (PBS) with pH 7.4. Erythrocytes were maintained 10⁸ cells per mL for each assay. 100 µL of each compound was mixed with human (10⁸ cells/mL) separately. Samples were incubated for 35 min at 37 °C and agitated after 10 min. Immediately after incubation the samples were placed on ice for 5 min then centrifuged for 5 min. Supernatant 100 µL were taken from each tube and diluted 10 times with chilled (4 °C) PBS. Triton X-100 (0.1% v/v) was taken as positive control and PBS was taken as negative control and pass through the same process. The absorbance was noted at 576 nm using µQuant (Bioteck, USA). The % RBCs lysis for each sample was calculated [33].

Molecular docking

The structures of all the synthesized inhibitors were constructed using MOE-Builder tool. The default parameters of MOE-Dock program were used for the molecular docking of the ligands. Ligands were allowed to be flexible in order to find the accurate conformations of the ligands and to obtain minimum energy structures. At the end of docking, the best conformations of the ligands were analyzed for their binding interactions.

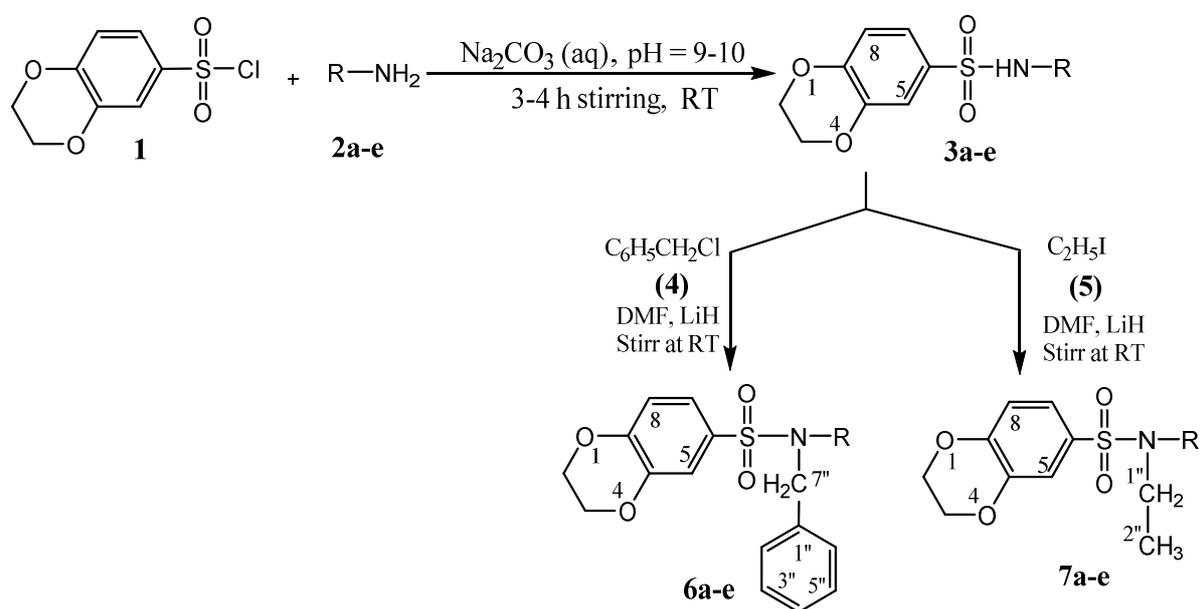
Results and Discussion

Chemistry

In the undertaken research, a series of heterocyclic compounds containing benzodioxane nucleus were synthesized as scheme 1. The parent compounds *N*-aryl-2,3-dihydrobenzo [1,4]dioxine-6-sulfonamide (3a-e), were prepared by a process similar to the known literature procedure [34] using 1,4-benzodioxane-6-sulfonyl chloride (1) and aryl amines (2a-e). Reactions of 3a-e with different electrophiles yielded a series of *N*-benzyl/ethyl-*N*-aryl-2,3-dihydrobenzo[1,4]dioxine-6-sulfonamide (6a-e and 7a-e) as represented in Scheme 1. Synthesis of all derivatives 6a-e and 7a-e was performed in DMF (*N,N'*-dimethylformamide) using lithium hydride (LiH) as the base. Complete conversion was achieved within 30 to 70 min by stirring. The products were isolated by adding cold distilled water in the reaction mixture and filtering off the precipitated solid. In some cases, compound was taken out through solvent extraction method by chloroform. Parent compound 3a was synthesized as light grey powder with yield of 92 % and melting point of 152-154 °C. HR-MS showing molecular ion peak at *m/z* 319.3772 confirming the formula C₁₆H₁₇NO₄S of a compound and total proton count was corroborated from ¹H-NMR spectrum. The FT-IR spectrum showed stretching frequencies at 3421, 3031, 1621, 1321 and 1115 cm⁻¹ provided the clues for the presence of N-H of sulfonamide, C-H and C=C of aromatic ring, -SO₂ of sulfonyl group and C-O-C of ether functionalities respectively. The EI-MS gave characteristic peaks at *m/z* 199 and 90 which were attributed to the formation of C₆H₃C₂H₄O₂SO₂⁺ and C₆H₃CH₃⁺

cations respectively. In the aromatic region of the $^1\text{H-NMR}$ spectrum signals appeared at δ 7.30 (d, J = 2.0 Hz, 1H, H-5), 7.21 (dd, J = 8.4, 2.4 Hz, 1H, H-7) and 6.85 (d, J = 8.4 Hz, 1H, H-8) were assigned to the phenyl ring attached to sulfonyl group; whereas three aromatic signals at δ 6.71 (s, 2H, H-2' and H-6'), 6.66 (s, 1H, H-4') were assigned to the benzene ring of 3,5-dimethylphenyl group. The signals appeared at 4.23-4.25 (m, 4H, CH_2 -2 and CH_2 -3), and 2.21 (s, 6H, CH_3 -1'' and CH_3 -2'') indicated the presence of 1,4-dioxane nucleus and two methyl groups attached to third

and fifth position of aniline in the molecule contributed to the aliphatic region of the spectrum. On these backgrounds, the structure was assigned to the parent compound *N*-(3,5-Dimethylphenyl)-2,3-dihydrobenzo[1,4]dioxine-6-sulfonamide 3a. Similarly, the structure of other compounds was characterized on above said spectral techniques. The physical data is provided in Table 1. The mass fragmentation pattern of *N*-(2-Phenylethyl)-2,3-dihydrobenzo[1,4]dioxine-6-sulfonamide (3e) is given in Fig. 1.

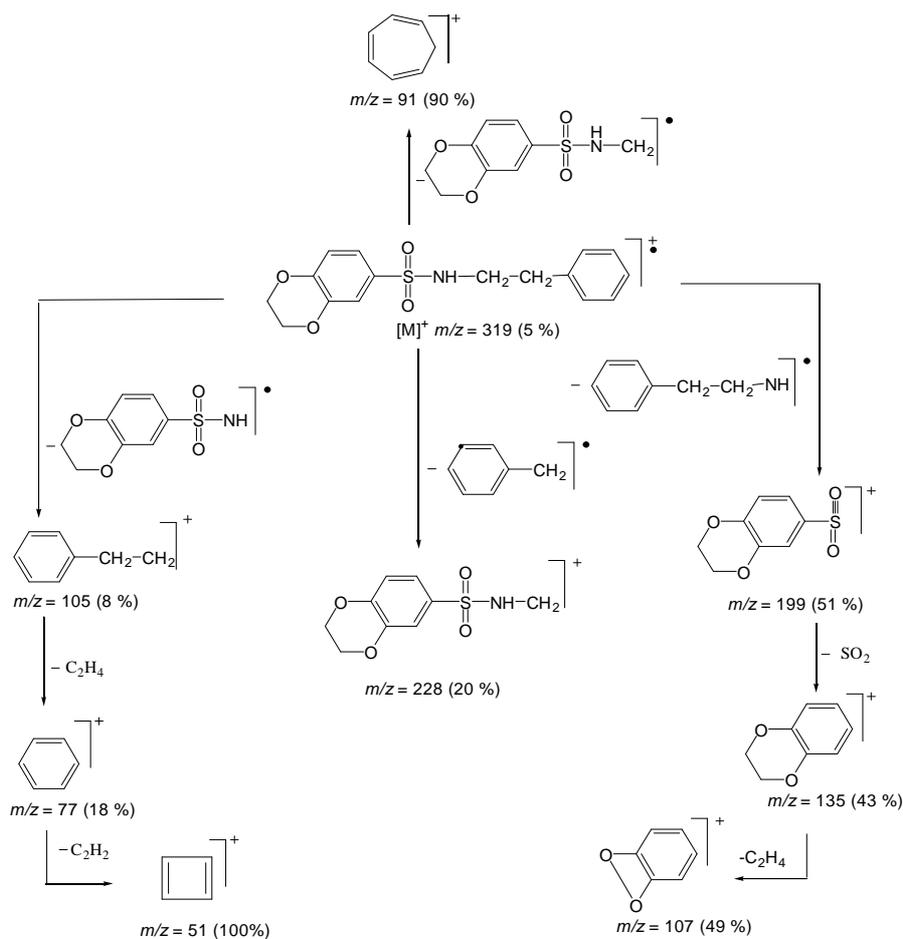


Compound	R	Compound	R
2a,3a,6a,7a		2d,3d,6d,7d	
2b,3b,6b,7b		2e,3e,6e,7e	
2c,3c,6c,7c			

Scheme 1: Synthetic scheme of various sulfonamides bearing benzodioxane nucleus

Table I. Physical data of the synthesized compounds.

Compound	Physical state	Color	Mol. formula	Mol. Wt.	M.P. °C	% yield
3a	Solid	Light grey	C ₁₆ H ₁₇ NO ₄ S	319	152-154	92
3b	Solid	White	C ₁₅ H ₁₅ NO ₄ S	305	119-121	87
3c	Solid	Light grey	C ₁₄ H ₁₃ NO ₅ S	307	112-114	75
3d	Solid	White	C ₁₅ H ₁₅ NO ₄ S	305	78-80	86
3e	Solid	White	C ₁₆ H ₁₇ NO ₄ S	319	91-93	94
6a	Solid	White	C ₂₃ H ₂₃ NO ₄ S	409	241-243	89
6b	Sticky solid	Dark brown	C ₂₂ H ₂₁ NO ₄ S	395	-	79
6c	Gummy solid	Yellowish brown	C ₂₁ H ₁₉ NO ₅ S	397	-	71
6d	Solid	Yellow	C ₂₂ H ₂₁ NO ₄ S	395	-	70
6e	Solid	White	C ₂₃ H ₂₃ NO ₄ S	409	152-154	92
7a	Solid	Off white	C ₁₈ H ₂₁ NO ₄ S	347	260-262	93
7b	Sticky solid	Brown	C ₁₇ H ₁₉ NO ₄ S	333	-	74
7c	Sticky solid	Dark brown	C ₁₆ H ₁₇ NO ₅ S	335	-	71
7d	Sticky solid	Yellowish brown	C ₁₇ H ₁₉ NO ₄ S	333	-	73
7e	Solid	White	C ₁₈ H ₂₁ NO ₄ S	347	80-82	89

Figure 1. Mass fragmentation pattern of *N*-(2-phenylethyl)-2,3-dihydrobenzo[1,4]dioxine-6-sulfonamide (3e)

Enzyme inhibition

The screening of all the synthesized compounds against butyrylcholinesterase enzyme revealed that only three compounds showed good activity, *N*-benzyl-*N*-(3-hydroxyphenyl)-2,3-dihydrobenzo[1,4]dioxine-6-sulfonamide (6c), *N*-benzyl-2,3-dihydrobenzo[1,4]dioxine-6-sulfonamide (3d), *N*-benzyl-*N*-(3,5-dimethylphenyl)-2,3-dihydrobenzo [1,4] dioxine-6-sulfonamide (6a) having IC₅₀ values of 71.27±0.01, 198.21±0.11 and 259.61±0.01 μmoles/L respectively, comparative to eserine (Table 2). The activity of the these compound was the most probably due to presence of hydroxyl group at third position of aniline ring in 6c, benzyl group in 3d and two alkyl groups at third and fifth position of aniline ring in 6a along with benzyl group attached to nitrogen of sulfonamide. For screening, against acetylcholinesterase enzyme, among all the synthesized compounds only three demonstrated better activity i.e. *N*-benzyl-*N*-(3-hydroxyphenyl)-2,3-dihydrobenzo[1,4]dioxine-6-sulfonamide (6c), *N*-(3-hydroxyphenyl)-2,3-dihydrobenzo[1,4]dioxine-6-sulfonamide (3c) and *N*-ethyl-*N*-(3,5-dimethylphenyl)-2,3-dihydrobenzo[1,4]dioxine-6-sulfonamide (7a) having IC₅₀ values of 99.93±0.19, 174.51±0.15, and 189.81±0.08 μmoles/L, respectively compare-

ative to standard. The proficient activity of first and second compound was the most likely due to occurrence of hydroxyl group at meta position of aniline ring and that of third compound was likely due to the presence of two alkyl groups on aniline ring along with ethyl group attached to nitrogen of sulfonamide. Against lipoxygenase enzyme, all the synthesized compounds showed beneficially good activity but the most active were *N*-(3,5-dimethylphenyl)-2,3-dihydrobenzo[1,4] dioxine-6-sulfonamide (3a), *N*-(3,5-dimethylphenyl)-*N*-ethyl-2,3-dihydrobenzo[1,4]dioxine-6-sulfonamide (7a) and *N,N*-dibenzyl-2,3-dihydrobenzo[1,4]dioxine-6-sulfonamide (6d) having IC₅₀ values of 91.25±0.16, 65.25±0.16 and 81.91±0.21, μmoles/L respectively, comparative to baicalein. The presented activity of first compound was the most probably due to occurrence of two alkyl groups, at third and fifth positions of aniline ring, that of second and third compounds was credibly due to the presence of two alkyl groups at 3rd/5th of position of aniline ring and benzyl group, along with benzyl and ethyl group respectively attached to nitrogen of sulfonamide. All the parent compounds (3a-e) can be further utilized for the synthesis of new derivatives with other different electrophiles to enhance their biological, antimicrobial and other activities.

Table 2. Bioactivity study of synthesized molecules.

C. No.	BChE			AChE			LOX		
	Conc./well (mM)	Inhibition (%)	IC ₅₀ (μmoles/L)	Conc. (mM)	Inhibition (%)	IC ₅₀ (μmoles/L)	Conc./well (mM)	Inhibition (%)	IC ₅₀ (μmoles/L)
3a	0.5	5.65±0.35	-	0.5	55.98±0.15	<400	0.5	79.17±0.63	91.25±0.16
3b	0.5	56.79±0.48	<400	0.5	18.52±0.77	-	0.5	53.51±0.11	<400
3c	0.5	11.89±0.14	-	0.5	72.31±0.11	174.51±0.15	0.5	65.53±0.28	209.81±0.13
3d	0.5	67.58±0.14	198.21±0.11	0.5	15.33±0.57	Nil	0.5	51.99±0.51	<400
3e	0.25	57.91±0.87	<400	0.5	18.47±0.87	-	0.5	70.29±0.91	104.11±0.17
6a	0.5	63.56±0.36	259.61±0.01	0.5	52.66±0.69	<400	0.5	39.87±0.16	-
6b	0.5	50.24±0.46	<500	0.5	13.33±0.22	>500	0.5	34.64±0.51	-
6c	0.5	79.12±0.13	71.27±0.01	0.5	87.79±0.11	99.93±0.19	0.5	12.64±0.11	-
6d	0.5	44.05±0.11	-	0.5	37.93±0.87	-	0.5	85.61±0.19	81.91±0.21
6e	0.5	30.81±0.11	-	0.5	10.26±0.92	-	0.5	30.39±0.24	-
7a	0.5	53.72±0.15	<400	0.5	70.14±0.18	189.81±0.08	0.5	88.64±0.14	65.25±0.16
7b	0.5	50.99±0.63	<500	0.5	11.67±0.87	>500	0.5	20.01±0.17	-
7c	0.5	28.28±0.62	-	0.5	16.91±0.85	-	0.5	45.74±0.61	-
7d	0.5	43.29±0.19	-	0.5	36.61±0.69	-	0.5	72.44±0.18	101.51±0.31
7e	0.5	36.39±0.13	>500	0.5	75.19±0.18	301.2±0.05	0.5	27.38±0.34	-
Control	Eserine	82.82±1.09	0.85±0.0001	Eserine	91.29±1.17	0.04±0.0001	Baicalein	93.79±1.27	22.4±1.3

Note: IC₅₀ values (concentration at which there is 50 % enzyme inhibition) of compounds were calculated using EZ-Fit Enzyme kinetics software (Perella Scientific Inc. Amherst, USA).

LOX = Lipoxygenase. AChE = Acetyl cholinesterase. BChE = Butyryl cholinesterase.

Antimicrobial activity

The *in vitro* antimicrobial properties of the parent compounds and their derivatives were tested (Table 3). Among the parent compounds 3a showed the antimicrobial activity against the selected panel of both bacterial and fungal species. Regarding the derivatives (6a-e) series; 6a showed higher activities in comparison to the rest of the members of its series. 7a and 7b were the members of 7a-e series which were the moderate to good active against both bacterial and fungal strains and 7c was relatively efficient antifungal candidate against selected strains. The remaining compounds possess very low or no activity against the assessed

microorganisms. The highest hemolytic activity was shown by 6a (92 %) but lower than the positive control (Triton-X-100). The lowest activity was shown by 6b and 7c (1.2 % and 1.6 % respectively) but higher than the negative controls PBS. These synthesized molecules inhibit the synthesis of cell wall protein of bacteria; interfering their growth and also break down red blood cells of host. Those antibacterial candidates would be selected; that have low hemolytic activity and high antibacterial or antifungal potential. On the basis of the previous results we may assume that the synthesized sulfonamides may be suitable leads for further improvement to address different targets.

Table 3. Antibacterial and antifungal studies on synthesized compounds.

Compound	Antibacterial activity					Antifungal activity			
	<i>Staphylococcus aureus</i>	<i>Bacillus subtilis</i>	<i>Pasturella multocida</i>	<i>Escherichia coli</i>	Hemolytic activity (Mean) % \pm S.D	<i>Candida albicans</i>	<i>Microspor umcanis</i>	<i>Aspergillus flavus</i>	<i>Fusarium solani</i>
	Zone of inhibition (mm)					Zone of inhibition (mm)			
3a	-	-	-	-	86.844 \pm 0.417	-	-	-	-
3b	14	12	14	14	86.765 \pm 0.212	17	12	14	18
3c	-	-	-	-	90.164 \pm 0.278	-	-	-	-
3d	-	-	-	-	74.426 \pm 0.031	-	-	-	-
3e	-	-	-	-	88.901 \pm 0.025	-	-	-	-
6a	18	14	16	16	92.973 \pm 7.063	19	14	15	14
6b	-	-	-	-	1.202 \pm 0.155	-	-	-	-
6c	-	-	-	-	87.720 \pm 0.284	-	-	-	-
6d	-	-	-	-	87.345 \pm 0.246	-	-	-	-
6e	-	-	-	-	91.956 \pm 1.425	-	-	-	-
7a	16	18	16	16	92.723 \pm 8.405	16	18	16	16
7b	19	22	20	18	58.557 \pm 2.983	11	13	10	14
7c	-	-	-	-	1.639 \pm 0.093	12	10	15	12
7d	-	-	-	-	86.678 \pm 0.250	-	-	-	-
7e	-	-	-	-	87.565 \pm 1.425	-	-	-	-
Streptomycin	30	28	28	30	Flumequine	29	27	26	31
PBS					0.00 \pm 0.0				
Triton (toxicity)					100 \pm 0.0				

Molecular docking

The results obtained from *in silico* approach were also favoring the fact that synthesized sulfonamides have shown good interaction with the target site. The interaction analysis shown that in every compound, the sulfonamide group is contributing in the interactions. The interaction of compound 3b with the active site of LOX and AChE is shown in Fig. 2 and Fig. 3. As depicted from stature; compound 3b showed good interaction with the amino acids residues of binding cavity. Against LOX, the both oxygen of sulfonyl group showed interactions with the Histidine 528 residue; whereas one oxygen of dioxane ring displayed attachment with Histidine 523 of LOX. In an AChE display; one doubly bonded oxygen of sulfonyl group exhibited hydrogen bonding with the serine 122 residue; while second one demonstrated attachment with Tyrosine 121 and phenylene group of benzodi-oxane displayed bonding to phenylalanine 130 amino acid. Thus molecular docking results were well in correlation with the experimentally determined data of enzyme inhibition against AChE and LOX.

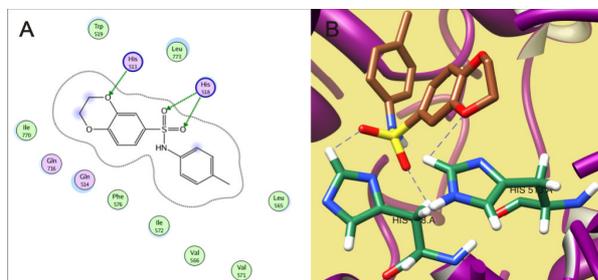


Figure 2. The interaction between compound 3b and lipoxygenase. 2D and 3D pose of the complex is shown in figure A and B respectively

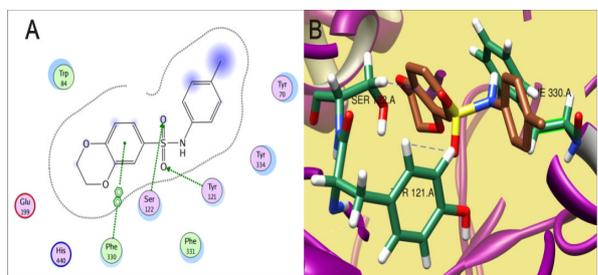


Figure 3. The interaction between compound 3b and acetylcholinesterase. 2D and 3D pose of the complex is shown in figure A and B respectively

Conclusion

The new series of sulfonamide bearing 1,4-benzodioxane ring were reported. The targets were characterized by FT-IR, ¹H-NMR and EIMS. All the compounds were screened for their antibacterial and antifungal activity by disc diffusion method. Compounds 3a, 6a and 7b exhibited good antimicrobial activity amongst all in comparison to standard (streptomycin). Compound 6c was more active against both BchE and AchE, while 7a exhibited good inhibition potential against LOX.

Acknowledgements

The Authors extend their appreciation to the Higher Education Commission of Pakistan for financial support.

References

1. G. L. Mandell and M. A. Sande, *Pergamon Press, New York*, (1990) 1047.
<https://books.google.com.pk/books/ISBN=0080527523>
2. S. Wang, H. Y. Zhang, L. Wang, Z. J. Duan and I. Kennedy, *Food Addit. Contam.*, 23 (2006) 362.
[doi:10.1080/02652030500499359](https://doi.org/10.1080/02652030500499359)
3. A. K. Sarmah, M. T. Meyer and A. B. A. Boxall, *Chemosphere*, 65 (2006) 725.
[doi:http://dx.doi.org/10.1016/j](https://doi.org/10.1016/j.chemosphere.2010.07.031)
4. X. L. Wang, K. Wan and C. H. Zhou, *Eur. J. Med. Chem.*, 45 (2010) 4631.
[doi: 10.1016/j.ejmech.2010.07.031](https://doi.org/10.1016/j.ejmech.2010.07.031)
5. Epstein, M. Amodio-Groton and N. S. Sadick, *J. Am. Acad. Dermatol.*, 37 (1997) 149.
<https://www.ncbi.nlm.nih.gov/pubmed/9270499>
6. J. D. Smilack, *Mayo Clin. Proc.*, 74 (1999) 730
<https://doi.org/10.4065/74.7.730>
7. M. E. Gadad, C. S. Mahajanshetti, S. Nimbalkar and A. Raichurkar, *Eur. J. Med. Chem.*, 35 (2000) 853.
[https://doi.org/10.1016/S0223-5234\(00\)00166-5](https://doi.org/10.1016/S0223-5234(00)00166-5)

8. B. L. Wilkinson, L. F. Bornaghi, T. A. Houston, A. Innocenti, C. Vullo, C. T. Supuran and S. A. Poulsen, *J. Med. Chem.*, 50 (2007) 1651.
doi: [10.1021/jm061320h](https://doi.org/10.1021/jm061320h)
9. C. Almansa, J. Bartroli, J. Belloc, F. L. Cavalcanti, R. Ferrando, L. A. Gomez, I. Ramis, E. Carceller, M. Merlos and J. J. Garcia-Rafanell, *Med. Chem.*, 47 (2004) 5579.
doi: [10.1021/jm040844j](https://doi.org/10.1021/jm040844j)
10. D. Vullo, De Luca, V. Scozzafava, A. Carginale, V. Rossi, M. Supuran, C. T. Capasso, *Bioorg. Med. Chem.*, 21 (2013) 4521.
doi: [10.1016/j.bmc.2013.05.042](https://doi.org/10.1016/j.bmc.2013.05.042)
11. W. Chu, J. Rothfuss, A. Avignon, C. Zeng, D. Zhou, R. S. Hotchkiss and R.H. Mach, *J. Med. Chem.*, 50 (2007) 3751.
doi: [10.1021/jm070506t](https://doi.org/10.1021/jm070506t)
12. El-Dien, G.G. Mohamed, E. Khaled and E.Y. Z. Frag, *J. Adv. Res.*, 1 (2010) 215.
doi: [10.1016/j.jare.2010.05.005](https://doi.org/10.1016/j.jare.2010.05.005)
13. W. Chu, J. Zhang, C. Zeng, J. Rothfuss, Z. Tu, Y. Chu, D. E. Reichert, M. J. Welch and R. H. Mach, *J. Med. Chem.*, 48 (2005) 7637.
<https://pubs.acs.org/doi/abs/10.1021/jm0506625>
14. D. J. Abraham, S. Takagi, R. D. Rosenstein, R. Shiono, H. Wagner, L. Horhammer, O. Seligmann and N. R. Farnsworth, *Tetrahedron Lett*, 31 (1970) 2675.
pubs.rsc.org/en/content/articlepdf/2003/ob/b300099k
15. W. Gu, X. Chen, X. Pan, A. S. C. Chan and Y. Teng-Kuei, *Tetrahedron: Asymmetry*, 11 (2000) 2801.
doi: [10.1021/ol034415b](https://doi.org/10.1021/ol034415b)
16. W. Chu, J. Zhang, C. Zeng, J. Rothfuss, Z. Tu, Y. Chu, D. E. Reichert, M. J. Welch and R. H. Mach, *J. Med. Chem.*, 48 (2005) 7637.
doi: [10.1021/jm0506625](https://doi.org/10.1021/jm0506625)
17. B. Kenny, S. Ballard, J. Blagg and D. Fox, *J. Med. Chem.*, 40 (1997) 1293.
doi: [10.1021/jm960697s](https://doi.org/10.1021/jm960697s)
18. R. R. Ruffolo, W. Bondinell and J. P. Hieble, *J. Med. Chem.*, 38 (1995) 3681.
<http://hyper.ahajournals.org/content/hypertensionaha/33/2/708.full.pdf>
19. A. Leonardi, R. Testa, P. G. De Benedetti, P. Hieble and D. Giardin, Elsevier, Amsterdam (1996) 135.
20. B. Ahmed, S. A. Khan and T. Alam, *Pharmazie*, 58 (2003) 173. PMID: 12685811,
<https://www.ingentaconnect.com/content/govi/pharmaz/2003/00000058/00000003/art00003>
21. H. Khalilullah, S. Khan, M.J. Ahsan and B. Ahmed, *Korean Chem. Soc. Bull.*, 33 (2012) 575.
doi: [10.5012/bkcs.2012.33.2.575](https://doi.org/10.5012/bkcs.2012.33.2.575)
22. S. A. Khan, B. Ahmad and T. Alam, *Pak. J. Pharm. Sci.*, 19 (2006) 290. PMID: 17105706
23. K. Nikolic, D. Agbaba, *Hem. Ind.*, 66 (2012) 619.
doi: [10.2298/HEMIND120221037N](https://doi.org/10.2298/HEMIND120221037N)
24. M. T. Vazquez, G. Rosell and M. D. Pujol, *Pharmaco*, 51 (1996) 215. PMID: 8688144
25. L. I. Pilkington and D. Barker, *Nat. Prod. Rep.*, 32 (2015) 1369.
doi: [10.1039/c5np00048c](https://doi.org/10.1039/c5np00048c)
26. A. Wadood, M. Riaz, S. B. Jamal, M. Shah and M. A. Lodhi, *Bioinformation*, 9 (2013) 309.
doi: [10.6026/97320630009309](https://doi.org/10.6026/97320630009309)
27. M. Irshad, M. A. Abbasi, Aziz-ur-Rehman, S. Z. Siddiqui, M. S. Ali, M. Ashraf, T. Ismail, I. Ahmad, S. Hassan, M. A. Lodhi and S. B. Jamal, *Pak. J. Pharm. Sci.*, 29 (2016) 1913.
<http://www.pjps.pk/wp-content/uploads/pdfs/29/6/Paper-4.pdf>
28. G. L. Ellman, K. D. Courtney, V. Andres and R. M. Featherstone, *Biochem. Pharmacol.*, 7 (1961) 88.
[https://doi.org/10.1016/0006-2952\(61\)90145-9](https://doi.org/10.1016/0006-2952(61)90145-9)
29. A. L. Tappel, *Arch. Biochem. Biophys.*, 54 (1955) 266.
[https://doi.org/10.1016/0003-9861\(55\)90039-4](https://doi.org/10.1016/0003-9861(55)90039-4)
30. H. C. Clapp, A. Banerjee and S. A. Rotenberg, *J. Biochem.*, 24 (1985) 1826.
doi: [org/10.1021/bi00329a004](https://doi.org/10.1021/bi00329a004)
31. C. Kemal, P. Louis-Flemborg, R. R. Krupinski-Olsen and A. L. Shorter, *J. Biochem.*, 26 (1987) 7064.
doi: [org/10.1021/bi00396a031](https://doi.org/10.1021/bi00396a031)

32. CLSI (The clinical Laboratory Standard Institute) *J. Clin. Microbiol*, 45 (2007) 2752. doi: [10.1128/JCM.00143-07](https://doi.org/10.1128/JCM.00143-07)
33. P. Sharma and J. D. Sharma, *J. Ethol*, 74 (2001) 239. Pubmed/11274824, [https://doi.org/10.1016/S0378-8741\(00\)00370-6](https://doi.org/10.1016/S0378-8741(00)00370-6)
34. X. Deng and N. S. Mani, *Asian J. Pharm. Hea. Sci.*, 8 (2006) 835. doi: [10.1039/b606127c](https://doi.org/10.1039/b606127c).